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Attorney Docket No. SALK1510-3

____ NEW PATENT APPLICATION _X_ CONTINUATION-IN-PART

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Sir:

Transmitted herewith for filing is the <u>new</u> patent application of

Inventors: Ronald M. Evans, J. Don Chen and Peter Ordentlich

FOI: A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR

This is a request for filing a continuation-in-part under 35 U.S.C. 111(A) and 37 C.F.R. 1.53(b), of U.S. Application Serial No. 08/522,726, filed September 1, 1995, now pending.

Enclosed are:

- _X 75 pages of the Specification, which includes _7 pages of the claims and _1 page of the Abstract;
- X 12 sheets of drawing(s) Formal; X Informal;
- X A Declaration (unexecuted);
- X 67-Page Sequence Listing;
- X computer readable disk containing Sequence Listing; and
- X Statement Under 37 C.F.R. §§1.821(f) and (g).

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In re Application of: Evans et al.

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PATENT Attorney Docket No.: SALK1510-3

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Multiple Dependent Claims Presented:	Yes X N	0		3	\$130	\$260				\$.00
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Respectfully submitted,

Date: March 10, 2000

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APPLICATION

For

UNITED STATES LETTERS PATENT

on

A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR

by

Ronald M. Evans, J. Don Chen and

Peter Ordentlich

Sheets of Drawings: Twelve (12) Docket No.: SALK 1510-3

Attorneys

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A Family of Transcriptional Co-repressors that Interact with Nuclear Hormone Receptors and Uses Therefor

5 RELATED APPLICATIONS

FIELD OF THE INVENTION

The present invention relates to intracellular receptors, methods for the modulation thereof, and methods for the identification of novel ligands therefor. In a particular aspect, the present invention relates to methods for the identification of compounds which function as ligands (or ligand precursors) for intracellular receptors. In another aspect, the present invention relates to novel chimeric constructs and uses therefor.

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BACKGROUND OF THE INVENTION

A central problem in eukaryotic molecular biology continues to be the elucidation of molecules and mechanisms that mediate specific gene regulation. As part of the scientific attack on this problem, a great deal of work has been done in efforts to identify ligands (i.e., exogenous inducers) which are capable of mediating specific gene regulation. Additional work has been done in efforts to identify other molecules involved in specific gene regulation.

Although much remains to be learned about the specifics of gene regulation, it is known that ligands modulate gene transcription by acting in concert with intracellular components, including intracellular receptors and discrete DNA sequences known as hormone response elements (HREs).

The identification of compounds that directly or indirectly interact with intracellular receptors, and thereby affect transcription of hormone-responsive genes, would be of significant value, e.g., for therapeutic applications.

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Transcriptional silencing mediated by nuclear receptors plays an important role in development, cell differentiation, and is directly linked to the oncogenic activity of v-erbA. The mechanism underlying this effect is unknown but is one key to understanding the molecular basis of hormone action. Accordingly, the identification of components involved in transcriptional silencing would represent a great advance in current understanding of mechanisms that mediate specific gene regulation.

Other information helpful in the understanding and practice of the present invention can be found in commonly assigned United States Patent Nos. 5,071,773, 4,981,784, 5,260,432, and 5,091,513, all of which are hereby incorporated herein by reference in their entirety.

BRIEF DESCRIPTION OF THE INVENTION

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The present invention overcomes many problems in the art by providing a family of receptor interacting co-repressors, referred to herein as "SMRT co-repressor", i.e., a silencing mediator (co-repressor) for retinoic acid receptor (RAR) and thyroid hormone receptor (TR). In vivo, members of the SMRT family of co-repressors function as potent co-repressors. A GAL4 DNA binding domain (DBD) fusion with a SMRT co-repressor behaves as a frank repressor of a GAL4-dependent reporter.

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Together, these observations identify a novel family of cofactors that is believed to represent an important mediator of hormone action.

Accordingly, the present invention provides isolated silencing mediators of retinoic acid and thyroid hormone receptors, and isoforms or peptide portions thereof (SMRT co-repressors), that modulate transcriptional potential of members of the nuclear receptor superfamily. Such SMRT co-repressors comprise a repression domain having less than about 83% identity with a Sin3A interaction domain of N-CoR (amino acids 255 to 312 of SEQ ID NO: 11); less than about 57% identity with repression domain 1 of N-CoR (amino acids 1 to 312 of SEQ ID NO: 11); less than about 66% identity with a SANT domain of N-CoR (amino acids 312 to 668 of SEQ ID NO: 11) and/or; less than about 30% identity with repression domain 2 of N-CoR (amino acids 736 to 1031 of SEQ ID NO: 11).

In accordance with yet another embodiment of the present invention, there are provided isolated peptides comprising at least a portion of the invention SMRT co-repressor six contiguous amino acids of an amino acid sequence selected from the group consisting of:

amino acids 1 to 1030 of SEQ ID NO: 5;
amino acids 1 to 1029 of SEQ ID NO: 7;
amino acids 1 to 809 of SEQ ID NO: 9;
and conservative variations thereof,
provided the peptide is not identical to a sequence of SEQ ID NO: 11.

In addition, there are provided isolated antibodies that bind specifically to invention isolated peptides. There are also provided chimeric molecules comprising invention isolated peptides and at least a second molecule. Also provided are complexes comprising an invention SMRT co-repressor and a member of the superfamily of nuclear receptors and isolated antibodies that bind to such complexes.

Accordingly, the present invention provides isolated polynucleotides encoding members of the newly described family of silencing mediators of retinoic acid and thyroid hormone receptor or an isoform or peptide portion thereof (SMRT co-repressor), or an isolated polynucleotide complementary thereto. In addition, there are provided vectors comprising invention polynucleotides, as well as host cells containing invention polynucleotides.

In additional embodiments of the present invention, there are provided methods for identifying agents that modulate the repressor potential of a SMRT corepressor.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates a function of an invention SMRT co-repressor.

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In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell.

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In another embodiment according to the present invention, there are provided methods of identifying a molecule that interacts specifically with a SMRT co-repressor.

BRIEF DESCRIPTION OF THE FIGURES

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Figure 1 shows the quantitation by phosphoimager of a dose-dependent dissociation of SMRT from RAR or TR by all-trans retinoic acid (atRA) or thyroid hormone (triiodothyronine or T3).

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Figure 2 presents amino acid (aa) sequences of SMRT (Genbank accession number XXXXX). The aa sequence presented in parentheses (i.e., residues

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1330-1376) is an alternatively spliced insert which is not present in the original two-hybrid clone (C-SMRT, aa 981 to C-terminal end). The proline-rich N-terminal domain (aa 1-160) and the glutamine-rich region (aa 1061-1132), as well as the ERDR and SG regions, are also indicated. The C-terminal region of SMRT (aa 1201 to C-terminal end) shows 48% aa identity to RIP13 (Seol et al., *Molecular Endocrinology* 9:72-85 (1995)). The rest of the sequence of RIP13 shows 22% aa identity to SMRT (aa 819-1200).

Figure 3 illustrates mediation of the silencing effect of hRAR α and hTR β by SMRT in vivo.

Figure 3(A) illustrates that v-erbA reverses the silencing effect of GAL-RAR (GAL4 DBD-hRAR\alpha 156-462) while SMRT restores the silencing effect.

Figure 3(B) illustrates that the RAR403 truncation mutant reverses the silencing effect of GAL-TR (GAL4 DBD-hTRβ 173-456) while SMRT restores the silencing effect.

 $Figure\ 3(C)\ illustrates\ that\ v-erbA\ and\ full\ length\ SMRT\ or\ C-SMRT$ $20 \quad have\ no\ effect\ on\ GAL-VP16\ activity.$

Figure 3(D) illustrates that a GAL4 DBD fusion of full length SMRT represses the thymidine kinase basal promoter activity containing four GAL4 binding sites. The fold of repression was calculated by dividing the normalized luciferase activity transfected with the GAL4 DBD alone by those transfected with indicated amount of GAL DBD fusion constructs.

Figure 4 provides an alignment of the human SMRT (SEQ ID NO: 5)
and mouse SMRTα (SEQ ID NO: 7) amino acid sequences. Proteins were aligned
30 using the CLUSTAL alignment program. Underlined sequence of mouse SMRTα corresponds to the amino acid sequences that are deleted in mouse SMRTβ. The

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arrow indicates the start point of the previously described human SMRT co-repressor (sSMRT).

Figures 5A and 5B provide alignments of the human SMRT and 5 human N-CoR co-repressors.

Figure 6A is a graph showing the results of transactivation experiments using transcripts encoding a detectable reporter and either wild type EcR (Ecr wt), a repression-Defective EcR allele Ecraa^{483T} (EcRA483T) or vp16 activation domain fused to Ultraspiracle (vp16-USP).

Figure 6B is a graph showing the results of transactivation experiments using CMV promoter-driven expression vectors. Wild-type EcR or EcR A483T was cotransfected with vp16-USP and Gal4-c-SMRT (aa 981 to C terminus) (Chen and Evans, *Nature* 377:454-457, (1995)) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element.

Figure 6C shows alignment of EcR, rTR, hRAR, and rRev-erbA receptor sequences and the secondary structure in the LBD signature motif region. Conserved residues are marked in dark. The mutation 483 (AT) is marked at the top of the corresponding residue.

Figure 7 is a graph showing β-galactosidase activity in a yeast two-hybrid screen with pAS-EcR as bait. pAS-EcR is a fusion gene with the region corresponding to aa 223-878 of EcRB1 fused C-terminally to the Gal4-DBD of the pAS1-CYH2 construct (Durfee et al., Genes Dev 7:555-569 (1993)); other Gal4-DBD-based nuclear receptor constructs used in this yeast two-hybrid assay include: USP
 (aa 50-508), hRAR (aa 186-462) and hTR (aa 121-410) (Schulman et al., Proc. Natl. Acad. Sci. USA, 92:8288-8292, (1995)), and SMRT (Chen and Evans, (1995), supra).

 β -galactosidase activities were quantified by liquid assay for yeast cells treated either without ligand or with 3 μ M of corresponding hormone. All-trans retinoic acid (ATRA) is a ligand of RAR; 3,3',5-triiodothyroacetic acid (T3) is a ligand of TR. RAR, retinoic acid receptor; TR, thyroid hormone receptor.

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Figure 8A shows the complete amino acid sequence of the SMRTER protein (SEQ ID NO: 12). The underlined regions represent the residues also conserved in SMRT and N-CoR. The gray box indicates the sequences of the E52 clone.

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Figure 8B is a schematic structural diagram of SMRTER, SMRT, and N-CoR showing the conserved SNOR, SANT, GST, ITS, D/ER repeat, and LSD motifs with their designated patterns positioned in their relative regions in each protein.

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Figure 9. Sequence Comparison of SMRTER, SMRT, N-CoR, and Other Related Proteins. The SANT domains of various proteins are listed. Percent identities/similarities compared to SMRTER are shown on the right. Two potential helices are predicted in the N-terminal half of the SANT domain. Black boxes indicate identical sequences; gray boxes, similar or partially identical sequences.

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Figure 10 is a schematic representation showing functional domains in SMRTER. Numbers on the left represent the regions in SMRTER used to generate the Gal4-DBD fusion genes. Black stippled bars indicate the locations of EcR-interacting domains; gray stippled bars indicate repression domains. Plus signs indicate that a positive interaction between SMRTER and the EcR complex and repression of basal activity by Gal4-SMRTER is significant. ERID = ecdysone receptor-interacting domain; SMRD = SMRTER repressor domain.

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Figure 11A is a graph showing the interaction of ERID1 AND ERID2 with the EcR complex. Figure 11B is a graph showing the results of competition

between ERID1, ERID2 and c-SMRT for binding to EcR. Figure 11C is a graph showing that EcR A483T disrupts the interaction with ERID1 and ERID2.

Figure 12A shows the results of mapping three repression domains. To

sexamine repressive activity, transcriptional activity of each Gal4-SMRTER fusion

was compared to the basal activity of Gal4-DBD on reporter. Only repression with

value approximately 5-fold or over is considered positive (+).

Figure 12B is a schematic representation of mapping the SMRTER-interacting domain in mSin3A and dSin3A. Yeast two-hybrid assays were used to assess the interaction between each Gal4-DBD-based fusion gene of each SMRD and the ACT-based fusion genes of mSin3A and dSin3A. The numbers indicate the region in either mSin3A or in dSin3A used to generate the ACT fusion genes. Constructs of mSin3A were described previously in Nagy et al., Cell 89:373-380, (1997).

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Figure 12C shows an alignment of SMRD3 of SMRTER and an mSin3-interacting domain of N-CoR. Conserved residues are boxed in gray. An asterisk indicates the region where the mutation (Gly) was generated. Minus signs indicate that the interaction between SMRD3 and Sin3A was not detectable in the yeast two-hybrid assays. Repression was measured by comparing the transcriptional activity of Gal4-SMRD3 M2 or Gal4-SMRD3 M3 to that of wild-type Gal4-SMRD3 using transfection experiments as described above.

DETAILED DESCRIPTION OF THE INVENTION

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In accordance with the present invention, there is provided a family of isolated SMRT co-repressors, and isoforms and peptide portions thereof, that modulate transcriptional potential of members of the nuclear receptor superfamily. Exemplary members of this family are co-repressors having substantially the same sequence as residues 1-1329 plus 1376-1495, as set forth in SEQ ID NO:1, optionally further

comprising the amino acid residues set forth in SEQ ID NO:2 (i. e., residues 1330-1375 of SEQ ID NO:1).

In another embodiment according to the present invention, the invention SMRT co-repressor comprises a repression domain having less than about 83% identity with a Sin3A interaction domain of N-CoR (as amino acids 255 to 312 of SEQ ID NO: 11); less than about 57% identity with repression domain 1 of N-CoR (amino acids 1 to 312 of SEQ ID NO: 11); less than about 66% identity with a SANT domain of N-CoR (amino acids 312 to 668 of SEQ ID NO: 11 and/or; less than about 30% identity with repression domain 2 of N-CoR (amino acids 736 to 1031 of SEQ ID NO: 11). Such an encoded SMRT co-repressor or peptide portion thereof is further characterized in that it can modulate transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor).

The invention SMRT co-repressors are additionally exemplified by a full length human SMRT co-repressor, (amino acids 1 to 2517 of SEQ ID NO: 5); and by two mouse SMRT isoforms, including a longer SMRT isoform designated mouse SMRTα, which has an amino acid sequence set forth as amino acids 1 to 2473 of SEQ ID NO: 7; and a shorter SMRT isoform designated mouse SMRTβ (amino acids 1 to 2253 of SEQ ID NO: 9). As compared to the mouse SMRTα isoform (SEQ ID NO: 7), the mouse SMRTβ isoform (SEQ ID NO: 9) has a deletion corresponding to amino acids 36 to 254 of SEQ ID NO: 7.

A peptide portion of a SMRT co-repressor is exemplified herein by amino acids 1 to 1031 of SEQ ID NO: 5; amino acids 1 to 1031 of SEQ ID NO: 7; and amino acids 1 to 813 of SEQ ID NO: 9, which includes the entire amino terminal domain of a SMRT co-repressor. Additional peptide portions of a SMRT co-repressor are exemplified by amino acids 1 to 303 of SEQ ID NO: 7; amino acids 845 to 986 of SEQ ID NO: 7; amino acids 427 to 663 of SEQ ID NO: 7; amino acids 845 to 1055 of SEQ ID NO: 7; amino acids 736 to 1031 of SEQ ID NO: 7; and amino acids 1 to 85 of SEQ ID NO: 9, which are sub-domains of the amino terminal domain

of mouse SMRTα that have nuclear receptor repressor potential, as well as by the corresponding peptide portions of human SMRT and corresponding peptide portions of mouse SMRTβ, which can modulate the transcriptional potential of a nuclear receptor, particularly a nuclear receptor that is in the form of a dimer, for example, a thyroid hormone receptor homodimer, a retinoic acid receptor homodimer, a retinoid X receptor homodimer, a thyroid hormone receptor-retinoid X receptor heterodimer, or a retinoic acid receptor-retinoid X receptor heterodimer. In addition, the invention relates to isolated peptides that contain at least six contiguous amino acids of an amino acid sequence set forth as amino acids 1 to 1030 of SEQ ID NO: 5; amino acids 1 to 1029 of SEQ ID NO: 5; or amino acids 1 to 809 of SEQ ID NO: 9, provided the SMRT peptide is not identical to a sequence of N-CoR (SEQ ID NO: 11).

Invention co-repressor can be an invertebrate SMRT co-repressor, such as the Drosophilia SMRTER co-repressor having an amino acid sequence as set forth in SEQ ID NO: 12, or conservative variations thereof.

Additional exemplary co-repressors are those containing one or both of the receptor interacting domains (ERID1 and ERID2) identified in the Drosophilia co-repressor. For example, co-repressors containing such receptor interacting domains can be selected from the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 1698-1924 of SEQ. ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, amino acids 1698-2063 of SEQ. ID NO:12, amino acids 2094-3040 of SEQ. ID NO:12, amino acids 2094-3040 of SEQ. ID NO:12, amino acids 542-950 of SEQ. ID NO:12, amino acids 542-950 of SEQ. ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2094-3040 of SEO ID NO:12, amino acids 2929-3040 of SEO ID NO:12, and

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amino acids 2951-3038 of SEQ ID NO:12, and conservative variations thereof.

Additional exemplary co-repressors are those containing one or more

of three autonomous repressor domains termed SMRD1, SMRD2, and SMRD3
identified in the SMRTER co-repressor. For example, invention co-repressors can
contain the following autonomous repressor domains derived from Drosophilia
SMRTER co-repressor (SEQ. ID 12):

amino acids 542-950 of SEQ. ID NO:12 amino acids 1698-1924 of SEQ ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, and conservative variations

thereof.

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Conservative variations of the above-described SMRT co-repressors are also contemplated to be within the scope of the present invention. Moreover, proteins, polypeptides and peptides having at least 80% sequence identity with any of the SMRT co-repressors described herein are also contemplated to be within the scope of the invention.

In another embodiment according to the present invention, there are provided chimeric molecules comprising invention isolated peptides and at least a second molecule. For example, the second molecule in invention chimeric molecule can be a polynucleotide or a polypeptide. In one embodiment, the chimeric molecule is a fusion polypeptide comprising a SMRT co-repressor operably linked to a DNA binding domain of a transcription factor.

In another embodiment according to the present invention, there are provided isolated antibodies that bind specifically to invention isolated peptides. In one embodiment, an antibody of the invention binds specifically to an epitope of a SMRT co-repressor. Such an antibody is characterized, in part, in that it does not substantially crossreact with an N-CoR polypeptide. In another embodiment, an

antibody of the invention binds specifically to a complex, which includes a SMRT corepressor or peptide portion thereof of the invention, a nuclear receptor and, optionally, a DNA regulatory element that is specifically bound by the nuclear receptor. Such an antibody is characterized, in part, in that it does not substantially crossreact with the nuclear receptor, either alone or bound to the DNA regulatory element. An antibody of the invention can be a monoclonal antibody, or can be one of a plurality of polyclonal antibodies, which essentially is a mixed population of monoclonal antibodies. The invention also relates to a cell line, which produces the monoclonal antibody of the invention.

Such antibodies can be employed for a variety of purposes, e.g., for studying tissue localization of invention SMRT co-repressor, the structure of functional domains, the purification of receptors, as well as in diagnostic applications, therapeutic applications, and the like. Preferably, for therapeutic applications, the antibodies employed will be monoclonal antibodies.

The above-described antibodies can be prepared employing standard techniques, as are well known to those of skill in the art, using the invention SMRT corepressor or portions thereof as antigens for antibody production. Both anti-peptide and anti-fusion protein antibodies can be used [see, for example, Bahouth et al. (1991)

Trends Pharmacol Sci. vol. 12:338-343; Current Protocols in Molecular Biology

(Ausubel et al., eds.) John Wiley and Sons, New York (1989). Factors to consider in selecting portions of invention SMRT co-repressor for use as immunogen (as either a synthetic peptide or a recombinantly produced bacterial fusion protein) include antigenicity, accessibility (i.e., where the selected portion is derived from, e.g., the ligand binding domain, DNA binding domain, dimerization domain, and the like), uniqueness of the particular portion selected (relative to known receptors and co-repressors therefor), and the like.

In another embodiment according to the present invention, there are provided complexes comprising an invention SMRT co-repressor and a member of

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the nuclear receptor superfamily and isolated antibodies that bind to such complexes. The nuclear receptor can be in the form of a monomer or dimer, for example, a thyroid hormone receptor homodimer, a retinoic acid receptor homodimer, a retinoid X receptor homodimer, a thyroid hormone receptor-retinoid X receptor heterodimer, a retinoic acid receptor-retinoid X receptor heterodimer, a ecdysone receptor-Ultraspiracle receptor heterodimer, and the like. Optionally or alternatively, the complex can include a DNA regulatory element, bound specifically by a DNA binding domain of the nuclear receptor.

The above-described complexes optionally further comprise a response element for the member of the nuclear receptor superfamily. Such response elements are well known in the art. Thus, for example, RAR response elements are composed of at least one direct repeat of two or more half sites separated by a spacer of five nucleotides. The spacer nucleotides can independently be selected from any one of A, C, G or T. Each half site of response elements contemplated for use in the practice of the invention comprises the sequence

-RGBNNM-,

wherein

R is selected from A or G;
B is selected from G, C, or T;
each N is independently selected from A, T, C, or G; and
M is selected from A or C;

with the proviso that at least 4 nucleotides of said -RGBNNM- sequence are identical with the nucleotides at corresponding positions of the sequence -AGGTCA-. Response elements employed in the practice of the present invention can optionally be preceded by N_x, wherein x falls in the range of 0 up to 5.

Similarly, TR response elements can be composed of the same half site repeats, with a spacer of four nucleotides. Alternatively, palindromic constructs as have been described in the art are also functional as TR response elements.

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The above-described SMRT co-repressor/dimeric receptor complexes can be dissociated by contacting the complex with a ligand for the member of the nuclear receptor superfamily.

As employed herein, the term "ligand (or ligand precursor) for a member of the nuclear receptor superfamily" (i.e., intracellular receptor) refers to a substance or compound which, in its unmodified form (or after conversion to its "active" form), inside a cell, binds to receptor protein, thereby creating a ligand/receptor complex, which in turn can activate an appropriate hormone response element. A ligand therefore is a compound which acts to modulate gene transcription for a gene maintained under the control of a hormone response element, and includes compounds such as hormones, growth substances, non-hormone compounds that modulate growth, and the like. Ligands include steroid or steroid-like hormone, retinoids, thyroid hormones, pharmaceutically active compounds, and the like. Individual ligands may have the ability to bind to multiple receptors.

Accordingly, as employed herein, "putative ligand" (also referred to as "test compound") refers to compounds such as steroid or steroid-like hormones, pharmaceutically active compounds, and the like, that are suspected to have the ability to bind to the receptor of interest, and to modulate transcription of genes maintained under the control of response elements recognized by such receptor.

In another embodiment according to the present invention, there are provided polynucleotides encoding members of the above-described family of silencing mediators of retinoic acid and thyroid hormone receptor, or an isoform or peptide portion thereof (SMRT co-repressors), or an isolated polynucleotide complementary thereto.

Invention polynucleotides include those encoding a SMRT corepressor comprises a repression domain having

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- a) less than about 83% identity with a Sin3A interaction domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
- b) less than about 57% identity with repression domain 1 of

N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11;

- c) less than about 66% identity with a SANT domain of
- N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
- d) less than about 30% identity with repression domain 2 of
- N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11.

In addition, an invention polynucleotide can encode a mouse SMRT β isoform having an amino acid sequence as set forth in SEQ ID NO: 9 or conservative variations thereof, or a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 8.

Further examples of invention polynucleotides are those comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 6; nucleotides 1 to 2801 of SEQ ID NO: 8; nucleotides 1 to 8388 of SEQ ID NO: 6; nucleotides 1 to 7465 of SEQ ID NO: 8; and nucleotides 1 to 8561 of SEQ ID NO: 4.

The invention polynucleotides further comprise those encoding a

25 human SMRT co-repressor having an amino acid sequence as set forth in SEQ ID

NO: 5, for example, a nucleotide sequence as set forth in SEQ ID NO: 4; by a

polynucleotide encoding a mouse SMRTα isoform having an amino acid sequence as

set forth in SEQ ID NO: 7, for example, a nucleotide sequence as set forth in SEQ ID

NO: 6; and by a polynucleotide encoding a mouse SMRTβ isoform having an amino

acid sequence as set forth in SEQ ID NO: 9, for example, a nucleotide sequence as set

forth in SEQ ID NO: 8. A polynucleotide of the invention is further exemplified by

polynucleotides encoding peptide portions of a SMRT co-repressor such as a polynucleotide containing nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 7; or nucleotides 1 to 2801 of SEQ ID NO: 8, which can repress the transcriptional activity of nuclear receptor, particularly a nuclear receptor that is in the form of dimer.

Additional invention polynucleotides include those encoding a full length insect SMRTER co-repressor having an amino acid sequence as set forth in SEQ ID NO: 12, or conservative variations thereof.

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Additional exemplary invention polynucleotides are those encoding one or both of the receptor interacting domains (ERID1 and ERID2) identified in invention co-repressors. For example, polynucleotides encoding such receptor interacting domains can be selected from those encoding the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 1698-1924 of SEQ. ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, amino acids 1698-2063 of SEQ. ID NO:12, amino acids 2094-3040 of SEQ. ID NO:12, amino acids 2929-3181 of SEQ. ID NO:12, amino acids 542-950 of SEQ. ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2929-3040 of SEQ ID NO:12, amino acids 2929-3040 of SEQ ID NO:12, and amino acids 2951-3038 of SEQ ID NO:12, and conservative variations thereof.

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Additional exemplary invention polynucleotides are those encoding one or more of three autonomous repressor domains termed SMRD1, SMRD2, and SMRD3 identified in the invention co-repressors. For example, polynucleotides encoding such autonomous repressor domains can be selected from those encoding the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 542-950 of SEQ. ID NO:12 amino acids 1698-1924 of SEQ ID NO:12,

amino acids 2951-3038 of SEQ. ID NO:12, and conservative variations

thereof.

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A polynucleotide that has at least 80% sequence identity or that hybridizes, (preferably under high stringency conditions) with any one of the abovedescribed polynucleotides is also contemplated to be within the scope of this invention.

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A polynucleotide of the invention can be operably linked to a second nucleotide sequence and, therefore, can encode a fusion polypeptide, for example, a SMRT co-repressor, or peptide portion thereof, operably linked to a DNA binding domain of a transcription factor.

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Additional examples of invention isolated oligonucleotides, are those which generally are at least about 15 nucleotides in length and can hybridize specifically to the polynucleotide of the invention, but not to a polynucleotide encoding an N-CoR polypeptide (SEQ ID NO: 11). An oligonucleotide of the invention can be useful as a probe, or as a primer for a PCR procedure, or can encode a peptide containing at least five contiguous amino acids of a SMRT co-repressor. In one embodiment, an oligonucleotide of the invention encodes at least five contiguous amino acids of a sequence such as that shown as amino acids 720 to 745 of SEQ ID NO: 5; or amino acids 716 to 742 of SEQ ID NO: 7; or amino acids 497 to 523 of SEQ ID NO: 9. In another embodiment, an oligonucleotide of the invention can hybridize specifically to a polynucleotide encoding human SMRT (SEQ ID NO: 5) or mouse SMRTα (SEQ ID NO: 7), and, optionally, to a polynucleotide encoding mouse SMRTβ (SEQ ID NO: 9).

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The phrase "substantially the same" as used herein in reference to a nucleotide sequence of DNA, a ribonucleotide sequence of RNA, or an amino acid

sequence of protein, means sequences that have slight and non-consequential sequence variations from the actual sequences disclosed herein. Species that are substantially the same are considered to be equivalent to the disclosed sequences and as such are within the scope of the appended claims. In this regard, "slight and non-consequential sequence variations" means that sequences substantially the same as the DNA, RNA, or proteins disclosed and claimed herein are functionally equivalent to the sequences disclosed and claimed herein. Functionally equivalent sequences will function in substantially the same manner to produce substantially the same compositions as the nucleic acid and amino acid compositions disclosed and claimed herein. In particular, functionally equivalent DNAs encode proteins that are the same as those disclosed herein or that have conservative amino acid variations, such as substitution of a non-polar residue for another non-polar residue or a charged residue for a similarly charged residue. These changes include those recognized by those of skill in the art as those that do not substantially alter the tertiary structure of the protein.

In another embodiment according to the present invention, there are provided vectors comprising an invention polynucleotide, and host cells containing invention polynucleotides. The invention vector can be an expression vector, including, for example, a viral vector, and the polynucleotide, or a vector containing the polynucleotide, can be contained in a host cell. In one embodiment, the polynucleotide of the invention is operably linked to a tissue specific DNA regulatory element. In another embodiment, a SMRT co-repressor or peptide portion thereof encoded by the polynucleotide is expressed in a host cell.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates the repressor potential of a SMRT co-repressor. In this embodiment, the invention method comprises contacting a host cell with an agent, and detecting a change in the level of expression of a first expressible nucleotide sequence in response to the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor. In such a method, the host cell is characterized, in part, in that it contains a first expressible nucleotide

sequence operably linked to a first DNA regulatory element, and expresses a fusion polypeptide composed of an invention SMRT co-repressor, or peptide portion thereof, and a DNA binding domain of a first transcription factor that can specifically bind the first DNA regulatory element. Binding of the DNA binding domain of the first transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence in the host cell.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates a function of an invention SMRT co-repressor. In this embodiment, the invention method comprises contacting an invention SMRT co-repressor, a member of the nuclear receptor superfamily, and an agent, and detecting an altered activity of the SMRT co-repressor in the presence of the agent as compared to the absence of the agent, thereby identifying an agent that modulates a function of the SMRT co-repressor.

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A method of the invention can be performed, for example, by contacting a host cell with an agent, and detecting a change in the level of expression of a first expressible nucleotide sequence in response to the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor. In such a method, the host cell is characterized, in part, in that it contains a first expressible nucleotide sequence operably linked to a first DNA regulatory element, and expresses a fusion polypeptide composed of a SMRT co-repressor or peptide portion thereof of the invention, and a DNA binding domain of a first transcription factor, which can specifically bind the first DNA regulatory element; binding of the DNA binding domain of the first transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence in the host cell. The first expressible nucleotide sequence can be an endogenous gene, which is normally present in the host cell, or can be a sequence that has been introduced into the host cell, either transiently or stably, using methods of recombinant DNA technology. In one embodiment, the first DNA binding domain is a GAL4 DNA binding domain and the first DNA regulatory element is a GAL4 DNA regulatory element that is operably

linked to an expressible nucleotide sequence, for example, a reporter gene, and is introduced into the host cell.

Thus, the invention method can identify an agent that increases or decreases the repressor potential of the SMRT co-repressor, or of an agent that increases or decreases the function of the SMRT co-repressor. The agent can directly interact with the SMRT co-repressor or peptide portion thereof, thereby modulating the repressor potential or function of the SMRT co-repressor, or can interact with a cellular molecule that, in turn, can alter the repressor potential or function of a SMRT co-repressor, thereby increasing or decreasing the repressor potential of the SMRT co-repressor.

The host cell can optionally contain a second expressible nucleotide sequence operably linked to a second DNA regulatory element, and can express a second fusion polypeptide, which is composed of an N-CoR polypeptide, or a repressor domain thereof, and a DNA binding domain of a second transcription factor, which can specifically bind the second DNA regulatory element. By comparing the level of expression of the first expressible nucleotide sequence and the second expressible nucleotide sequence in the host cell upon contacting the host cell with the agent, an agent that independently or coordinately modulates SMRT and N-CoR repressor activity. For example, detecting a change in the level of expression of the first expressible nucleotide sequence, but not in the level of expression of the second expressible nucleotide sequence, due to contacting the host cell with the agent identifies an agent that modulates the repressor potential of a SMRT co-repressor, but not of an N-CoR polypeptide can be identified.

In practicing a method of the invention, the SMRT co-repressor, or peptide portion thereof, can be, for example, an amino acid sequence such as amino acids 1 to 1031 of SEQ ID NO: 5; amino acids 1 to 1031 of SEQ ID NO: 7; or amino acids 1 to 813 of SEQ ID NO: 9. The agent can be, for example, an antibody or antigen binding fragment thereof, a peptide, or a small organic molecule.

In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing an invention isolated polynucleotide into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.

In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing an invention isolated polynucleotide into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.

In performing a method of the invention, an agent that alters an interaction of the SMRT co-repressor, or peptide portion thereof, with the nuclear receptor can be identified using a binding assay, such as an electrophoretic mobility shift assay wherein the level of expression of an expressible nucleotide sequence. Such a method can also identify an agent that alters the ability of the invention SMRT co-repressor, or peptide portion thereof, to interact specifically with the nuclear receptor, but does not alter the level of expression of the expressible nucleotide sequence; or an agent that alters the level of expression of the expressible nucleotide sequence, but does not alter interaction of the SMRT co-repressor or peptide portion thereof with the nuclear receptor; or an agent that alters an interaction of the SMRT co-repressor, or peptide portion thereof, with the nuclear receptor and alters the level of expression of the expressible nucleotide sequence. The agent can, but need not be, a ligand for the nuclear receptor, and the method can be performed in a cell or in a reaction mixture in vitro.

Alternatively, an invention polynucleotide can be introduced into the cell, whereby the polynucleotide, or an expression product of the polynucleotide, alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor. The polynucleotide can encode an invention SMRT co-repressor or peptide, portion thereof, which can be expressed in the cell, thereby increasing the level of a SMRT co-repressor, or peptide portion thereof, in the cell. The polynucleotide also can be an antisense polynucleotide, that decreases the level of a SMRT co-repressor in the cell.

In another embodiment according to the present invention, there are provided methods of identifying a molecule that interacts specifically with a SMRT co-repressor. In this embodiment, invention methods comprise contacting the molecule with an invention SMRT co-repressor and detecting specific binding of the molecule to the SMRT co-repressor, thereby identifying a molecule that interacts specifically with a SMRT co-repressor.

The molecule can be any molecule that interacts specifically with a SMRT co-repressor, including, for example, a small organic molecule such as a drug, a peptide, a nucleic acid molecule, and the like. In one embodiment, the molecule is a cellular factor, for example, a cellular protein that modulates the ability of a SMRT co-repressor to repress transcriptional activity of a nuclear receptor. In another embodiment, the method further involves isolating the molecule that interacts specifically with the SMRT co-repressor or peptide portion thereof.

In accordance with yet another aspect of the present invention, there are provided methods to block the repressing effect of invention SMRT co-repressors, said method comprising administering an effective amount of an antibody as described herein. Alternatively, a silencing domain of a nuclear receptor can be employed. Those of skill in the art can readily determine suitable methods for administering said antibodies, and suitable quantities for administration, which will vary depending on

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numerous factors, such as the indication being treated, the condition of the subject, and the like

In accordance with another aspect of the present invention, there is

provided a method to repress (or silence) the activity of a member of the nuclear receptor superfamily containing a silencing domain that represses basal level promoter activity of target genes, said method comprising contacting said member of the nuclear receptor superfamily with a sufficient quantity of an invention SMRT co-repressor so as to repress the activity of said member. Members of the nuclear receptor superfamily

contemplated for repression in accordance with this aspect of the present invention include, for example, thyroid hormone receptor, retinoic acid receptor, vitamin D receptor, peroxisome proliferator activated receptor, and the like.

In accordance with yet another aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear receptor activity caused by an invention SMRT co-repressor, said method comprising comparing the size of the SMRT co-repressor/dimeric receptor complex (i.e., complexes comprising the invention SMRT co-repressor and a homodimeric or heterodimeric member of the nuclear receptor superfamily) upon exposure to test compound, relative to the size of said complex in the absence of test compound. An observed size corresponding to intact complex is indicative of an inactive compound, while an observed size that reflects dissociation of the complex is indicative of a compound that disrupts the complex, thereby relieving the repression caused thereby. Optionally, the complex employed in this assay further comprises a response element for said member of the nuclear receptor superfamily.

The size of the above-described complex can readily be determined employing various techniques available in the art. For example, electrophoretic mobility shift assays (EMSA) can be employed (wherein receptor alone or receptor-SMRT corepressor complex is bound to target DNA and the relative mobility thereof determined).

Those of skill in the art can readily identify other methodology which can be employed to determine the size of the complex as a result of exposure to putative ligand.

In accordance with a still further aspect of the present invention, there is

provided a method to identify compounds which relieve the repression of nuclear
receptor activity caused by an invention SMRT co-repressor, without substantially
activating said receptor, said method comprising:

comparing the reporter signal produced by two different expression

systems in the absence and presence of test compound,

wherein said first expression system comprises a complex

comprising:

a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer,

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter gene, and optionally, invention SMRT co-repressor, and

wherein said second expression system comprises a complex comprising:

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes,

the same response element-reporter combination as employed in said first expression system, and

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optionally, invention SMRT co-repressor, and thereafter selecting those compounds which provide:

a higher reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of said compound, and

substantially the same reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of an invention SMRT co-suppressor but substantially lacking the ability to activate nuclear receptor activity.

The addition of invention SMRT co-repressor is optional in the above-described assay because it is present endogenously in most host cells employed for such assays. It is preferred, to ensure the presence of a fairly constant amount of SMRT co-repressor, and to ensure that SMRT co-repressor is not a limiting reagent, that SMRT co-repressor be supplied exogenously to the above-described assays.

Mutant receptors contemplated for use in the practice of the present invention are conveniently produced by expression plasmids, introduced into the host cell by transfection. Mutant receptors contemplated for use herein include RAR403 homodimers, RAR403-containing heterodimers, TR160 homodimers, TR160-containing heterodimers, and the like.

Reporter constructs contemplated for use in the practice of the present invention comprise:

- (a) a promoter that is operable in the host cell,
- 30 (b) a hormone response element, and

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(c) a DNA segment encoding a reporter protein,
wherein the reporter protein-encoding DNA segment is
operatively linked to the promoter for transcription of the DNA

segment, and

wherein the hormone response element is operatively linked to the promoter for activation thereof.

Hormone response elements contemplated for use in the practice of the present invention are well known in the art, as has been noted previously.

Exemplary reporter genes include chloramphenicol transferase (CAT), luciferase (LUC), beta-galactosidase (β -gal), and the like. Exemplary promoters include the simian virus (SV) promoter or modified form thereof (e.g., SV), the thymidine kinase (TK) promoter, the mammary tumor virus (MTV) promoter or modified form thereof (e.g., Δ MTV), and the like [see, for example, Mangelsdorf et al., in Nature 345:224-229

(1990), Mangelsdorf et al., in Cell <u>66</u>:555-561 (1991), and Berger et al., in J. Steroid Biochem. Molec. Biol. <u>41</u>:733-738 (1992).

As used herein in the phrase "operative response element" or

"operatively linked" the word "operative" means that the respective DNA sequences
(represented by the terms "GAL4 response element" and "reporter gene") are
operational, i.e., work for their intended purposes; such that after the two segments are
linked, upon appropriate activation by a ligand-receptor complex, the reporter gene will
be expressed as the result of the fact that the "GAL4 response element" was "turned on"

or otherwise activated.

In practicing the above-described functional bioassay, the expression plasmid and the reporter plasmid are co-transfected into suitable host cells. The transfected host cells are then cultured in the presence and absence of a test compound to determine if the test compound is able to produce activation of the promoter operatively linked to the response element of the reporter plasmid. Thereafter, the transfected and

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cultured host cells are monitored for induction (i.e., the presence) of the product of the reporter gene sequence.

Any cell line can be used as a suitable "host" for the functional bioassay contemplated for use in the practice of the present invention. Thus, cells contemplated for use in the practice of the present invention include transformed cells, non-transformed cells, neoplastic cells, primary cultures of different cell types, and the like. Exemplary cells which can be employed in the practice of the present invention include Schneider cells, CV-1 cells, HuTu80 cells, F9 cells, NTERA2 cells, NB4 cells, HL-60 cells, 293 cells, Hela cells, yeast cells, and the like. Preferred host cells for use in the functional bioassay system are COS cells and CV-1 cells. COS-1 (referred to as COS) cells are monkey kidney cells that express SV40 T antigen (Tag); while CV-1 cells do not express SV40 Tag. The presence of Tag in the COS-1 derivative lines allows the introduced expression plasmid to replicate and provides a relative increase in the amount of receptor produced during the assay period. CV-1 cells are presently preferred because they are particularly convenient for gene transfer studies and provide a sensitive and well-described host cell system.

The above-described cells (or fractions thereof) are maintained under

20 physiological conditions when contacted with physiologically active compound.

"Physiological conditions" are readily understood by those of skill in the art to comprise an isotonic, aqueous nutrient medium at a temperature of about 37°C.

In accordance with yet another aspect of the present invention, there is

25 provided a method to identify compounds which activate nuclear receptor activity, but
substantially lack the ability to relieve the repression caused by an invention SMRT corepressor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

comprising:

wherein said first expression system comprises a complex

a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor 5 homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer, a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and 10 optionally, invention SMRT co-repressor, and wherein said second expression system comprises a complex comprising: a homodimeric or heterodimeric form of the same 15 member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, 20 the same response element-reporter combination as employed in said first expression system, and optionally, invention SMRT co-repressor, and thereafter selecting those compounds which provide: 25 a higher reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of compound, and substantially the same reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in 30 the absence of said compound,

wherein said selected compounds are capable of activating nuclear receptor activity, but substantially lacking the ability to relieve the repression caused by a SMRT co-repressor having a structure and function characteristic of, an invention SMRT co-repressor for retinoic acid and thyroid receptors.

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In accordance with a still further aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear receptor activity caused by an invention SMRT co-repressor, and activate said receptor, said method comprising:

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comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

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a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer,

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a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and optionally, invention SMRT co-repressor, and

wherein said second expression system comprises a complex

comprising:

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a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes.

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the same response element-reporter combination as employed in said first expression system, and optionally, invention SMRT co-repressor, and thereafter

5 selecting those compounds which provide:

increased reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of said compound, and

substantially increased reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, and activating said receptor.

20 including:

full length silencing mediator for retinoic acid and thyroid receptors plus

GAL4 DNA binding domain.

full length silencing mediator for retinoic acid and thyroid receptors plus GAL4 activation domain.

full length silencing mediator for retinoic acid and thyroid receptors plus glutathione S-transferase (GST) tag,

and the like.

The above-described modified forms of invention SMRT co-repressor can be used in a variety of ways, e.g., in the assays described herein.

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An especially preferred modified SMRT co-repressor of the invention comprises full length silencing mediator for retinoic acid and thyroid receptors plus GAL4 activation domain.

In accordance with a still further embodiment of the present invention, there is provided a method to identify compounds which disrupt the ability of an invention SMRT co-repressor to complex with nuclear receptors, without substantially activating said receptor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above, a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer or retinoic acid receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

wherein said second expression system comprises a complex comprising:

said modified SMRT co-repressor,

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost

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its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

selecting those compounds which provide:

a lower reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of said compound, and

substantially the same reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of disrupting the ability of a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors to complex with nuclear receptors, without substantially activating said receptor.

Mutant receptors contemplated for use in this embodiment of the present

20 invention include RAR403 homodimers, RAR403-containing heterodimers, TR160

homodimers, TR160-containing heterodimers, and the like.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT (i.e., silencing mediator (SMRT corepressor) for retinoic acid receptor (RAR) and thyroid hormone receptor (TR)) is not endogenous to yeast.

In accordance with yet another embodiment of the present invention, there is provided a method to identify compounds which activate nuclear receptor activity, but substantially lack the ability to disrupt a complex comprising a nuclear receptor and an invention SMRT co-repressor, said method comprising:

comparing the reporter signal produced by two different expression

5 systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above, a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer or retinoic acid receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

wherein said second expression system comprises: said modified SMRT co-repressor,

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

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selecting those compounds which provide:

a higher reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of compound, and

substantially the same reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of compound,

wherein said selected compounds are capable of activating nuclear receptor activity, but substantially lack the ability to disrupt the complex of an invention SMRT co-repressor.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT is not endogenous to yeast.

In accordance with a still further embodiment of the present invention, there is provided a method to identify compounds which activate a nuclear receptor, and disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above,
a homodimeric or heterodimeric member of the nuclear
receptor superfamily selected from thyroid hormone receptor
homodimer, thyroid hormone receptor-retinoid X receptor
heterodimer, retinoic acid receptor homodimer or retinoic acid
receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

5 wherein said second expression system comprises a complex comprising:

said modified SMRT co-repressor,

the same homodimeric or heterodimeric member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

selecting those compounds which provide:

a reduction in reporter signal upon exposure of compound to said first expression system, relative to reporter signal in the absence of said compound, and

increased reporter signal upon exposure of compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of activating a nuclear 25 receptor and disrupting a complex comprising nuclear receptor and a SMRT corepressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT is not endogenous to yeast.

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In accordance with yet another aspect of the present invention, there is provided a method to identify compounds which activate a nuclear receptor and/or disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:

comparing the reporter signals produced by a combination expression system in the absence and presence of test compound,

wherein said combination expression system comprises:

a first homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer.

a second homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first homodimer or heterodimer, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes (i.e., provides basal level expression),

> wherein either said first homodimer (or heterodimer) or said second homodimer (or heterodimer) is operatively linked to a GAL4 DNA binding domain,

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a first reporter,

a GAL4 response element, wherein said response element is operatively linked to a second reporter, and

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optionally a SMRT co-repressor of nuclear receptor activity, said SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, and thereafter

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identifying as capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, but substantially lacking the ability to activate nuclear receptor activity those compounds which provide:

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a higher reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and

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substantially the same reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, or

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identifying as capable of activating nuclear receptor activity, but substantially lacking the ability to relieve the repression caused by a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors those compounds which provide:

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a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of compound, and

substantially the same reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, or

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identifying as capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of

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the silencing mediator for retinoic acid and thyroid receptors, and activating said receptor those compounds which provide:

a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, and

a greater increase in reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound.

Thus, the change in expression level of the two different reporters introduced in a single transfection can be monitored simultaneously. Based on the results of this single transfection, one can readily identify the mode of interaction of test compound with the receptor/SMRT complex.

Exemplary GAL4 response elements are those containing the palindromic 17-mer:

5'-CGGAGGACTGTCCTCCG-3' (SEQ ID NO:3),

20 such as, for example, 17MX, as described by Webster et al., in Cell 52:169-178 (1988), as well as derivatives thereof. Additional examples of suitable response elements include those described by Hollenberg and Evans in Cell 55:899-906 (1988); or Webster et al. in Cell 54:199-207 (1988).

In accordance with still another embodiment of the present invention, there is provided a method to identify compounds which activate a nuclear receptor and/or disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:

comparing the reporter signals produced by a combination expression system in the absence and presence of test compound,

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wherein said combination expression system comprises:

a modified SMRT co-repressor as described above,
a first homodimeric or heterodimeric member of the
nuclear receptor superfamily selected from thyroid hormone
receptor homodimer, thyroid hormone receptor-retinoid X
receptor heterodimer, retinoic acid receptor homodimer, or

retinoic acid receptor-retinoid X receptor heterodimer,

a second homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first homodimer or heterodimer, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes,

> wherein either said first homodimer (or heterodimer) or said second homodimer (or heterodimer) is operatively linked to a GAL4 DNA binding domain,

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a first reporter,

a GAL4 response element, wherein said response element is operatively linked to a second reporter, and thereafter

identifying as capable of disrupting the ability of a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors to complex with a nuclear receptor, without substantially activating nuclear receptor, those compounds which provide:

a lower reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and

substantially the same reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, or

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identifying as capable of activating nuclear receptor activity, but substantially lacking the ability to disrupt a complex comprising a nuclear receptor and a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, those compounds which provide:

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a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of compound, and substantially the same reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound.

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or

identifying as capable of disrupting a complex comprising a nuclear receptor and a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, and activating said receptor those compounds which provide:

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a reduction in reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and

increased reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound.

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In accordance with a still further aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear

receptor activity caused by an invention SMRT co-repressor, said method comprising determining the effect of adding test compound to an expression system comprising:

a modified member of the nuclear receptor superfamily, wherein said
 modified member contains an activation domain which renders said receptor
 constitutively active.

a fusion protein comprising the receptor interaction domain of SMRT operatively linked to the GAL4 DNA binding domain, and

a GAL4 response element operatively linked to a reporter.

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Prior to addition of an effective ligand for the member of the nuclear receptor superfamily employed herein, the association of the modified member and the fusion protein will be effective to bind the GAL4 response element and activate transcription of the reporter. The presence of an effective ligand is indicated by a reduction of reporter signal upon exposure to ligand, which disrupts the interaction of the modified member and fusion protein.

Activation domains contemplated for use in the practice of the present invention are well known in the art and can readily be identified by the artisan.

Examples include the GAL4 activation domain, BP64, and the like.

To summarize, a novel family of nuclear receptor SMRT co-repressor which mediates the transcriptional silencing of RAR and TR has been identified. This discovery is of great interest because transcriptional silencing has been shown to play an important role in development, cell differentiation and the oncogenic activity of v-erbA (Baniahmad et al., EMBO J. 11:1015-1023 (1992)); Gandrillon et al., Cell 49:687-697 (1989)); Zenke et al., Cell 61:1035-1049 (1990); Barlow et al., EMBO J. 13:4241-4250 (1994); Levine and Manley, Cell 59:405-408 (1989); Baniahmad et al., Proc. Natl. Acad. Sci. USA 89:10633-10637 (1992b); and Saitou et al., Nature 374:159-162 (1995)). In fact, v-erbA mutants that harbor the Pro160->Arg change in the TR neither repress basal

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transcription nor are capable of oncogenic transformation (Damm and Evans, (1993), supra).

The function of SMRT as a silencing mediator (co-repressor) of RAR and TR is analogous to mSin3 in the Mad-Max-Sin3 ternary complex (Schreiber-Agus et al., Cell 80:777-786 (1995); and Ayer et al., Cell 80:767-776 (1995)). Because GAL-SMRT functions as a potent repressor when bound to DNA, it is reasonable to speculate that the function of the unliganded receptors is to bring with them SMRT to the template via protein-protein interaction. Thus, the repressor function is intrinsic to SMRT as opposed to the TR or RAR itself (Baniahmad et al., Proc. Natl. Acad. Sci. USA 90:8832-8836 (1993); and Fondell et al., Genes Dev 7:1400-1410 (1993)). It is demonstrated herein that the ligand triggers a dissociation of SMRT from the receptor, which would lead to an initial step in the activation process. This would be followed (or be coincident) with an induced conformational change in the carboxy-terminal transactivation domain (c, also called AF2), allowing association with co-activators 15 on the transcription machinery (Douarin et al., EMBO J. 14:2020-2033 (1995); Halachmi et al., Science 264:1455-1458 (1994); Lee et al., Nature 374:91-94 (1995); and Cavailles et al., Proc. Natl. Acad. Sci. USA 91:10009-10013 (1994)). Thus, as has previously been suggested (Damm and Evans, (1993), supra), the ligand dependent activation of TR would represent two separable processes including relief of repression 20 and net activation. The isolation of SMRT now provides a basis for dissecting the molecular basis of trans-repression.

The invention will now be described in greater detail by reference to the following non-limiting examples. 25

Example 1 Isolation of SMRT

Using a GAL4 DBD-RXR fusion protein (see, for example, USSN 08/177,740, incorporated by reference herein in its entirety) as a bait in a yeast

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two-hybrid screening system (Durfee et al., (1993), supra), several cDNA clones encoding receptor interacting proteins were isolated. One of these proteins, SMRT, interacts strongly with unliganded RAR and TR but only weakly with RXR or other receptors in yeast. This protein was selected for further characterization.

Example 2

Far-western blotting procedure

Total bacteria extracts expressing GST fusions of hRAR α (aa 156-462) or hRXR α LBD (aa 228-462) and control extracts expressing GST alone or GST-PML fusion protein were subjected to SDS/PAGE and electroblotted onto nitrocellulose in transfer buffer (25 mM Tris, pH 8.3/192 mM glycine/ 0.01% SDS). After denaturation/renaturation from 6 M to 0.187 M guanidine hydrochloride in HB buffer (25 mM HEPES, pH 7.7/25 mM NaCl/5 mM MgCl₂/1 mM DTT) filters were saturated at 4°C in blocking buffer (5% milk, then 1% milk in HB buffer plus 0.05% NP40). *In vitro* translated 35 S-labeled proteins were diluted into H buffer (20 mM Hepes, pH 7.7/75 mM KCl/0.1 mM EDTA/2.5 mM MgCl₂/0.05% NP40/ 1% milk/1 mM DTT) and the filters were hybridized overnight at 4°C with (1 μ M) or without ligand. After three washes with H buffer, filters were dried and exposed for autoradiography or quantitated by phosphoimager.

GST-SMRT is a GST fusion of the C-SMRT encoded by the yeast two hybrid clone. GST-SMRT has been purified, but contains several degradation products.

25 For yeast two-hybrid screening, a construct expressing the GAL4
DBD-hRXRα LBD (aa 198-462) fusion protein was used to screen a human lymphocyte
cDNA library as described (Durfee et al., (1993), *supra*). Full length SMRT cDNA was
isolated from a human HeLa cDNA library (Clontech) using the two-hybrid insert as a
probe.

Using the above-described far-western blotting procedure, ³⁵S-labeled SMRT preferentially complexes with bacterial extracts expressing the RAR, marginally associates with RXR and shows no association with control extracts. In contrast, ³⁵S-PPAR selectively associates with its heterodimeric partner, RXR, but not with RAR. In a similar assay, ³⁵S-labeled RAR or TR interacts strongly with SMRT and their heterodimeric partner, RXR, but not with degraded GST products, while ³⁵S-RXR interacts only weakly with SMRT. Binding of ligand to RAR or TR reduces their interactions with SMRT but not with RXR, while binding of ligand to RXR has only slight effect. Figure 1 shows the quantitation of a dose-dependent dissociation of SMRT from RAR or TR by all-trans retinoic acid (atRA) or thyroid hormone (triiodothyronine or T3), demonstrating that the amount of ligand required for 50% dissociation in both cases are close to the kds for both ligands (Munoz et al. EMBO J. 7:155-159 (1988); Sap et al., Nature 340:242-244 (1989); and Yang et al., Proc. Natl. Acad. Sci. USA 88:3559-3563 (1991)).

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Full length SMRT encodes a polypeptide of 1495 amino acids rich in proline and serine residues (see Figure 2 and SEQ ID NO:1). Genbank database comparison reveals similarity of the C-terminal domain of SMRT to a partial cDNA encoding another receptor interacting protein, RIP13 (Seol et al., (1995), *supra*), whose role in receptor signaling is unknown. Within this region, there can be identified several potential heptad repeats which might mediate protein-protein interaction with the "a-helical sandwich" structure (Bourguet et al., *Nature* 375:377-382 (1995)) of the ligand binding domain (LBD) of receptors.

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Example 3 Characterization of SMRT

Unlike other nuclear receptors, unliganded RAR and TR possess a strong silencing domain which represses basal level promoter activity of their target genes

(Damm et al., *Nature* 339:593-597 (1989); Brent et al., *New Biol.* 1:329-336 (1989);

Baniahmad et al., *Cell* 61:505-514 (1990); and Baniahmad et al., *EMBO J.*

11:1015-1023 (1992)). The preferential interaction of SMRT with RAR and TR in the absence of hormone suggests that SMRT may play a role in mediating the transcriptional silencing effect of the receptor.

To further investigate the involvement of SMRT in silencing, the interaction of SMRT with mutant receptors which display distinct silencing and/or transactivation activities was tested as follows. 35 S-methionine labeled receptors were used as probes to hybridize immobilized GST-SMRT in the presence (10 μ M) or absence of all-trans retinoic acid (atRA). The total bacteria extract expressing GST-RXR was included as a control.

When quantitated by phosphoimager, RAR403 shows a 4-fold better interaction with SMRT than wild type RAR. Both full length RAR or a deletion mutant expressing only the ligand binding domain (LBD, referred to as $\Delta\Delta$ R) associate with SMRT; this association is blocked by ligand.

These results confirm that the LBD alone is sufficient in the interaction. The carboxy-terminal deletion mutant RAR403 is a potent dominant negative repressor of basal level promoter activity of RAR target genes (Damm et al., *Proc. Natl. Acad. Sci. USA* 90:2989-2993 (1993); Tsai and Collins, *Proc. Natl. Acad. Sci. USA* 90:7153-7157 (1993); and Tsai et al., *Genes Dev* 6:2258-2269 (1992)). As might be predicted from the above studies, RAR403 and its amino terminal deletion derivative, R403, interact strongly with SMRT in either the presence or absence of ligand, consistent with SMRT mediating the repressor activity of this mutant.

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Example 4 Interaction of SMRT with TR Mutants

The interaction of SMRT with two different classes of TR mutants was analyzed next. The first mutant employed is the naturally occurring oncogene, v-erbA, which has strong silencing ability but no transactivation activity (Sap et al., (1989),

supra; Sap et al., Nature 324:635-640 (1986); Weinberger et al., Nature 318:670-672 (1985); and Weinberger et al., Nature 324:641-646 (1986)). The second mutant employed is a single amino acid change (Pro 160 -> Arg) of the rTRa (TR160) which has previously been shown to lose its capacity in basal level repression but retains hormone dependent transactivation (Thompson et al., Science 237:1610-1614 (1987); and Damm and Evans, Proc. Natl. Acad. Sci. USA 90:10668-10672 (1993)). If SMRT is involved in silencing, it would be expected that SMRT should interact with the v-erbA, but show little or no association with the silencing-defective TR160 mutant.

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Interaction of the oncogenic v-erbA and rTR α R160 mutant (TR160) with GST-SMRT was determined in a far-western assay as described above (see Example 2). When quantitated by phosphoimager, the v-erbA shows an 18-fold better interaction with SMRT than hTR β , and the TR160 mutant shows a 10-fold lower signal than the rTR α .

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As one might expect, v-erbA interacts strongly with SMRT both in presence or absence of ligand. In contrast, full length TR160 mutant or LBD of TR160 ($\Delta\Delta$ TR160) does not interact significantly with SMRT when compared to the wild type receptor.

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These data demonstrate that SMRT plays an important role in mediating transcriptional silencing effects of both RAR and TR. These data also suggest that the release of SMRT from receptors could be a prerequisite step in ligand-dependent transactivation by nuclear receptors.

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Example 5 Formation of ternary complexes containing SMRT

RAR and TR form heterodimers with RXR, resulting in a complex with 30 high DNA binding ability (Bugge et al., EMBO J. 11:1409-1418 (1992); Yu et al., Cell 67:1251-1266 (1991); and Kliewer et al., Nature 355:446-449 (1992)). Since SMRT

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interacts with RAR and TR, tests were conducted to determine whether SMRT can also interact with the receptor-DNA complex. Thus, the interaction of SMRT with RXR-RAR heterodimer on a DR5 element (i.e., an AGGTCA direct repeat spaced by five nucleotides) was determined in a gel retardation assay, which is carried out as follows. *In vitro* translated receptor or unprogrammed reticulocyte lysate (URL) was incubated with 1 µg of poly dIdC on ice for 15 minutes in a total volume of 20 µl containing 75 mM KCl, 7.5% glycerol, 20 mM Hepes (pH 7.5), 2 mM DTT and 0.1% NP-40, with or without ligand (in the range of about 10-100 nM employed). A ³²P labeled, double stranded oligonucleotide probe was added into the binding reaction (10,000 cpm per reaction), and the reaction was further incubated for 20 minutes at room temperature. The protein-DNA complex was separated on a 5% native polyacrylamide gel at 150 volts.

SMRT is seen to form a ternary complex with the RXR-RAR heterodimer on a DNA response element in the gel retardation assay. Addition of ligand releases SMRT from this complex in a dose-dependent manner.

Similarly, SMRT is seen to form a ternary complex with the RXR-TR heterodimer on a TR response element; addition of T3 disrupts the formation of this complex.

These data demonstrate that SMRT can be recruited to DNA response elements via protein-protein interaction with RAR or TR in the absence of hormone. Binding of hormone disrupts receptor-SMRT interaction and releases SMRT from the receptor-DNA complex.

Example 6 Transient transfection assay

30 CV-1 cells were plated in 24 well plates at a density of 50,000 cells per well. Expression plasmids were transfected into cells by lipofection using DOTAP. In each transfection, 5 ng of GAL-RAR and 15 ng of v-erbA or SMRT were used together with 150 ng of reporter construct containing 4 copies of GAL4 binding sites in front of a minimal thymidine kinase promoter and a CMX-β-gal construct as an internal control.

The relative luciferase activity was calculated by normalizing to the β-gal activity.

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Example 7 Reversal of transcriptional silencing

Recently, it has been shown that over expression of RAR or TR could

reverse the transcriptional silencing effect of the GAL4 DBD fusion of TR (GAL-TR) or
RAR (GAL-RAR) (Baniahmad et al., Mol Cell Biol 15:76-86 (1995); and Casanova et
al., Mol Cell Biol 14:5756-5765 (1994)), presumably by competition for a limiting
amount of a SMRT co-repressor. A similar effect is observed herein when over
expression of v-erbA or RAR403 mutants are shown to reverse the silencing effect of

GAL-RAR and GAL-TR on the basal activity of a luciferase reporter (see Figure 3A and
3B).

In principle, over expression of SMRT should restore repressor activity when co-expressed with v-erbA or RAR403 competitors. Indeed, results presented in 20 Figure 3C show that both the full length and the C-terminal domain of SMRT (C-SMRT) can titrate out v-erbA or RAR403 competitor activity and re-endow GAL-RAR and GAL-TR with silencing activity. In contrast, neither v-erbA nor SMRT show any effect on the transactivation activity of GAL-VP16 fusion. Thus, SMRT is able to block the titration effect of v-erbA and RAR403 and functionally replaces the putative SMRT co-repressor in this system.

Example 8 Direct recruitment of SMRT to a heterologous promoter

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If SMRT is the mediator of transcription silencing of TR and RAR by interaction with template-bound unliganded receptors, then direct recruitment of SMRT

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to a heterologous promoter should result in repression of basal level activity. This was tested by fusing full length SMRT to the GAL4 DBD (GAL-SMRT). The effect of the resulting fusion protein on the activity of the thymidine kinase promoter containing four GAL4 binding sites was analyzed. Figure 3D shows that GAL-SMRT, like GAL-TR, can silence basal promoter activity in a dose-dependent manner. In contrast, GAL-RXR shows no repression.

These data suggest that SMRT, when recruited to a promoter by direct DNA binding or via association with an unliganded receptor, functions as a potent transcriptional repressor.

Example 9 Cloning Of Human And Mouse SMRT co-repressors

This example describes the cloning of a full length human silencing mediator of retinoic acid and thyroid hormone receptor (SMRT co-repressor) and of two mouse SMRT isoforms. m-SMRT α and m-SMRT β .

An examination of the previously described human SMRT co-repressor 20 revealed that the first eight amino acids and unstream sequences were derived from a portion of ribonucleoprotein K sequence. Accordingly, a mouse spleen cDNA lambda ZAP II library (Stratagene; La Jolla CA) was screened at low stringency with a probe corresponding to approximately the 5' 1,000 base pairs (bp) of the previously identified human SMRT (s-SMRT). A 3.5 kilobase (kb) cDNA fragment was obtained that 25 contained a unique sequence in addition to known s-SMRT sequence. The 5' end of this cDNA, and subsequently obtained clones, was used in successive rounds of screening of the mouse spleen cDNA library and a mouse brain cDNA library (Stratagene) and the full-length SMRTα isoform cDNA (SEQ ID NO: 6) and SMRTα isoform cDNA (SEQ ID NO: 10) were obtained. The mouse SMRT (m-SMRT) 5' sequence then was used at 30 low stringency to screen a human pituitary cDNA library (Stratagene) to obtain the fulllength human SMRT (h-SMRT) cDNA (SEQ ID NO: 1). All cDNA clones were

sequenced on both strands using standard methods, and have been deposited with GenBank as Accession No. AF103003 (h-SMRT; SEQ ID NOS: 3 and 5); Accession No. 113001 (m-SMRTα; SEQ ID NOS: 6 and 7); and Accession No. 113002 (m-SMRTβ; SEO ID NOS: 8 and 9).

By sequentially shifting between the mouse spleen and mouse brain cDNA libraries, several clones containing a potential starting methionine and 5' untranslated region sequences were obtained. The complete polypeptide sequences of m-SMRT (SEQ ID NO: 7) and h-SMRT (SEQ ID NO: 5) are provided. In addition, a splice variant isolated from the mouse brain cDNA library encoded an m-SMRT corepressor containing a deletion of amino acids 36 to 254 of SEQ ID NO: 7 (see SEQ ID NO: 3). The two m-SMRT co-repressors are designated SMRTα (SEQ ID NO: 7) and SMRTβ (SEQ ID NO: 9). Based on sequence similarity to N-CoR (see below), this deletion in m-SMRTβ removes the majority of the sequence in h-SMRT and m-SMRTα that is homologous to N-CoR repression domain 1 (RD1), including a portion of the Sin3A binding region.

The cloned h-SMRT (SEQ ID NO: 3) encodes a polypeptide that contains an additional 1130 amino acids at the amino terminus as compared to the previously described human SMRT co-repressor. The full length h-SMRT shares 84% identity with m-SMRTa. A comparison of h-SMRT (SEQ ID NO: 5) and N-CoR (SEQ ID NO: 11) revealed that the N-terminal extension of h-SMRT (amino acids 1 to 1030) and N-CoR (amino acids 1 to 1031) share approximately 41% identity, which is somewhat higher that the 36% identity shared between the full length proteins. However, regions within the N-CoR and SMRT N-termini share striking homology (Figures 4A and 4B).

Amino acids 1 to 160 of N-CoR are moderately conserved in h-SMRT (and m-SMRT α), sharing about 36% identity. This region of N-CoR has been reported to interact with Siah2 (Zhang et al., (1998), *supra*) and, similarly, can be involved in an

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interaction of Siah2 with h-SMRT or m-SMRTα. In particular, highly conserved sequences in this region can be the specific Siah2 interaction sites (see Figure 4A).

A 52 amino acid segment from N-CoR (amino acids 255 to 312)

mediates an interaction with Sin3A (Heinzel et al., *Nature* **387**:43-48 (1997)), and was presumed to represent the core of the larger RD1 region (Horlein et al., (1995), *supra*). This small interaction domain is highly conserved (about 83% identity) in h-SMRT, and the overall identity shared between SMRT and N-CoR RD1 is about 57%.

Amino acids 312 to 668 of N-CoR also are well conserved (66% identity) in h-SMRT (and m-SMRT α), and two internal blocks of sequences in this region share even greater similarity (see Figure 1B; shaded regions). These blocks are homologous to each other and to part of the SANT domain, which was identified in the yeast chromatin remodeling factor, SWI3, the yeast adapter protein, ADA2, the basal transcription factor TFIIIB, and other proteins (Aasland et al., *Trends Biochem. Sci.* 21:87-88 (1996)), suggesting that these domains share a common and important function. The amino acids of N-CoR RD2 (see Horlein et al., (1995) supra) are the least conserved in h-SMRT, sharing about 30% identity.

These results demonstrate that isoforms of SMRT co-repressors are expressed in cells, as exemplified by m-SMRT α and m-SMRT β . In addition, the results demonstrate that the previously undescribed amino terminus of SMRT co-repressors shares regions of substantial homology with N-CoR, and regions of homology are identified that indicate these sequences can mediate previously uncharacterized functions

Example 10 Expression And Chromosomal Localization Of Smrt Co-Repressors

30 This example describes the tissue distribution of SMRT RNA and the chromosomal localization of human SMRT. Total RNA was prepared from adult CB6F1 mouse tissues using TRIZOL reagent (GIBCO/BRL), and poly(A) RNA was purified from total RNA using an OLIGOTEX mRNA Kit (Qiagen, Valencia, CA). RNA was separated on 1.25% agarose/6% formaldehyde gels and transferred to a NYTRAN membrane (Scheicher & Schuell). A 720 bp m-SMRT/PstI fragment was used as a probe. Following hybridization with the SMRT probe, the filters were stripped and hybridized with a murine glyceraldehyde-3-phosphate dehydrogenase cDNA probe to allow normalization for RNA loading.

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Chromosomal localization of SMRT was determined by fluorescence in situ hybridization using the 5.3 kb h-SMRT cDNA clone. The probe was labeled by nick-translation with biotin-11-dUTP, then hybridized to normal male human metaphase chromosomes. Chromosomes were counterstained with 4',6-diamidino-2-phenylindole (DAPI). Chromosome identification was carried out by computer inversion of the gray scale DAPI image on a PSI Imaging System (Perceptive Scientific Instruments; League City TX). Chromosome 12 confirmation was carried out using a chromosome 12-specific alpha satellite probe (Vysis; Downers Grove IL).

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Previous studies using the short human SMRT co-repressor suggested that SMRT was expressed ubiquitously in various tissues. To confirm this result, expression of the full length m-SMRT was determined by northern blot analysis by using a probe consisting of nucleotides 2760 to 3620 of m-SMRT (SEQ ID NO: 6). The expression pattern was ubiquitous, as previously described, although higher levels were detected in lung, spleen, and brain. Similarly, h-SMRT was expressed ubiquitously as determined using a multiple tissue blot (CLONTECH; Palo Alto CA). It is noteworthy that two isoforms of SMRT were present in the majority of the mouse tissues and likely correspond to the m-SMRTα and m-SMRTβ isoforms.

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The chromosomal location of the h-SMRT and N-CoR genes was mapped. The h-SMRT clone hybridized to the q arm of one of the C group

chromosomes. Computer-mediated banding of the DAPI stained chromosomes identified the labeled chromosome as chromosome 12, band q24. The chromosome 12 localization was confirmed by cohybridization of SMRT and a chromosome 12 alpha satellite probe, D12Z3 (Vysis), which labels the pericentromeric region of chromosome 12. The location for the human N-CoR gene was determined through a mapped human bacterial artificial chromosome clone, hCIT529I10, which is 158 kb of genomic N-CoR and resides on chromosome 11p11.2. The SMRT and N-CoR chromosomal locations can be accessed through GENEMAP98 from the Human Genome Project at http://www.ncbi.nlm.nih.gov/genemap.

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These results demonstrate that the full length SMRT co-repressors and the SMRT co-repressors are expressed in various tissues. The results also demonstrate that the human SMRT gene is located on chromosome 12.

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Example 11 Functional Characterization Of SMRT Amino Terminus Domains

This example demonstrates that various domains of the SMRT amino 20 terminus can repress nuclear receptor transcriptional activity.

Experiments were performed using the plasmids pCMX-GAL4 DBD and pMH100-TK-luc (Nagy et al., (1997), *supra*). Standard PCR amplifications were used to generate GAL4 fusion constructs. All constructs were verified by double-stranded sequencing to confirm identity and reading frame.

Monkey CV-1 cells were grown in DMEM supplemented with 10% resin-charcoal stripped fetal bovine serum (FBS), 50 units/ml of penicillin G, and 50 μ g/ml of streptomycin sulfate at 37°C in 7% CO₂. V-1 cells (60-70% confluence, 48-well plate) were cotransfected with 16 ng of pCMX-GAL4, 100 ng of pMH100-TK-luc, and 100 ng of pCMX- β galactosidase in 200 μ l of DMEM containing 10% super-

stripped fetal calf serum (FCS) by the N-(1-(2,3-dioleoyloxy)propyl)-N,N,N-trimethylammonium methylsulfate (DOTAP)-mediated procedure (Nagy et al., (1997), supra). The amount of DNA in each transfection was kept constant by addition of pCMX. After 24 hr, the medium was replaced; cells were harvested and assayed for luciferase activity 36 to 48 hr after transfection. Luciferase activity was normalized by the level of β-galactosidase activity. Each transfection was performed in triplicate and repeated at least three times.

Based on the high degree of identity between regions of the SMRT amino terminus and the corresponding N-CoR region, the ability of regions in the SMRT amino terminus to act in transcriptional repression was examined. A nested series of nucleotide sequences encoding portions of the SMRT amino terminus fused to the GAL4 DNA binding domain (GAL-DBD) was prepared in mammalian expression vectors (Figure 5A). The constructs were cotransfected with a GAL4-TK-luciferase reporter plasmid to determine the regulatory properties of the GAL4-SMRT fusions. Repression was determined relative to the basal activity of the reporter in the presence of the GAL-DBD alone.

The entire SMRT amino terminus region (GAL4-SMRT(1-1031)) demonstrated the greatest amount of repression (approximately 38-fold), and virtually extinguished reporter activity. In comparison, GAL4-SMRT (1-303), which is equivalent to N-CoR RD1, demonstrated 6-fold repression; and GAL4-SMRT (736-1031), which is equivalent to N-CoR RD2, demonstrated about 2.6-fold repression. Surprisingly, the highly conserved SANT domain conferred a significant amount of repression (about 3.3-fold).

A smaller region (amino acids 845 to 986) within the RD2 homology region shows a higher level of sequence conservation as compared to the entire RD2 region. Deletion constructs were generated to determine whether this minimal region was sufficient for the repression activity of RD2. Deletion of flanking amino acids 736 to 845 or of amino acids 987 to 1055 did not affect the level of repression, demonstrating

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that the repressor function of RD2 is contained within a 141 amino acid core sequence of RD2

Based on sequence similarity to N-CoR, the deletion of amino acids 36 to 254 in the m-SMRTβ isoform removes the majority of RD1, including a portion of the Sin3A binding region. The effect of this deletion on SMRT function was examined by cotransfection experiments comparing repression by SMRTα to SMRTβ. These experiments revealed that SMRTβ has substantially less repressor activity than SMRTα. A construct containing the entire amino terminus of m-SMRTβ (amino acids 1 to 813) repressed activity about 2.6 fold, as compared to m-SMRTα amino acids 1 to 1031, which repressed activity about 38.1-fold. In addition, a GAL4 construct containing m-SMRT amino acids 1 to 83 repressed activity only about 1.4-fold. These results indicate that alternative splicing can add further diversity to expand the function of SMRT gene products.

Example 12 Yeast Two-Hybrid Screen and Assays

To investigate whether repression by EcR in CV-1 cells is mediated by its association with a vertebrate corepressor and whether such an interaction, if it does occur, is impaired by the A483T mutation, a mammalian two-hybrid assay with Gal4-c-SMRT was conducted.

A yeast two-hybrid screen (Fields and Song, Nature, 340:245-246,

25 (1989)) was performed by transforming approximately 2 X 10⁶ Y190 yeast cells with
a pAS-EcR construct and a Drosophila (0-8 hr) embryonic c-DNA two-hybrid library
(Yu et al., Nature, 385:552-555, (1997)). Transformants were selected onto DO-LeuTrp-His plates containing 40 mM 3-aminotriazole (Sigma) for 3-4 days. Surviving
yeast colonies were picked as primary positives and restreaked on selection plates to
30 isolate single clones. Activation domain plasmids were rescued from the selected
positive transformants for further analysis. Each clone was evaluated by testing its

potential interaction with several other nuclear receptors using the yeast two-hybrid assays. E52 was isolated and further pursued based on this selection criterion. Quantitative liquid assay of β -galactosidase was performed on positive clones 16 hr after treating the yeast cells with no ligand, or with 3 μ M ligand.

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pAS-EcR is a fusion gene with the region corresponding to amino acids 223-878 of EcRB1 fused C-terminally to the Gal4-DBD of the pAS1-CYH2 construct (Durfee et al., (1993), supra); other Gal4-DBD-based nuclear receptor constructs used in this yeast two-hybrid assay include: USP (amino acids 50-508), hRAR (amino acids 186-462) and hTR (amino acids 121-410) (Schulman et al., (1995), supra), and SMRT (Chen and Evans, (1995), supra). β-galactosidase activities were quantified by liquid assay for yeast cells treated either without ligand or with 3 μM of corresponding hormone. All-trans retinoic acid (ATRA) is a ligand of RAR; 3,3',5-triiodothyroacetic acid (T3) is a ligand of TR.

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Similar yeast two-hybrid assays were also used to examine the interaction between SMRTER and mSin3A and dSin3A.

Example 12 Cloning SMRTER

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To isolate full-length SMRTER cDNA, a XhoI insert fragment isolated from the E52 clone was used to screen male and female Tudor c-DNA libraries (gift of Tulle Hazelrigg). This initial screen resulted in isolating three overlapping c-DNA clones covering the region of amino acid 2094 to the C terminus of SMRTER Additional regions were obtained from three consecutive library screens using two cosmid clones isolated from the Tamkun genomic library (gift of John Tamkun). Sequences of these overlapping c-DNA and genomic clones were assembled to obtain a conceptual open reading frame of SMRTER 3446 amino acids in length (SEQ ID NO:12; Figure 8A). The translational initiation codon was designated based on the sequences that match the consensus Kozak codons and is preceded by three in-frame

consecutive stop codons in the upstream region. Both strands of the sequences of the c-DNA clones were determined using an ABI prism Big Dye® terminator cycle sequencing ready reaction kit (PE Biosystems) and ABI 377 instrument.

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Example 14 Plasmids

CMV promoter-driven expression plasmids of EcR, USP, RXR, c-SMRT, β-galactosidase, and pMH100-TK-luc reporter, and yeast plasmids of RAR, TR, and SMRT have been described previously (Yao et al., (1992), *supra*, Yao et al., (1993), *supra*; Chen and Evans, (1995), *supra*; Schulman et al., (1995), *supra*; Chen et al., *Proc. Natl. Acad. Sci. USA* 93:7567-7571, (1996); Nagy et al., (1997), *supra*). hsp27EcR-TK-Luc, a reporter with six copies of the hsp27EcRE, is a gift of Barry Forman. CMV vector-driven EcR A483T and Gal4-SMRD3 mutations were generated using the Transformer® site-directed mutagenesis kit (Clontech) with proper selection primers and the mutagenic primers that correspond to the missense mutation (A483T) of EcR and to the designated mutations, M2 and M3, in the SMRD3 domain, respectively. Other plasmids were constructed with standard techniques, including various enzyme digestions or PCR amplification.

Example 15 Cell Culture and Transfection

25 CV-1 cells were grown in Dulbecco's modified Eagles medium at 37°C in 5% CO₂. The media were supplemented with 10% AG1-X8 resin charcoal double-stripped calf bovine serum, 50 U/ml penicillin G, and 50 μg/ml streptomycin sulfate. Approximately 20 hr after CV-1 cells (10⁵ cells) were plated in 48-well cell culture clusters (Costar), cells were transiently transfected with plasmids using 30 DOTAP according to the instructions of the manufacturer (Boehringer Mannheim). The amount of CMV promoter-driven expression vectors, β-galactosidase gene

expression vector, CMX-lacZ, and reporter, pMH100-TK-luc or hsp27EcRE-TK-Luc, were in the range of 100-200 ng, 500 ng, and 400 ng, respectively, for six wells of each 48-well clusters in each transfection experiments. At least 4 hr after transfection, each medium was replaced with medium either without ligand, or with 1 μ M of MurA. Cells were harvested and assayed approximately 48 hr after transfection. All experiments were performed in triplicate and repeated with similar results.

CV-1 cells were transfected with wild-type EcR or EcR A483T, along with vp16-USP and a reporter, hsp27EcRE-TK-Luc, which contains six copies of the hsp27EcRE fused to the thymidine kinase (TK) promoter-luciferase reporter. VP16-USP fusion contains the region of USP (amino acids 50-508) fused C-terminally to the VP16 domain. Muristerone A (MurA) is a potent ecdysone agonist (Christopherson et al., *Proc. Natl. Acad. Sci. USA*, 89:6314-6318, (1992)). In all experiments, cells were also cotransfected with CMV-lacZ, which is used to normalize the luciferase activity. As shown in Figure 6A, the ability to dimerize with USP is reflected in reporter activity without treatment with hormone (open bar), and the ability to respond to hormone is reflected in reporter activity when cells were treated with 1 µM Muristerone A (closed bar).

CMV promoter-driven expression vector including wild-type EcR or EcR A483T was cotransfected with VP16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), *supra*) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine→threonine) at the 483 site of EcR (Bender et al., (1997), *supra*). The results of this experiment (Figure 6B) show that EcR A483T disrupts the interaction with SMRT.

Example 16 In Vitro Interacting Assays

Glutathione S-transferase fusion proteins, including GST-X, GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1), and GST-ERID2 (amino acids 2951-3038 of SEO ID NO:1), were expressed in E. coli DH5 cells, and extracts were affinity purified by binding to glutathione Sepharose 4B beads. Bound proteins used as affinity matrices in pull-down experiments were first equilibrated with the binding buffer (20 mM HEPES [pH 7.9], 150 mM NaCl, 1 mM EDTA, 4 mM MgCl2, 1 mM DTT, 0.06% NP40, 10% Glycerol, 0.25 mM PMSF, 1 mg BSA). For pull-down 10 assays using GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) and GST-ERID2 (amino acids 2951-3038 of SEQ ID NO:1), additional hsp27EcRE (0.05 µg/ml) was added to the binding buffer. In this experiment, 30 ul of 50% GST-protein beads slurry, containing approximately 1 µg of proteins, were incubated with 10 µl of 35Smethionine-labeled proteins in 300 µl of the binding buffer (with or without 3 µM of 15 MurA as indicated) for 30 min at room temperature. After the incubation, beads were washed three times with the binding buffer (with or without ligand) and resuspended in SDS-PAGE sample buffer before loading. After electrophoresis, bound radiolabeled proteins were visualized by autoradiography. 35S-methionine-labeled EcR, USP were generated in a coupled transcription-translation system, TNT (Promega), 20 using CMX-EcR (T7) and CMX-uspK (T7) constructs as templates, respectively.

Example 17 Immunohistochemistry and Immunofluorescence

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Antibodies against SMRTER were raised in rabbits immunized with bacterially expressed glutathione S-transferase fusion proteins corresponding to the region (amino acids 2477-2648 of SEO ID NO:1) of SMRTER. Specific antibodies were purified by affinity chromatography through antigen-linked columns and used at 1:200 dilution for tissue staining. Tissues for whole-mount staining were dissected at the wandering third instar stage of the Canton-S strain larvae and fixed (4%

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formaldehyde in 1? PBS, 50 mM EGTA) for at least 30 min. Preincubation, secondary antibodies, washes, and peroxidase reactions are described in the protocol of the Elite ABC (Rabbit IgG) kit (Vector). For the pilot experiments, partially purified IgG from preimmunization serum was used. For polytene chromosome staining, salivary glands were dissected according to the method described in Zink et al., EMBO J., 10:153-162, (1991).

Chromosome spreads were costained with affinity-purified anti-SMRTER (1:100) polyclonal antibody and with anti-USP monoclonal antibody (ABIII/AD5; gift of F. Kafatos, 1:100 dilution). SMRTER was detected with Texas red-conjugated donkey anti-rabbit secondary antibody (1:100 dilution), and USP was detected with FITC-conjugated donkey anti-mouse secondary antibody (1:100 dilution) (Jackson ImmunoResearch Labs).

Example 18 ER Interacts Genetically with DSinA

In keeping with the evidence that dSin3A is a component in EcR regulatory pathway, an experiment was conducted to examine whether dSin3A interacts genetically with EcR using several previously characterized Drosophila EcR and dSin3A mutants (Bender et al., (1997), supra; Neufeld et al., (1998), supra). In the experiment, in which female dSin3AK07401 were crossed with male EcRE261st using techniques known in the art (see Table 1 below), only approximately 14% of the scored EcRE261st/dSin3AK07401 progenies survived, a percent that is significantly lower than the expected 33.3%. This suggests that a large portion of the EcRE261st/dSin3AK07401 flies either die prior to eclosion or fail to eclose. Additionally, surviving EcRE261st/dSin3AK07401 escapers showed delayed development and wing defects, in which wings are held horizontally at 45°-90° angle from the body axis. These results suggest that dSin3A shares an overlapping regulatory pathway with EcR.

In a reverse genetic cross, in which female EcRE261st were crossed with male dSin3AK07401, none of the EcRE261st/dSin3AK07401 flies survived to adulthood. These results suggest that EcRE261st/dSin3AK07401 results in a genetically sensitized background. When the maternally deposited EcR in embryos descended from female EcRE261st/SM6b was cut in half, the lethality for EcRE261st/dSin3AK07401 was further increased. These results reveal that, in addition to its previously known zygotic function, EcR also contributes maternally to Drosophila development.

10 Table 1

		EcR ^{E261st} /DSin3A ^{RO7401}
Cross		Surviving Rate (%)
DSin3AKO7401/CyO	φ	
×		14 (n = 141)
EcR ^{261st} /SM6b	₫	
EcR ^{261st} /SM6b	Ŷ.	
×		0 (n = 144)
DSin3AKO7401/CyO	₫	
A similar wing held-out phen Df(2R)nap11/DSin3A ^{KO7401} ,	otype is also observ	red in EcRE261st/DSin3Axe374,
Df(2R)nap11/DSin3AKO7401,	and Df(2R)nap11/D	Osin2A ^{xe374} . EcR ^{E261st} and
Df(2R)nap11 are both describ	ed in Figure 6. Ds	in2AKO7401 is an allele with a P
element insertion within the 5	' intron of Sin3A.	DSin3Axe374 is an X ray-generated
allele (Neufeld et al., (1998)).	n=the number of	surviving flies scored. Note that
CvO/SM6b is lethal.		

EcRA483T flies showed developmental abnormalities in wings and tergites.

- 15 A similar phenotype, although with a lower penetration rate, has been also observed in EcRA483T/Df(2R)20B and in EcRA483T/Df(2R)nap11. Df(2)20B and Df(2)nap11 are both deficiencies in which EcR is deleted (Bender et al., (1997), supra). Sequence alignment of EcR with the vertebrate TR, RAR, and v-erbA, an oncogenic TR variant, revealed that alanine 483 is located within a highly conserved.
- 20 23-amino acid (aa) loop region connecting helices 3 and 4, termed the LBD signature

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motif (Wurtz et al., Nat. Struct. Biol., 3:206, (1996)) (see Figure 6C). Based on structural studies of vertebrate nuclear receptors (for review, see Moras and Gronemeyer, (1998), supra), this alanine residue appears to be on the exposed surface, consistent with it being a potential corepressor binding site for nuclear receptors.

These in vivo studies indicate that EcRA483T is a semilethal allele (Bender et al., (1997), *supra*). When EcRA483T is in trans with EcRE261st, an allele that removes both the DBD and LBD domains of EcR, animals are primarily lethal (>95%). The few surviving EcRA483T/EcRE261st flies, however, display significant delays in development, blistered wings, and defective tergites, indicating that EcR is involved in the development of these tissues. The ability of EcR to bind a vertebrate corepressor and the loss of this property in EcR A483T suggests that the defects observed in EcRA483T flies may result from the disruption of its interaction with an as yet unidentified Drosophila corepressor.

Example 19 Isolation of an EcR-Interacting Factor

20 The CMV promoter-driven expression vector including wild-type EcR or EcR A483T, was cotransfected with vp16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), *supra*) into CV-1 cells to examine its effect on the interaction of the invertebrate SMRTER with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine→threonine) at the 483 site of EcR (Bender et al., (1997), *supra*). Although EcR readily interacted with SMRT in both mammalian and yeast cells (Figure 6B; Figure 7), repeated low-stringency hybridization screens failed to identify a Drosophila homolog of SMRT. No

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Example 20 Isolation and Characterization of an

EcR-Interacting Clone - Yeast Two-hybrid screen

To pursue the isolation of an EcR corepressor, a yeast two hybrid interaction screen was performed of a Drosophila embryonic cDNA library using pAS-EcR as bait. E52 was isolated as one of the complementary positive clones from a yeast two-hybrid screen with pAS-EcR as bait, as described in Example 12.

Example 21

Characterization of a Repression-Defective EcR Allele, EcRA483T

- (A) CV-1 cells were transfected with wild-type EcR or EcR A483T, along with vp16-USP and a reporter, hsp27EcRE-TK-Luc, which contains six copies of the hsp27EcRE fused to the thymidine kinase (TK) promoter-luciferase reporter. In all experiments, cells were also cotransfected with CMV-lacZ, which is used to normalize the luciferase activity. The ability to dimerize with USP was reflected in reporter activity without treatment with hormone (open bar), and the ability to respond to hormone was reflected in reporter activity when cells were treated with 1 μM Muristerone A (closed bar). vp16-USP fusion contains the region of USP (amino acids 50-508) fused C-terminally to the vp16 domain. Muristerone A (MurA) is a potent ecdysone agonist (Christopherson et al., (1992), supra). In these tests EcR A483T was selectively defective in repression.
- 25 (B) CMV promoter-driven expression vector including wild-type EcR or EcR A483T was cotransfected with vp16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), supra) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine threonine) at the 483 site of EcR (Bender et al., (1997), supra). The

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results of this test show that EcR A483T disrupts the interaction with SMRT.

(C) Sequence alignment of EcR with the vertebrate TR, RAR, and verbA, an oncogenic TR variant, reveals that the alanine 483 of the EcRA4831T mutant is located within a highly conserved 23-amino acid (aa) loop region connecting helices 3 and 4, termed the LBD signature motif (Wurtz et al., (1996), supra) (Figure 6C). Based on structural studies of vertebrate nuclear receptors (for review, see Moras and Gronemeyer, (1998), supra), this alanine residue appears to be on the exposed surface, consistent with it being a potential corepressor binding site for nuclear receptors.

In vivo studies indicated that EcRA483T is a semilethal allele (Bender et al., (1997), supra). When EcRA483T is in trans with EcRE261st, an allele that removes both the DBD and LBD domains of EcR, animals are primarily lethal (>95%). The few surviving EcRA483T/EcRE261st flies, however, display significant delays in development, blistered wings, and defective tergites, indicating that EcR is involved in the development of these tissues. The ability of EcR to bind a vertebrate corepressor and the loss of this property in EcR A483T suggested to us that the defects observed in EcRA483T flies may result from the disruption of its interaction with an as yet unidentified Drosophila corepressor.

Example 22 Isolation of an EcR-Interacting Factor

25 Although EcR readily interacts with SMRT in both mammalian and yeast cells (Figure 6B; Figure 7), repeated low-stringency hybridization screens failed to identify a Drosophila homolog of SMRT. Given that no SMRT/N-CoR homolog is found in C. elegans, it was believed that either a SMRT/N-CoR-like corepressor is not conserved in invertebrates or, alternatively, invertebrate corepressors may diverge significantly from their vertebrate counterparts. To pursue the isolation of an EcR corepressor, a yeast interaction screen of a Drosophila embryonic cDNA library using

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EcR as bait was conducted as described in Example 19. This screen resulted in the isolation of a clone, E52, whose protein product interacts with EcR as well as with the vertebrate RAR and TR, but notably not with USP (Figure 7). Unlike the interaction between E52 and RAR, which can be dissociated by all-trans retinoic acid, the interaction between E52 and EcR, or the interaction between SMRT and EcR, is not dissociated by Muristerone A (MurA). This result suggests that other factors essential for the dissociation of E52 from EcR, such as USP, are missing in yeast (see below).

Example 23

Isolation and Characterization of an EcR-Interacting Clone

E52 was isolated as one of the complementary positive clones from a yeast two-hybrid screen. Isolation of overlapping cDNA and genomic clones led to the identification of a full-length sequence encoding a large protein of 3446 amino acids (Figure 8A). This protein contains several unusually long stretches of Gln, Ala, Gly, and Ser repeats. Comparative analysis reveals it to be a novel protein with limited regions of clear homology with the vertebrate nuclear receptor corepressors SMRT and N-CoR (Chen and Evans, (1995), supra; Hörlein et al., (1995), supra; Ordentlich et al., (1999), supra; Park et al., (1999), supra). This protein SMRTER, SMRT-related ecdysone receptor-interacting factor, was shown by Northern blot analysis to encode large transcripts (>12 kb) expressed broadly throughout the embryonic stage and three larvae stages, as well as in adult Drosophila flies.

Example 24

Molecular and Biochemical Analysis for ERID1 and ERID2

Interaction with the EcR complex was evaluated based on transient transfection with the Gal4-SMRTER fusion genes. USP, EcR-vp16 (VP16 transactivating domain was fused C-terminally to the end of the EcRB1 isoform), and the reporter, pMH100-TK-Luc.

In vitro pull down assays (Example 12) were conducted to determine whether EcR interacts with ERID1 and ERID2. In vitro translated 35S-methionine-labeled EcRB1 alone, or a mixture of 35S-methionine-labeled EcRB1 and unlabeled USP, or 35S-methionine-labeled USP alone, were incubated with GST, GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1), or GST-ERID2 (amino acids of SEQ ID NO:1). GST-ERID1 and GST-ERID2, but not GST alone, pull down labeled EcR, whereas little interaction is found between USP and any of the three GST proteins. In addition, the pull-down complex was disrupted by the addition of 3μM MurA when USP is present. These in vitro results establish that SMRTER and EcR may interact directly.

Further in vitro tests were conducted to determine ERID1, ERID2, and c-SMRT compete with each other to bind EcR. Gal4-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) or Gal4-ERID2 (amino acids 2929-3181 of SEQ ID NO:1), along with EcR-vp16 and USP, were transfected in CV-1 cells as described above. In this competition experiment, additional ERID1, ERID2, and c-SMRT (Chen et al., (1996), supra) were cotransfected into cells. ERID1 (1698-2063) and ERID2 ((amino acids 2929-3038 of SEQ ID NO:1) were tagged with the nuclear targeting signal (MAPKKKRKV) (SEQ ID NO:3) to ensure that these proteins were localized in nuclei. As shown in Figure 11C, interaction between each Gal4-ERID fusion and EcR-vp16:USP was significantly decreased by both ERIDs and by c-SMRT. Interestingly, a more prominent effect was observed in experiments when Gal4-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) was challenged by ERID2, and, conversely, a more efficient competition was achieved by ERID1 to Gal4-ERID2 (amino acids 2094-3181 of SEQ ID NO:1). Together, these results suggest that ERID1, ERID2, and c-SMRT may bind similar or overlapping surface(s) in EcR.

Example 25 SMRTER Colocalizes with the EcR on Polytene Chromosomes

SMRTER antibodies were prepared as described in Example 12 to examine its cytological and chromosomal localization patterns of SMRTER.

Consistent with its action as a corepressor of EcR, SMRTER was localized to nuclei of salivary glands and of fat bodies, as well as to nuclei of eye, wing, and leg imaginal dises isolated from the third instar larvae.

Next association of SMRTER with the EcR:USP complex on chromosomes was examined. The USP staining pattern was used as an index for EcRs presence on chromosomes. Since USP and EcR colocalized with each other on polytene chromosomes (Yao et al., (1993), supra), chromosomal spreads prepared from the salivary glands of wandering third instar larvae (prior to pupariation) were subjected to simultaneous immunological staining with antibodies against SMRTER and USP. SMRTER was detected with antibody conjugated with Texas red, USP with FITC.

To visualize the band, interband, and puffing patterns of the polytene chromosomes, the chromosomes were counterstained with DAPI to show the banding regions while leaving the interbands and puffs unstained or lightly stained. Indirect immunofluorescence staining revealed that SMRTER is a chromosome-bound protein and colocalizes with USP (FITC) at a majority of chromosomal sites; whereas in a pilot experiment, no such staining patterns were detected using the preimmunization serum. The strongest SMRTER staining was primarily associated with the boundary between band and interband regions as well as within the interband regions of chromosomes counterstained with DAPI. This result confirms that, as an EcR-associating factor, SMRTER is recruited by the EcR:USP heterodimers to their specific target chromosomal loci.

SMRTER staining can still be detected in puffed regions, such as the 2B puff. Since the polytene chromosomes consist of a parallel arrangement of several hundred to two thousand copies of the euchromatic portions of the chromosomes, an individual binding protein like SMRTER may be cycling on and off, resulting in a steady state of signals detected in the broader chromatin regions. Whether or not SMRTER levels actually change prior to or after the peak of ecdysone pulses remains to be established.

While the invention has been described in detail with reference to certain

10 preferred embodiments thereof, it will be understood that modifications and variations
are within the spirit and scope of that which is described and claimed.

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That which is claimed is:

- An isolated polynucleotide encoding a member of a family of silencing mediators of retinoic acid receptor and thyroid hormone receptor, or an isoform or peptide portion thereof (SMRT co-repressor), or an isolated polynucleotide complementary thereto.
- The polynucleotide of claim 1, which modulates transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor).
- 3. The polynucleotide of claim 2, wherein the SMRT co-repressor comprises a repression domain having
 - a) less than about 83% identity with a Sin3A interaction domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
 - b) less than about 57% identity with repression domain 1 of N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11;
 - c) less than about 66% identity with a SANT domain of N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
 - d) less than about 30% identity with repression domain 2 of N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11, and polynucleotides that hybridize thereto under stringent conditions.
- 4. The polynucleotide of claim 1, wherein the SMRT co-repressor is a human SMRT co-repressor having an amino acid sequence as set forth in SEQ ID NO: 5 or conservative variations thereof.
- A polynucleotide which hybridizes under stringent conditions with a polynucleotide according to claim 2.

- A polynucleotide that has at least 80% sequence identity with a
 polynucleotide according to claim 2.
- 7. The polynucleotide of claim 4, which has a nucleotide sequence as set forth in SEQ ID NO: 4, and conservative variations thereof.
- 8. The polynucleotide of claim 1, wherein the SMRT co-repressor is a mouse SMRT α isoform.
- The polynucleotide of claim 6, having an amino acid sequence as set forth in SEQ ID NO: 7 or conservative variations thereof.
- $10.\,$ The polynucleotide of claim 4, which has a nucleotide sequence as set forth in SEQ ID NO: 6.
- 11. The polynucleotide of claim 1, wherein the SMRT co-repressor is a mouse SMRT β isoform.
- 12. The polynucleotide of claim 11, having an amino acid sequence as set forth in SEQ ID NO: 9 or conservative variations thereof.
- The polynucleotide of claim 11, which has a nucleotide sequence as set forth in SEQ ID NO: 8.

14. The polynucleotide of claim 1, comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 6; and nucleotides 1 to 2801 of SEO ID NO: 8.

- 15. A polynucleotide that under stringent conditions with a polynucleotide according to claim 14, provided that the polynucleotide does not contain a sequence identical to SEQ ID NO: 11.
- 16. A polynucleotide that has at least 80% sequence identity with a polynucleotide according to claim 14, provided that the polynucleotide does not contain a sequence identical to SEQ ID NO: 11.
- 17. A polynucleotide of claim 1, comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 8388 of SEQ ID NO: 6; and nucleotides 1 to 7465 of SEQ ID NO: 8.

- 18. The polynucleotide of claim 1, comprising nucleotides 1 to 8561 of SEQ ID NO: 4.
- 19. The polynucleotide of claim 1, which is operably linked to a second nucleotide sequence.

- 20. The polynucleotide of claim 19, which encodes a fusion polypeptide comprising the SMRT co-repressor operably linked to a DNA binding domain of a transcription factor.
 - 21. A vector comprising the polynucleotide of claim 1.
 - 22. A host cell containing the polynucleotide of claim 1.
- 23. An isolated oligonucleotide, comprising at least 15 nucleotides that can hybridize specifically to the polynucleotide of claim 1, but not to a polynucleotide encoding SEQ ID NO: 11 or to a polynucleotide encoding an amino acid sequence consisting of amino acids 1031 to 2517 of SEQ ID NO: 5.
- 24. The oligonucleotide of claim 23, wherein the polynucleotide encodes at least five contiguous amino acids of a sequence selected from the group consisting of:

amino acids 720 to 745 of SEQ ID NO: 5; amino acids 716 to 742 of SEQ ID NO: 7; and amino acids 497 to 523 of SEQ ID NO: 9.

25. The oligonucleotide of claim 23, which can hybridize specifically to a polynucleotide encoding SEQ ID NO: 5 or SEQ ID NO: 7, but not to a polynucleotide encoding SEQ ID NO: 9.

26. An isolated silencing mediator of retinoic acid and thyroid hormone receptor, or isoform or peptide portion thereof (SMRT co-repressor), wherein the co-repressor modulates transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor).

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- 27. An isolated co-repressor comprising a repression domain having
 - a) less than about 83% identity with a Sin3A interaction
- domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
 - b) less than about 57% identity with repression domain 1 of
- N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11; c) less than about 66% identity with a SANT domain of
 - c) less than about 60% identity with a SANY domain of
- N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
- d) less than about 30% identity with repression domain 2 of N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11.

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28. An isolated peptide, comprising at least six contiguous amino acids of an amino acid sequence selected from the group consisting of:

amino acids 1 to 1030 of SEQ ID NO: 5;

amino acids 1 to 1029 of SEQ ID NO: 7;

amino acids 1 to 809 of SEQ ID NO: 9;

and conservative variations thereof,

provided the peptide is not identical to a sequence of SEQ ID NO: 11.

- An isolated antibody that binds specifically to the peptide of claim
 - 30. A cell line, which produces the antibody of claim 29.
- A chimeric molecule, comprising the SMRT co-repressor of claim
 and at least a second molecule.

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- A complex, comprising a SMRT co-repressor of claim 26 and a member of the nuclear receptor superfamily (nuclear receptor).
- 33. The complex of claim 32, wherein the nuclear receptor is in the form of a dimer.
- 34. A method for identifying an agent that modulates the repressor potential of a SMRT co-repressor, the method comprising:
 - a) contacting a host cell with an agent,
 wherein the host cell contains a first expressible nucleotide
 sequence operably linked to a first DNA regulatory element, and
 expresses a fusion polypeptide comprising a SMRT corepressor of claim 26, and a DNA binding domain of a first transcription
 factor, which can specifically bind the first DNA regulatory element,
 and wherein binding of the DNA binding domain of the first

transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence; and

 b) detecting a change in the level of expression of the first expressible nucleotide sequence due to contacting the host cell with the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor.

- 35. A method for identifying an agent that modulates a function of a SMRT co-repressor, the method comprising:
 - a) contacting a SMRT co-repressor of claim 26,
 a member of the nuclear receptor superfamily (nuclear receptor), and

an agent; and

b) detecting an altered activity of the SMRT co-repressor in the presence of the agent as compared to the absence of the agent, thereby identifying an agent that modulates a function of the SMRT co-repressor.

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- 36. A method of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing a polynucleotide of claim 1 into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.
- 37. A method of identifying a molecule that interacts specifically with a SMRT co-repressor, the method comprising:
 a) contacting the molecule with the SMRT co-repressor of
 - claim 26; and

 b) detecting specific binding of the molecule to the SMRT corepressor, thereby identifying a molecule that interacts specifically with a
 SMRT co-repressor.

ABSTRACT OF THE INVENTION

The present invention relates to isolated polynucleotides encoding a family of silencing mediators of retinoic acid and thyroid hormone receptor (SMRT) isoforms, including vertebrate and invertebrate isoforms thereof. For example, a full length human SMRT co-repressor, two isoforms of a mouse SMRT-- a longer form. mouse SMRT α , and a shorter form, mouse SMRT β , and an isoform of an insect (Drosophilia), SMRTER -- as well as peptide portions of the SMRT co-repressors that can modulate transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor); to oligonucleotides that can hybridize specifically to such a polynucleotide; to vectors and to host cells containing such polynucleotides. The invention also relates to polypeptide SMRT co-repressors encoded by such invention SMRT polynucleotides, and to peptide portions thereof that can modulate transcriptional potential of a nuclear receptor; including peptide portions of a SMRT co-repressor that are not present in an N-CoR polypeptide. In addition, the invention relates to chimeric molecules and to complexes containing a SMRT co-repressor or peptide portion thereof, to antibodies that specifically bind such compositions, and to methods for identifying an agent that modulates the repressor potential of a SMRT corepressor. The invention also provides methods for identifying an agent that modulates a function of a SMRT co-repressor; for modulating the transcriptional potential of a nuclear receptor in a cell using the compositions of the invention; and for identifying a molecule that interacts specifically with a SMRT co-repressor.

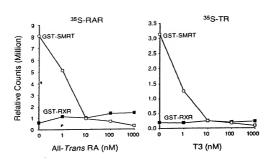


FIGURE 1

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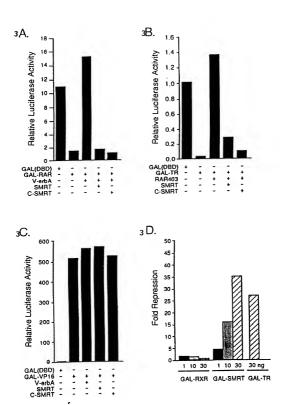


FIGURE 3

| Part |

FIGURE 4

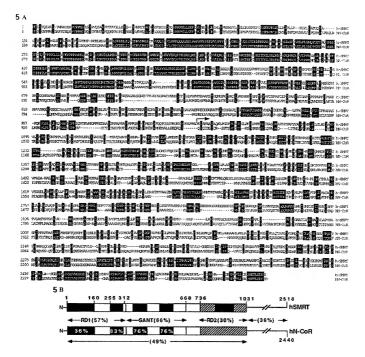


FIGURE 5

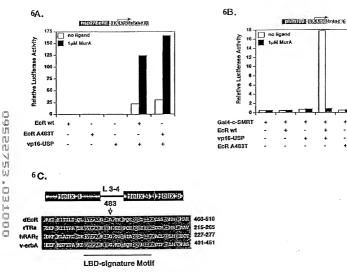


FIGURE 6

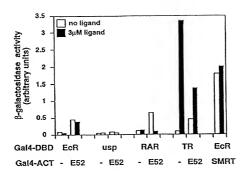


FIGURE 7

DECLARATION FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship is as stated below next to my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR, which is a C-I-P of 08/522,726, filed on September 1, 1995, the specification of which

applic	able (the "Application").	
	, and was amended on,	f
X	was filed on March 10, 2000, as U.S. Application Serial No.	
X	is attached hereto. (SALK1510-3)	

 $\label{thm:continuous} In the reby authorize and request insertion of the application serial number of the Application when officially known.$

I hereby state that I have reviewed and understand the contents of the aboveidentified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, Code of Federal Regulations ("C.F.R."), § 1.56.

With respect to the Application, I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

(Application Serial No.)	(Filing Date)
(Application Serial No.)	(Filing Date)
(Application Serial No.)	(Filing Date)

With respect to the Application, I hereby claim the benefit under 35 U.S.C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of the application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, C.F.R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of the Application:

08/522,726 (Application Serial No.)	09/01/95 (Filing Date)	
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so 2 GT\6172856.1

made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of first inventor: Ronald M. Evans
Inventor's signature:
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Citizenship: USA
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Inventor's signature:
Date:
Residence: San Diego, California
Citizenship: Taiwan
Post Office Address: 7548 Charmant Drive, #1416 San Diego, California 92126
Full name of third inventor: Peter Ordentlich
Inventor's signature:
Date:
Residence:
Citizenship:
Post Office Address:

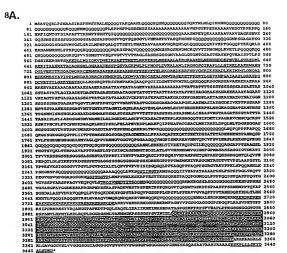




FIGURE 8

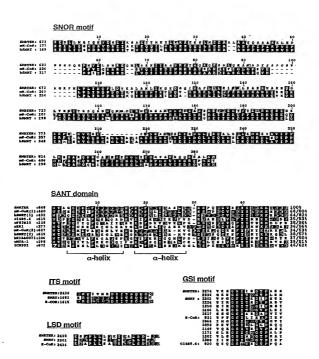


FIGURE 9

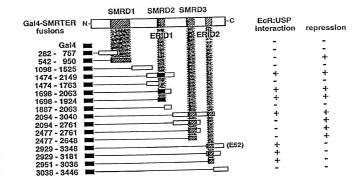
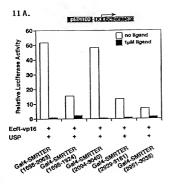


FIGURE 10



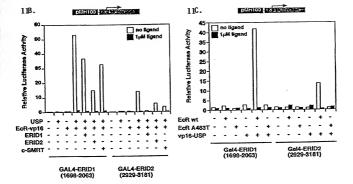
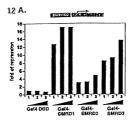
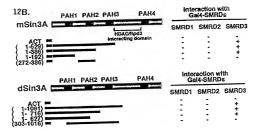


FIGURE 11





SMRTER: 2564 EXECUTORAL SARPENDO SERVESK 44

GSINSA Interaction - - G

G GGH-SMRTS GGH

FIGURE 12

SEQUENCE LISTING

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<120> A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS

AND USES THEREFOR

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Gly Leu His Asp Thr Ala Arg Pro Val Leu Pro Arg Pro Pro Thr Ile 70 75

Ser Asn Pro Pro Pro Leu Ile Ser Ser Ala Lys His Pro Ser Val Leu 85 90

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Val Pro Tyr Ser Glu His Ala Lys Ala Pro Val Gly Pro Val Thr Met 120 125 115

Gly Leu Pro Leu Pro Met Asp Pro Lys Lys Leu Ala Pro Phe Ser Gly 135

Val Lys Gln Glu Gln Leu Ser Pro Arg Gly Gln Ala Gly Pro Pro Glu 150 155

Ser Leu Gly Val Pro Thr Ala Gln Glu Ala Ser Val Leu Arg Gly Thr 165 170

Ala Leu Gly Ser Val Pro Gly Gly Ser Ile Thr Lys Gly Ile Pro Ser 180 185

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1210

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ctg Leu	aat Asn 610	gag Glu	agt Ser	tct Ser	cgc Arg	tgg Trp 615	aca Thr	gaa Glu	gaa Glu	gaa Glu	atg Met 620	gaa Glu	aca Thr	gcc Ala	aag Lys	1873
					cac His 630											1921
					gtg Val											1969
					ctc Leu											2017
atg Met	gag Glu	aag Lys 675	gag Glu	agg Arg	aac Asn	gcg Ala	cgg Arg 680	agg Arg	aag Lys	aag Lys	aag Lys	aaa Lys 685	gcg Ala	ccg Pro	gcg Ala	2065

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	gcg Ala	gcc Ala 690	agc Ser	gag Glu	gag Glu	gct Ala	gca Ala 695	ttc Phe	ccg Pro	ccc Pro	gtg Val	gtg Val 700	gag Glu	gat Asp	gag Glu	gag Glu	21	13
	atg Met 705	gag Glu	gcg Ala	tcg Ser	ggc Gly	gtg Val 710	agc Ser	gga Gly	aat Asn	gag Glu	gag Glu 715	gag Glu	atg Met	gtg Val	gag Glu	gag Glu 720	21	61
	gct Ala	gaa Glu	gcc Ala	tta Leu	cat His 725	gcc Ala	tct Ser	gly aaa	aat Asn	gag Glu 730	gtg Val	ccc Pro	aga Arg	gly aaa	gaa Glu 735	tgc Cys	22	09
	agt Ser	ggc Gly	cca Pro	gcc Ala 740	act Thr	gtc Val	aac Asn	aac Asn	agc Ser 745	tca Ser	gac Asp	acc Thr	gag Glu	agc Ser 750	atc Ile	ccc Pro	22	57
	tct Ser	cct Pro	cac His 755	act Thr	gag Glu	gcc Ala	gcc Ala	aag Lys 760	gac Asp	aca Thr	gly ggg	cag Gln	aat Asn 765	gly ggg	ccc Pro	aag Lys	23	05
	ccc Pro	cca Pro 770	gcc Ala	acc Thr	ctg Leu	ggc Gly	gcc Ala 775	gac Asp	ggg gly	cca Pro	ccc Pro	cca Pro 780	ggc Gly	cca Pro	ccc Pro	acc Thr	23	53
	cca Pro 785	cca Pro	cgg Arg	agg Arg	aca Thr	tcc Ser 790	cgg Arg	gcc Ala	ccc Pro	att Ile	gag Glu 795	ccc Pro	acc Thr	ccg Pro	gcc Ala	tct Ser 800	24	01
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u : ::	gca Ala	cct Pro	cct Pro	cct Pro 820	gtg Val	gtc Val	ccc Pro	aag Lys	gag Glu 825	gag Glu	aag Lys	gag Glu	gag Glu	gag Glu 830	acc Thr	gca Ala	24	97
	gca Ala	gcg Ala	ccc Pro 835	cca Pro	gtg Val	gag Glu	gag Glu	999 Gly 840	gag Glu	gag Glu	cag Gln	aag Lys	ccc Pro 845	ccc Pro	gcg Ala	gct Ala	25	45
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	gag Glu 865	tgc Cys	acg Thr	gag Glu	gaa Glu	gcc Ala 870	gag Glu	gag Glu	ggg Gly	ccg Pro	gcc Ala 875	aag Lys	ggc Gly	aag Lys	gac Asp	gcg Ala 880	26	541
	gag Glu	gcc Ala	gct Ala	gag Glu	gcc Ala 885	acg Thr	gcc Ala	gag Glu	ggg	gcg Ala 890	ctc Leu	aag Lys	gca Ala	gag Glu	aag Lys 895	aag Lys	26	89
	gag Glu	ggc Gly	Gly aaa	agc Ser 900	ggc	agg Arg	gcc Ala	acc Thr	act Thr 905	gcc Ala	aag Lys	agc Ser	tcg Ser	ggc Gly 910	gcc Ala	ccc Pro	27	737
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ctc Leu 945	acc Thr	ccg Pro	act Thr	ggc Gly	gac Asp 950	ccc Pro	cgg Arg	gcc Ala	aat Asn	gcc Ala 955	tca Ser	ccc Pro	cag Gln	aag Lys	cca Pro 960	2881
ctg Leu	gac Asp	ctg Leu	aag Lys	cag Gln 965	ctg Leu	aag Lys	cag Gln	cga Arg	gcg Ala 970	gct Ala	gcc Ala	atc Ile	ccc Pro	ccc Pro 975	atc Ile	2929
cag Gln	gtc Val	acc Thr	aaa Lys 980	gtc Val	cat His	gag Glu	ccc Pro	ccc Pro 985	cgg Arg	gag Glu	gac Asp	gca Ala	gct Ala 990	ccc Pro	acc Thr	2977
aag Lys	cca Pro	gct Ala 99	ccc Pro	cca Pro	gcc Ala	cca Pro	ccg Pro 1000	Pro	ccg Pro	caa Gln	aac Asn	ctg Leu 1009	Gln	ccg Pro	gag Glu	3025
		Ala	cct Pro				Gly					Gly				3073
agc Ser 102	Pro	gca Ala	ccc Pro	ccc Pro	gcc Ala 103	Asp	aag Lys	gag Glu	gcc Ala	ttc Phe 103!	Ala	gcc Ala	gag Glu	gcc Ala	cag Gln 1040	3121
aag Lys	ctg Leu	cct Pro	ggg Gly	gac Asp 104	Pro	cct Pro	tgc Cys	tgg Trp	act Thr 105	Ser	ggc Gly	ctg Leu	ccc Pro	ttc Phe 105	Pro	3169
gtg Val	ccc Pro	ccc Pro	cgt Arg 106	Glu	gtg Val	atc Ile	aag Lys	gcc Ala 106	Ser	ccg Pro	cat His	gcc Ala	ecg Pro 107	Asp	ccc Pro	3217
			tcc Ser 5					Gly					Leu			3265
cat His	gac Asp 109	Thr	gcc Ala	cgg Arg	ccc Pro	gtc Val 109	Leu	ccg Pro	cgc Arg	cca Pro	ccc Pro 110	Thr	atc Ile	tcc Ser	aac Asn	3313
ccg Pro 110	Pro	ccc	ctc Leu	atc Ile	tcc Ser 111	Ser	gcc Ala	aag Lys	cac His	ecc Pro 111	Ser	gtc Val	ctc Leu	gag Glu	agg Arg 1120	3361
caa Gln	ata Ile	ggt Gly	gcc Ala	atc Ile 112	Ser	caa Gln	gga Gly	atg Met	tcg Ser 113	Val	cag Gln	ctc Leu	cac His	gtc Val 113	Pro	3409
tac Tyr	tca Ser	gag Glu	cat His 114	Ala	aag Lys	gcc Ala	ccg Pro	gtg Val 114	Gly	cct Pro	gtc Val	acc Thr	atg Met 115	Gly	ctg Leu	3457
ccc Pro	ctg Leu	Pro	atg Met 5	gac Asp	ccc Pro	aaa Lys	aag Lys 116	Leu	gca Ala	ccc Pro	ttc Phe	agc Ser 116	Gly	gtg Val	aag Lys	3505

cag Gln	gag Glu 1170	Gln	ctg Leu	tcc Ser	cca Pro	cgg Arg 1175	Gly	cag Gln	gct Ala	G1 999	cca Pro 1180	Pro	gag Glu	agc Ser	ctg Leu	3553
999 Gly 1189	Val	ccc Pro	aca Thr	gcc Ala	cag Gln 1190	Glu	gcg Ala	tcc Ser	gtg Val	ctg Leu 1195	aga Arg	gly gag	aca Thr	gct Ala	ctg Leu 1200	3601
ggc Gly	tca Ser	gtt Val	ccg Pro	ggc Gly 1205	Gly	agc Ser	atc Ile	acc Thr	aaa Lys 1210	Gly	att Ile	ccc Pro	agc Ser	aca Thr 1215	Arg	3649
gtg Val	ccc Pro	tcg Ser	gac Asp 1220	Ser	gcc Ala	atc Ile	aca Thr	tac Tyr 1225	Arg	ggc Gly	tcc Ser	atc Ile	acc Thr 1230	His	ggc Gly	3697
acg Thr	cca Pro	gct Ala 1235	Asp	gtc Val	ctg Leu	tac Tyr	aag Lys 1240	Gly	acc Thr	atc Ile	acc Thr	agg Arg 124!	Ile	atc Ile	ggc Gly	3745
gag Glu	gac Asp 1250	Ser	ccg Pro	agt Ser	cgc Arg	ttg Leu 1255	Asp	cgc Arg	ggc	cgg Arg	gag Glu 1260	Asp	agc Ser	ctg Leu	ccc Pro	3793
aag Lys 126	Gly	cac His	gtc Val	atc Ile	tac Tyr 1270	Glu	ggc Gly	aag Lys	aag Lys	ggc Gly 1279	cac His	gtc Val	ttg Leu	tcc Ser	tat Tyr 1280	3841
gag Glu	ggt Gly	ggc Gly	atg Met	tct Ser 128	Val	acc Thr	cag Gln	tgc Cys	tcc Ser 129	Lys	gag Glu	gac Asp	ggc Gly	aga Arg 129	Ser	3889
agc Ser	tca Ser	gga Gly	pro	Pro	cat His	gag Glu	acg Thr	gcc Ala 130	Ala	ccc Pro	aag Lys	cgc Arg	acc Thr 131	Tyr	gac Asp	3937
atg Met	atg Met	gag Glu 131	Gly	cgc Arg	gtg Val	ggc	aga Arg 132	Ala	atc Ile	tcc Ser	tca Ser	gcc Ala 132	Ser	atc Ile	gaa Glu	3985
ggt Gly	ctc Leu 133	Met	ggc Gly	cgt Arg	gcc Ala	atc Ile 133!	Pro	ccg Pro	gag Glu	cga Arg	cac His 134	Ser	ccc Pro	cac His	cac His	4033
ctc Leu 134	Lys	gag Glu	cag Gln	cac His	cac His 135	Ile	cgc Arg	ggg	tcc Ser	atc Ile 135	aca Thr 5	caa Gln	ggg Gly	atc Ile	cct Pro 1360	4081
cgg Arg	tcc Ser	tac Tyr	gtg Val	gag Glu 136	Ala	cag Gln	gag Glu	gac Asp	tac Tyr 137	Leu	cgt Arg	cgg Arg	gag Glu	gcc Ala 137	Lys	4129
ctc Leu	cta Leu	aag Lys	cgg Arg 138	Glu	ggc Gly	acg Thr	cct Pro	ccg Pro 138	Pro	cca Pro	ccg Pro	Pro	tca Ser 139	Arg	gac Asp	4177
ctg Leu	acc Thr	gag Glu 139	Ala	tac Tyr	aag Lys	acg Thr	cag Gln 140	Ala	ctg Leu	ggc Gly	ccc	ctg Leu 140	Lys	ctg Leu	aag Lys	4225

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ccg Pro	gcc Ala 1410	His	gag Glu	ggc Gly	ctg Leu	gtg Val 1415	Ala	acg Thr	gtg Val	aag Lys	gag Glu 1420	Ala	ggc Gly	cgc Arg	tcc Ser	4273
atc Ile 1425	cat His	gag Glu	atc Ile	ccg Pro	cgc Arg 1430	Glu	gag Glu	ctg Leu	cgg Arg	cac His 1435	Thr	ccc Pro	gag Glu	ctg Leu	ccc Pro 1440	4321
ctg Leu	gcc Ala	ccg Pro	cgg Arg	ccg Pro 1445	Leu	aag Lys	gag Glu	ggc Gly	tcc Ser 1450	Ile	acg Thr	cag Gln	ggc Gly	acc Thr 1455	Pro	4369
ctc Leu	aag Lys	tac Tyr	gac Asp 1460	Thr	ggc Gly	gcg Ala	tcc Ser	acc Thr 1465	Thr	ggc Gly	tcc ser	aaa Lys	aag Lys 147	His	gac Asp	4417
gta Val	cgc Arg	tcc Ser 1475	Leu	atc Ile	ggc Gly	agc Ser	ccc Pro 1480	Gly	cgg Arg	acg Thr	ttc Phe	cca Pro 148	Pro	gtg Val	cac His	4465
ccg Pro	ctg Leu 1490	Asp	gtg Val	atg Met	gcc Ala	gac Asp 1495	Ala	cgg Arg	gca Ala	ctg Leu	gaa Glu 1500	Arg	gcc Ala	tgc Cys	tac Tyr	4513
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cgg Arg	cag Gln	agc Ser	ccc Pro 154	Leu	acc Thr	tat Tyr	gag Glu	gac Asp 154	His	ggg Gly	gca Ala	ccc Pro	ttt Phe 155	Ala	ggc Gly	4657
	ctc Leu		Arg					Thr					Thr			4705
ctg L e u	cag Gln 1570	Glu	ggc Gly	agc Ser	ctt Leu	tcg Ser 157	Ser	agc Ser	aag Lys	gca Ala	tcc Ser 158	Gln	gac Asp	cga Arg	aag Lys	4753
ctg Leu 158	acg Thr 5	tcg Ser	acg Thr	cct Pro	cgt Arg 159	Glu	atc Ile	gcc Ala	aag Lys	tcc Ser 159	Pro	cac His	agc Ser	acc Thr	gtg Val 1600	4801
ccc Pro	gag Glu	cac His	cac His	cca Pro 160	His	ccc Pro	atc Ile	tcg Ser	ccc Pro 161	Tyr	gag Glu	cac His	ctg Leu	ctt Leu 161	Arg	4849
ggc Gly	gtg Val	agt Ser	ggc Gly 162	Val	gac Asp	ctg Leu	tat Tyr	cgc Arg 162	Ser	cac His	atc Ile	ccc Pro	ctg Leu 163	Ala	ttc Phe	4897
gac Asp	ccc	acc Thr	Ser	ata Ile	ccc Pro	cgc Arg	ggc Gly 164	Ile	cct Pro	ctg Leu	gac Asp	gca Ala 164	Ala	gct Ala	gcc Ala	4945

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				Gly	cca Pro				Thr					Thr		5377
			Arg		cga Arg			Asp					Arg			5425
		Glu			atc Ile		Thr					Val				5473
	Ile				ggt Gly 183	Thr					Gly					5521
agc Ser	ggc Gly	gly ggg	ggt Gly	ggg Gly 184	ggc Gly 5	agc Ser	agc Ser	agc Ser	cgc Arg 185	Pro	gcc Ala	tcc Ser	cac His	tcc Ser 185	His	5569
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cag Gln	aga Arg	ccc Pro 187	Ser	gtg Val	ctt Leu	cac His	aac Asn 188	Thr	ggc Gly	atg Met	aag Lys	ggt Gly 188	Ile	atc Ile	acc Thr	5665

13										
gct gtg ga Ala Val Gl 1890	g ccc agc u Pro Ser	aag ccc Lys Pro 1895	Thr Val	ctg agg Leu Arg	tcc acc Ser Thr 1900	tcc a Ser T	cc tcc hr Ser	5713		
tca ccc gt Ser Pro Va 1905					Ala Thr			5761		
ctg ggc gg Leu Gly Gl	c acc ctc y Thr Leu 192	Asp Gly	gtc tac Val Tyr	cct acc Pro Thr 1930	ctc atg Leu Met	Glu P	cc gtc Pro Val .935	5809		
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gca gac ac Ala Asp Th 19	c ggc cat r Gly His 55	gcc ttc Ala Phe	ctc gcc Leu Ala 1960	aag ccc Lys Pro	cca gcc Pro Ala 196	Arg S	cc ggg Ser Gly	5905		
ctg gag co Leu Glu Pr 1970	c gcc tcc o Ala Ser	tcc ccc Ser Pro 1975	Ser Lys	ggc tcg Gly Ser	gag ccc Glu Pro 1980	cgg c Arg P	cc cta Pro Leu	5953		
gtg cct cc Val Pro Pr 1985	t gtc tct o Val Ser	ggc cac Gly His 1990	gcc acc Ala Thr	atc gcc Ile Ala 199	Arg Thr	cct g Pro A	gcg aag Ala Lys 2000	6001		
aac ctc go Asn Leu Al		His Ala				Pro P		6049		
teg gee te Ser Ala Se	g gac ccg r Asp Pro 2020	cac cgg His Arg	gaa aag Glu Lys 202	Thr Gln	agt aaa Ser Lys	Pro P 2030	tt tcc Phe Ser	6097		
atc cag ga Ile Gln Gl 20						Ser S		6145		
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acc cac ga Thr His As 2065	c aag ggg p Lys Gl	ctc ccc Leu Pro 2070	aag cac Lys His	ctg gaa Leu Glu 207	Glu Leu	gac a Asp I	aag agc Lys Ser 2080	6241		
cac ctg ga His Leu Gl	ig ggg gag u Gly Glu 208	ı Leu Arg	ccc aag Pro Lys	cag cca Gln Pro 2090	ggc ccc Gly Pro	Val I	aag ctt Lys Leu 2095	6289		
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cag ccc to Gln Pro Se 2:						Val I		6385		

14																
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cag Gln 2145	Asp					His					Ser					6481
gcc Ala					Phe					Cys					Leu	6529
cgc Arg				Ser					Pro					Gly		6577
ccg Pro			Gly		ccc Pro			Glu					Ser			6625
Pro	aac Asn 2210	Lys	acg Thr	tcg Ser	gtc Val	ttg Leu 2215	Gly	ggt Gly	ggt Gly	gag Glu	gac Asp 222	Gly	att Ile	gaa Glu	cct Pro	6673
gtg Val 2225	Ser	cca Pro	ccg Pro	gag Glu	ggc Gly 2230	Met	acg Thr	gag Glu	cca Pro	999 Gly 223	His	tcc Ser	cgg Arg	agt Ser	gct Ala 2240	6721
gtg Val	tac Tyr	ccg Pro	ctg Leu	ctg Leu 224!		cgg Arg	gat Asp	ggg Gly	gaa Glu 2250	Gln	acg Thr	gag Glu	ccc Pro	agc Ser 225	Arg	6769
				Ser	cca Pro				Ser					Phe		6817
			Thr		agc Ser			Ala					Lys			6865
		Asn			ctg Leu		Thr					Glu				6913
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					acc Thr 5					Ala					Ala	7009
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aaa Lys	tat Tyr	gac Asp 235	Gln	tgg Trp	gaa Glu	gag Glu	tcc Ser 236	Pro	ccg Pro	ctc Leu	agc Ser	gcc Ala 236	Asn	gct Ala	ttt Phe	7105

15	
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gcc tgg gac gag gag ccc aag cca ctg ctc tgc tcg cag tac gag aca Ala Trp Asp Glu Glu Pro Lys Pro Leu Leu Cys Ser Gln Tyr Glu Thr 2500 2505 2510	7537
ctc tcc gac agc gag tga ctcagaacag ggcggggggg ggcgggcggt Leu Ser Asp Ser Glu * 2515	7585
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<210> 5 <211> 2517

<212> PRT <213> Homo sapiens

435

Met Ser Gly Ser Thr Gln Leu Val Ala Gln Thr Trp Arg Ala Thr Glu Pro Arg Tyr Pro Pro His Ser Leu Ser Tyr Pro Val Gln Ile Ala Arg Thr His Thr Asp Val Gly Leu Leu Glu Tyr Gln His His Ser Arg Asp 40 Tyr Ala Ser His Leu Ser Pro Gly Ser Ile Ile Gln Pro Gln Arg Arg 55 Arg Pro Ser Leu Leu Ser Glu Phe Gln Pro Gly Asn Glu Arg Ser Gln 70 75 Glu Leu His Leu Arg Pro Glu Ser His Ser Tyr Leu Pro Glu Leu Gly 85 90 Lys Ser Glu Met Glu Phe Ile Glu Ser Lys Arg Pro Arg Leu Glu Leu 100 105 Leu Pro Asp Pro Leu Leu Arg Pro Ser Pro Leu Leu Ala Thr Gly Gln 120 Pro Ala Gly Ser Glu Asp Leu Thr Lys Asp Arg Ser Leu Thr Gly Lys 135 140 Leu Glu Pro Val Ser Pro Pro Ser Pro Pro His Thr Asp Pro Glu Leu 150 155 Glu Leu Val Pro Pro Arg Leu Ser Lys Glu Glu Leu Ile Gln Asn Met 170 Asp Arg Val Asp Arg Glu Ile Thr Met Val Glu Gln Gln Ile Ser Lys 185 Leu Lys Lys Lys Gln Gln Gln Leu Glu Glu Glu Ala Ala Lys Pro Pro 200 195 Glu Pro Glu Lys Pro Val Ser Pro Pro Pro Ile Glu Ser Lys His Arg 215 Ser Leu Val Gln Ile Ile Tyr Asp Glu Asn Arg Lys Lys Ala Glu Ala 230 235 Ala His Arg Ile Leu Glu Gly Leu Gly Pro Gln Val Glu Leu Pro Leu 245 250 Tyr Asn Gln Pro Ser Asp Thr Arg Gln Tyr His Glu Asn Ile Lys Ile 265 Asn Gln Ala Met Arg Lys Lys Leu Ile Leu Tyr Phe Lys Arg Arg Asn 280 His Ala Arg Lys Gln Trp Lys Gln Lys Phe Cys Gln Arg Tyr Asp Gln 295 300 Leu Met Glu Ala Leu Glu Lys Lys Val Glu Arg Ile Glu Asn Asn Pro 310 315 Arg Arg Arg Ala Lys Glu Ser Lys Val Arg Glu Tyr Tyr Glu Lys Gln 325 330 Phe Pro Glu Ile Arg Lys Gln Arg Glu Leu Gln Glu Arg Met Gln Ser 345 340 Arg Val Gly Gln Arg Gly Ser Gly Leu Ser Met Ser Ala Ala Arg Ser 360 Glu His Glu Val Ser Glu Ile Ile Asp Gly Leu Ser Glu Gln Glu Asn 375 380 Leu Glu Lys Gln Met Arg Gln Leu Ala Val Ile Pro Pro Met Leu Tyr 390 395 Asp Ala Asp Gln Gln Arq Ile Lys Phe Ile Asn Met Asn Gly Leu Met 405 410 Ala Asp Pro Met Lys Val Tyr Lys Asp Arg Gln Val Met Asn Met Trp 425 Ser Glu Gln Glu Lys Glu Thr Phe Arg Glu Lys Phe Met Gln His Pro

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Pro Ala His Glu Gly Leu Val Ala Thr Val Lys Glu Ala Gly Arg Ser 1410 1415 1420 Ile His Glu Ile Pro Arg Glu Glu Leu Arg His Thr Pro Glu Leu Pro

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Arg Gln Ser Pro Leu Thr Tyr Glu Asp His Gly Ala Pro Phe Ala Gly 1540 1550

His Leu Pro Arg Gly Ser Pro Val Thr Met Arg Glu Pro Thr Pro Arg 1555 1560 Leu Gln Glu Gly Ser Leu Ser Ser Ser Lys Ala Ser Gln Asp Arg Lys

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Tyr Pro Pro Tyr Leu Ile Arg Gly Tyr Pro Asp Thr Ala Ala Leu Glu 1665 $1670\,$ 1675 $1675\,$ Asn Arg Gln Thr Ile Ile Asn Asp Tyr Ile Thr Ser Gln Gln Met His

Asn Arg Gin Thr lie lie Asn Asp Tyr lie Thr Ser Gin Gin Met His 1685 1695 His Asn Thr Ala Thr Ala Met Ala Gin Arg Ala Asp Met Leu Arg Gly

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Pro Ile Trp Arg Pro Gly Thr Glu Gln Ser Ser Gly Ser Ser Gly Ser 1825 1830 1835

Ser Gly Gly Gly Gly Ser Ser Ser Arg Pro Ala Ser His Se

Ala His Gln His Ser Pro Ile Ser Pro Arg Thr Gln Asp Ala Leu Gln
1860 1865 1870

Gln Arg Pro Ser Val Leu His Asn Thr Gly Met Lys Gly Ile Ile Thr 1875 1880 1885

- Ala Val Glu Pro Ser Lys Pro Thr Val Leu Arg Ser Thr Ser Thr Ser 1895 1900
- Ser Pro Val Arq Pro Ala Ala Thr Phe Pro Pro Ala Thr His Cys Pro 1910 1915 Leu Gly Gly Thr Leu Asp Gly Val Tyr Pro Thr Leu Met Glu Pro Val
- 1930 1935 1925 Leu Leu Pro Lys Glu Ala Pro Arg Val Ala Arg Pro Glu Arg Pro Arg
- 1940 1945 1950 Ala Asp Thr Gly His Ala Phe Leu Ala Lys Pro Pro Ala Arg Ser Gly 1955 1960 1965
- Leu Glu Pro Ala Ser Ser Pro Ser Lys Gly Ser Glu Pro Arg Pro Leu
- 1970 1975 1980 Val Pro Pro Val Ser Gly His Ala Thr Ile Ala Arg Thr Pro Ala Lys
- 1990 1995 Asn Leu Ala Pro His His Ala Ser Pro Asp Pro Pro Ala Pro Pro Ala 2005 2010 2015
- Ser Ala Ser Asp Pro His Arg Glu Lys Thr Gln Ser Lys Pro Phe Ser 2020 2025 2030
- Ile Gln Glu Leu Glu Leu Arg Ser Leu Gly Tyr His Gly Ser Ser Tyr 2035 2040 2045
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- 2085 2090 2095 Gly Gly Glu Ala Ala His Leu Pro His Leu Arg Pro Leu Pro Glu Ser
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- Ala Pro Leu Tyr Ser Phe Pro Gly Ala Ser Cys Pro Val Leu Asp Leu 2165 2170 2175 Arg Arg Pro Pro Ser Asp Leu Tyr Leu Pro Pro Pro Asp His Gly Ala
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- 2290 2295 2300 Asn Ile Ser Gln Pro Gly Thr Glu Ile Phe Asn Met Pro Ala Ile Thr 2310 2315
- Gly Thr Gly Leu Met Thr Tyr Arg Ser Gln Ala Val Gln Glu His Ala 2325 2330
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	Thr					Glu					att Ile)					7420
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gca Ala	ggc Gly	agc Ser	999 Gly 244	Pro	cta Leu	gct Ala	ggt Gly	Pro 245	His	cac His	gcc Ala	tgg Trp	gat Asp 245	Glu	gag Glu	7996

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310 315 Arg Arg Arg Ala Lys Glu Ser Lys Val Arg Glu Tyr Tyr Glu Lys Gln 325 330 Phe Pro Glu Ile Arg Lys Gln Arg Glu Leu Gln Glu Arg Met Gln Ser 345 Arg Val Gly Gln Arg Gly Ser Gly Leu Ser Met Ser Ala Ala Arg Ser 360 Glu His Glu Val Ser Glu Ile Ile Asp Gly Leu Ser Glu Gln Glu Asn 375 Leu Glu Lys Gln Met Arg Gln Leu Ala Val Ile Pro Pro Met Leu Tyr 390 395 Asp Ala Asp Gln Gln Arg Ile Lys Phe Ile Asn Met Asn Gly Leu Met 405 410 Asp Asp Pro Met Lys Val Tyr Lys Asp Arg Gln Val Thr Asn Met Trp 420 425 Ser Glu Gln Glu Arg Asp Thr Phe Arg Glu Lys Phe Met Gln His Pro 440 Lys Asn Phe Gly Leu Ile Ala Ser Phe Leu Glu Arg Lys Thr Val Ala 455 Glu Cys Val Leu Tyr Tyr Tyr Leu Thr Lys Lys Asn Glu Asn Tyr Lys 470 475 Ser Leu Val Arg Arg Ser Tyr Arg Arg Gly Lys Ser Gln Gln Gln 485 490 Gln Gln Gln Gln Gln Gln Gln Gln Gln Met Ala Arg Ser Ser 505 Gln Glu Glu Lys Glu Lys Glu Lys Glu Lys Glu Ala Asp Lys Glu 520 Glu Glu Lys Gln Asp Ala Glu Asn Glu Lys Glu Glu Leu Ser Lys Glu 535 Lys Thr Asp Asp Thr Ser Gly Glu Asp Asn His Glu Lys Glu Ala Val 550 555 Ala Ser Lys Gly Arg Lys Thr Ala Asn Ser Gln Gly Arg Arg Lys Gly 570 Arg Ile Thr Arg Ser Met Ala Asn Glu Ala Asn His Glu Glu Thr Ala 585 Thr Pro Gln Gln Ser Ser Glu Leu Ala Ser Met Glu Met Asn Glu Ser 600 Ser Arg Trp Thr Glu Glu Glu Met Glu Thr Ala Lys Lys Gly Leu Leu 615 Glu His Gly Arg Asn Trp Ser Ala Ile Ala Arg Met Val Gly Ser Lys 630 635 Thr Val Ser Gln Cys Lys Asn Phe Tyr Phe Asn Tyr Lys Lys Arg Gln 650 645 Asn Leu Asp Glu Ile Leu Gln Gln His Lys Leu Lys Met Glu Lys Glu 665 660 Arg Asn Ala Arg Arg Lys Lys Lys Thr Pro Ala Ala Ala Ser Glu 680 Glu Thr Ala Phe Pro Pro Ala Ala Glu Asp Glu Glu Met Glu Ala Ser 695 700 Gly Ala Ser Ala Asn Glu Glu Glu Leu Ala Glu Glu Ala Glu Ala Ser 715 710 Gln Ala Ser Gly Asn Glu Val Pro Arg Val Gly Glu Cys Ser Gly Pro 730 725 Ala Ala Val Asn Asn Ser Ser Asp Thr Glu Ser Val Pro Ser Pro Arg 745 740 Ser Glu Ala Met Lys Asp Thr Gly Pro Lys Pro Thr Gly Thr Glu Ala 765 760 Leu Pro Ala Ala Thr Gln Pro Pro Val Pro Pro Pro Glu Glu Pro Ala 780 775 Val Ala Pro Ala Glu Pro Ser Pro Val Pro Asp Ala Ser Gly Pro Pro

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35 1270 1275 Tyr Asp Met Met Glu Gly Arg Val Gly Arg Thr Val Thr Ser Ala Ser 1290 1295 1285 Ile Glu Gly Leu Met Gly Arg Ala Ile Pro Glu Gln His Ser Pro His 1305 1310 Leu Lys Glu Gln His His Ile Arg Gly Ser Ile Thr Gln Gly Ile Pro 1320 1325 1315 Arg Ser Tyr Val Glu Ala Gln Glu Asp Tyr Leu Arg Arg Glu Ala Lys 1330 1340 Leu Leu Lys Arg Glu Gly Thr Pro Pro Pro Pro Pro Pro Pro Pro Arg Asp 1345 1350 1355 Leu Thr Glu Thr Tyr Lys Pro Arg Pro Leu Asp Pro Leu Gly Pro Leu 1365 1370 1375 Lys Leu Lys Pro Thr His Glu Gly Val Val Ala Thr Val Lys Glu Ala 1380 1385 1390 Gly Arg Ser Ile His Glu Ile Pro Arg Glu Glu Leu Arg Arg Thr Pro 1395 1400 1405 Glu Leu Pro Leu Ala Pro Arg Pro Leu Lys Glu Gly Ser Ile Thr Gln 1410 1415 1420 Gly Thr Pro Leu Lys Tyr Asp Ser Gly Ala Pro Ser Thr Gly Thr Lys 1425 1430 1435 1440 Lys His Asp Val Arg Ser Ile Ile Gly Ser Pro Gly Arg Pro Phe Pro 1445 1450 1455 Ala Leu His Pro Leu Asp Ile Met Ala Asp Ala Arg Ala Leu Glu Arg 1460 1465 1470 Ala Cys Tyr Glu Glu Ser Leu Lys Ser Arg Ser Gly Thr Ser Ser Gly 1475 1480 1485 Ala Gly Gly Ser Ile Thr Arg Gly Ala Pro Val Val Val Pro Glu Leu 1490 1495 1500 Gly Lys Pro Arg Gln Ser Pro Leu Thr Tyr Glu Asp His Gly Ala Pro 1505 1510 1515 Phe Thr Ser His Leu Pro Arg Gly Ser Pro Val Thr Thr Arg Glu Pro 1525 1530 Thr Pro Arg Leu Gln Glu Gly Ser Leu Leu Ser Ser Lys Ala Ser Gln 1540 1545 1550 Asp Arg Lys Leu Thr Ser Thr Pro Arg Glu Ile Ala Lys Ser Pro His 1555 1560 1565 Ser Thr Val Pro Glu His His Pro His Pro Ile Ser Pro Tyr Glu His 1570 1575 1580 Leu Leu Arg Gly Val Thr Gly Val Asp Leu Tyr Arg Gly His Ile Pro 1585 1590 1595 Leu Ala Phe Asp Pro Thr Ser Ile Pro Arg Gly Ile Pro Leu Glu Ala 1605 1610 1615 Ala Ala Ala Ala Tyr Tyr Leu Pro Arg His Leu Ala Pro Ser Pro Thr 1620 1625 Tyr Pro His Leu Tyr Pro Pro Tyr Leu Ile Arg Gly Tyr Pro Asp Thr 1635 1640 1645 Ala Ala Leu Glu Asn Arg Gln Thr Ile Ile Asn Asp Tyr Ile Thr Ser 1650 1655 1660 Gln Gln Met His His Asn Ala Ala Ser Ala Met Ala Gln Arg Ala Asp 1670 1675 Met Leu Arg Gly Leu Ser Pro Arg Glu Ser Ser Leu Ala Leu Asn Tyr 1685 1690 1695 Ala Ala Gly Pro Arg Gly Ile Ile Asp Leu Ser Gln Val Pro His Leu 1700 1705 1710 Pro Val Leu Val Pro Pro Thr Pro Gly Thr Pro Ala Thr Ala Ile Asp 1715 1720 1725 Arg Leu Ala Tyr Leu Pro Thr Ala Pro Pro Pro Phe Ser Ser Arg His 1735 1740 Ser Ser Pro Leu Ser Pro Gly Gly Pro Thr His Leu Ala Lys Pro

36 1750 1755 1745 Thr Ala Thr Ser Ser Ser Glu Arg Glu Arg Glu Arg Glu Arg Glu Arg 1765 1770 1775 Asp Lys Ser Ile Leu Thr Ser Thr Thr Thr Val Glu His Ala Pro Ile 1780 1785 Trp Arg Pro Gly Thr Glu Gln Ser Ser Gly Ala Gly Gly Ser Ser Arg 1795 1800 Pro Ala Ser His Thr His Gln His Ser Pro Ile Ser Pro Arq Thr Gln 1810 1815 1820 Asp Ala Leu Gln Gln Arg Pro Ser Val Leu His Asn Thr Ser Met Lys 1830 1835 1840 Gly Val Val Thr Ser Val Glu Pro Gly Thr Pro Thr Val Leu Arg Trp 1845 1850 1855 Ala Arg Ser Thr Ser Thr Ser Ser Pro Val Arg Pro Ala Ala Thr Phe 1860 1865 1870 Pro Pro Ala Thr His Cys Pro Leu Gly Gly Thr Leu Glu Gly Val Tyr 1875 1880 1885 Pro Thr Leu Met Glu Pro Val Leu Leu Pro Lys Glu Thr Ser Arg Val 1890 1895 1900 Ala Arg Pro Glu Arg Ala Arg Val Asp Ala Gly His Ala Phe Leu Thr 1910 1915 1920 Lys Pro Pro Gly Arg Glu Pro Ala Ser Ser Pro Ser Lys Ser Ser Glu 1925 1930 1935 Pro Arg Ser Leu Ala Pro Pro Ser Ser Ser His Thr Ala Ile Ala Arg 1940 1945 Thr Pro Ala Lys Asn Leu Ala Pro His His Ala Ser Pro Asp Pro Pro 1955 1960 1965 Ala Pro Thr Ser Ala Ser Asp Leu His Arg Glu Lys Thr Gln Ser Lys 1970 1975 1980 Pro Phe Ser Ile Gln Glu Leu Glu Leu Arg Ser Leu Gly Tyr His Ser 1985 1990 1995 Gly Ala Gly Tyr Ser Pro Asp Gly Val Glu Pro Ile Ser Pro Val Ser 2005 2010 2015 Ser Pro Ser Leu Thr His Asp Lys Gly Leu Ser Lys Pro Leu Glu Glu 2020 2025 2030 Leu Glu Lys Ser His Leu Glu Gly Glu Leu Arg His Lys Gln Pro Gly 2035 2040 2045 Pro Met Lys Leu Ser Ala Glu Ala Ala His Leu Pro His Leu Arg Pro 2050 2055 2060 Leu Pro Glu Ser Gln Pro Ser Ser Ser Pro Leu Leu Gln Thr Ala Pro 2070 2075 2080 Gly Ile Lys Gly His Gln Arg Val Val Thr Leu Ala Gln His Ile Ser 2085 2090 2095 Glu Val Ile Thr Gln Asp Tyr Thr Arg His His Pro Gln Gln Leu Ser 2105 2110 2100 Gly Pro Leu Pro Ala Pro Leu Tyr Ser Phe Pro Gly Ala Ser Cys Pro 2115 2120 2125 Val Leu Asp Leu Arg Arg Pro Pro Ser Asp Leu Tyr Leu Pro Pro Pro 2135 2140 Asp His Gly Thr Pro Ala Arg Gly Ser Pro His Ser Glu Gly Gly Lys 2150 2155 Arg Ser Pro Glu Pro Ser Lys Thr Ser Val Leu Gly Ser Ser Glu Asp 2165 2170 2175 Ala Ile Glu Pro Val Ser Pro Pro Glu Gly Met Thr Glu Pro Gly His 2180 2185 Ala Arg Ser Thr Ala Tyr Pro Leu Leu Tyr Arg Asp Gly Glu Gln Gly 2195 2200 2205

Glu Pro Arg Met Gly Leu Glu Ser Pro Gly Asn Thr Ser Gln Pro Pro

Thr Phe Phe Ser Lys Leu Thr Glu Ser Asn Ser Ala Met Val Lys Ser

2215

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2225					2230)				2235	5				2240	
Lys	Lys	Gln	Glu	Ile 2245		Lys	Lys	Leu	Asn 2250		His	Asn	Arg	Asn 2255		
Pro	Glu	Tyr	Asn 2260	Ile		Gln	Pro	Gly 2265	Thr		Ile	Phe	Asn 2270	Met		
Ala	Ile	Thr 2275	Gly		Gly	Leu	Met 2280	Thr		Arg	Ser	Gln 2285	Ala	Val	Gln	
Glu	His 2290	Ala		Thr	Asn	Met 2295	Gly		Glu	Ala	Ile 2300	Ile		Lys	Ala	
Leu 2305	Met		Lys	Tyr	Asp 2310	Gln		Glu	Glu	Pro 2315	Pro		Leu	Gly	Ala 2320	
		Phe	Asn		Leu		Ala	Ser	Ala 2330	Ser		Pro	Ala	Ala 2335	Ala	
Met	Pro	Tle	Thr	2325 Thr		Δen	Glv	Ara			His	Ala	Len	Thr		
			2340)		_		2345	5				2350	כ		
Pro	Gly	Gly 2355		GIY	Lys	Ala	Lys 2360		Ser	GIY	Arg	Pro 2365		Ser	Arg	
ГÀЗ	Ala 2370		Ser	Pro	Ala	Pro 2375		Leu	Ala	Ser	Gly 2380		Arg	Pro	Pro	
ser 2385	Val		Ser	Val	His 2390	Ser		Gly	Asp	Cys 2395		Arg	Arg	Thr	Pro 2400	
		Asn	Arg		${\tt Trp}$		Asp	Arg		Ser		Ala	Gly	Ser	Thr	
Pro	Phe	Pro				Leu	Ile				Gln	Ala		2415 Val		
Ala	Ser	Pro	2420 Pro		Pro	Gly	Leu	2425 Ala		Gly	Ser			Leu	Ala	
Gly	Pro	2435 His		Ala	Trp	Asp	2440 Glu		Pro	Lys	Pro	2445 Leu		Cys	Ser	
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															ctacca	360
CC a	tg t	ca c	gga t	cc a	aca o	cag o	cct o	gtg g	gca o	cag a	aca t	egg (egg 9	gct g Ala <i>l</i>	gct Na	407
r	1	er (3 T Y C	,61	5	3111 1	.10	val 1	ara (10			mg /	rand P	15	
nan	ccc	aaa	tac	cca	ccc	cat	aac	atc	tee	tac	cca	at.a	caq	ata	acc	455

gag ccc cgc tac cca ccc cat ggc atc tcc tac ccg gtg cag ata gcc Glu Pro Arg Tyr Pro Pro His Gly Ile Ser Tyr Pro Val Gln Ile Ala $20 \hspace{1cm} 25 \hspace{1cm} 30$

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		tac Tyr						935
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		tgg Trp						1031
		cct Pro						1079
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		aag Lys						1175

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atg Met	gag Glu 385	atg Met	aac Asn	gag Glu	agt Ser	tct Ser 390	cgc Arg	tgg Trp	act Thr	gag Glu	gaa Glu 395	gag Glu	atg Met	gag Glu	aca Thr	1:	559
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					gag Glu											1'	751
					gag Glu											1	799
					tca Ser 485											1:	847
					tca Ser											1	895

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						agc Ser			1943
						gac Asp			1991
						cag Gln 555			2039
						ccc Pro			2087
						cat His			2135
						ccc Pro			2183
						ctg Leu			2231
						tct Ser 635			2279
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						gtg Val			2375
						gtg Val			2423
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						ggc Gly 715			2519
						ccc Pro			2567
						aag Lys			2615

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					ccc Pro 965											3287
					cac His											3335

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		Asp					Gly					gag Glu)			3431
	His					Glu					Val	tca Ser			3479
Lys					Ser					Pro		gag Glu			3527
				Tyr					Gly			ggc Gly		Thr	3575
			Ser					Met				atc Ile 108	Pro		3623
		Pro					Gln					ggc Gly			3671
	Gly					Tyr					Glu	gac Asp			3719
Arg					Leu					Thr		cca Pro			3767
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			Leu					Thr				gtg Val 116	Val		3863
		Glu					Ile					aga Arg			3911
	Arg					Pro					Pro	ctg Leu			3959
Ser					Thr					Asp		gly ggg			4007
				Lys					Ser			ggc Gly		Pro	4055

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			gag Glu)					Glu					Ser			4151
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	Val		gaa Glu			Lys					${\tt Pro}$					4247
			gca Ala		Phe					Pro					Val	4295
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			ccc Pro					Leu					Leu			4631
ggc Gly	tac Tyr 142	Pro	gac Asp	acg Thr	gcg Ala	gcc Ala 143	Leu	gag Glu	aac Asn	cgc Arg	cag Gln 143	Thr	atc Ile	atc Ile	aat Asn	4679
gac Asp 144	Tyr	atc Ile	acc Thr	tcg Ser	cag Gln 144	Gln	atg Met	cac His	cac His	aac Asn 145	Ala	gcc Ala	tcc Ser	gcc Ala	atg Met 1455	4727
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	cgc cac agt Arg His Ser 152	Ser Ser					4967
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	ccc atc tgg Pro Ile Trp 0		Gly Thr				5111
ggg ggc agc Gly Gly Ser 1585	agc cgc ccc Ser Arg Pro	gcc tcc Ala Ser 1590	cac acc His Thr	cac cag His Gln 159	His Ser	ccc atc Pro Ile	5159
	acc cag gac Thr Gln Asp 160	Ala Leu					5207
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	agg tgg gcc Arg Trp Ala 1635					Val Arg	5303
cca gct gcc Pro Ala Ala 165	aca ttc cca Thr Phe Pro 0	cct gcc Pro Ala 1655	Thr His	tgc cca Cys Pro	ctt ggt Leu Gly 1660	ggc acc Gly Thr	5351
	gtc tac cct Val Tyr Pro				Leu Leu		5399
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	ccg ccg gcg Pro Pro Ala					5639
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	gtg agc tcc Val Ser Ser 1795		Thr His		Leu Ser	5783
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cac aag cag His Lys Gln 1825						5879
	cgg cca ctg Arg Pro Leu 184	Pro Glu Ser		Ser Ser Ser		5927
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gct cag cac Ala Gln His	atc agc gag Ile Ser Glu 1875		Gln Asp		His His	6023
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	ccc ccc gac Pro Pro Asp 192	His Gly Thr		Arg Gly Ser		6167
	ggc aaa agg Gly Lys Arg 1940					6215

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			Gly					Thr					Leu	tat Tyr		6311
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	Ser					Phe					Thr			aac Asn		6407
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	Āsp					Val					Ser			gac Asp		6887
					Leu					Trp				ccc Pro 2190	Ser	6935

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	ggg Gly 222	Pro					His					Glu				7079
	ctg Leu 10					Tyr					Asp					7124
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	.3> Mi 0> 9	ıs mi	ıscu.	lus												
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Pro	Arg	Tyr	Pro 20	Pro	His	Gly	Ile	Ser 25		Pro	Val	Gln	Ile 30	Ala	Arg	
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Glr	Arg	Tyr	Asp	Gln 85	Leu	Met	Glu	Ala	Trp	Glu	Lys	Lys	Val	Glu 95	Arg	
Il€	Glu	Asn	Asn 100	Pro	Arg	Arg	Arg	Ala 105	Lys	Glu	Ser	Lys	Val	Arg	Glu	
Tyr	Tyr	Glu 115	Lys	Gln	Phe	Pro	Glu 120	Ile	Arg	Lys	Gln	Arg 125	Glu	Leu	Gln	
Glu	Arg	Met	Gln	Ser	Arg	Val 135	Gly	Gln	Arg	Gly	Ser 140	Gly	Leu	Ser	Met	
Ser 145	Ala	Ala	Arg	Ser	Glu 150		Glu	Val	Ser	Glu 155		Ile	Asp	Gly	Leu 160	
	Glu	Gln	Glu	Asn 165		Glu	Lys	Gln	Met 170		Gln	Leu	Ala	Val 175		
Pro	Pro	Met	Leu 180		Asp	Ala	Asp	Gln 185		Arg	Ile	Lys	Phe		Asn	
Met	Asn	Gly 195		Met	Asp	Asp	Pro 200		Lys	Val	Tyr	Lys 205		Arg	Gln	
Val	Thr 210		Met	Trp	Ser	Glu 215		Glu	Arg	Asp	Thr 220		Arg	Glu	Lys	
Phe 225	Met	Gln	His	Pro	Lys 230		Phe	Gly	Leu	Ile 235		Ser	Phe	Leu	Glu 240	
	Lys	Thr	Val	Ala		Cys	Val	Leu	Tyr		Tyr	Leu	Thr	Lys		

245 250 Asn Glu Asn Tyr Lys Ser Leu Val Arg Arg Ser Tyr Arg Arg Gly 260 265 Met Ala Arg Ser Ser Gln Glu Glu Lys Glu Glu Lys Glu Lys Glu Lys 295 300 Glu Ala Asp Lys Glu Glu Glu Lys Gln Asp Ala Glu Asn Glu Lys Glu 310 315 Glu Leu Ser Lys Glu Lys Thr Asp Asp Thr Ser Gly Glu Asp Asn Asp 330 Glu Lys Glu Ala Val Ala Ser Lys Gly Arg Lys Thr Ala Asn Ser Gln 345 Gly Arg Arg Lys Gly Arg Ile Thr Arg Ser Met Ala Asn Glu Ala Asn 360 365 His Glu Glu Thr Ala Thr Pro Gln Gln Ser Ser Glu Leu Ala Ser Met 370 375 380 Glu Met Asn Glu Ser Ser Arg Trp Thr Glu Glu Glu Met Glu Thr Ala 390 395 Lys Lys Gly Leu Leu Glu His Gly Arg Asn Trp Ser Ala Ile Ala Arg 405 410 Met Val Gly Ser Lys Thr Val Ser Gln Cys Lys Asn Phe Tyr Phe Asn 420 425 Tyr Lys Lys Arg Gln Asn Leu Asp Glu Ile Leu Gln Gln His Lys Leu 435 440 Lys Met Glu Lys Glu Arg Asn Ala Arg Arg Lys Lys Lys Thr Pro 455 Ala Ala Ala Ser Glu Glu Thr Ala Phe Pro Pro Ala Ala Glu Asp Glu 470 475 Glu Met Glu Ala Ser Gly Ala Ser Ala Asn Glu Glu Glu Leu Ala Glu 485 490 Glu Ala Glu Ala Ser Gln Ala Ser Gly Asn Glu Val Pro Arg Val Gly 505 Glu Cys Ser Gly Pro Ala Ala Val Asn Asn Ser Ser Asp Thr Glu Ser 520 Val Pro Ser Pro Arg Ser Glu Ala Thr Lys Asp Thr Gly Pro Lys Pro 535 Thr Gly Thr Glu Ala Leu Pro Ala Ala Thr Gln Pro Pro Val Pro Pro 550 555 Pro Glu Glu Pro Ala Val Ala Pro Ala Glu Pro Ser Pro Val Pro Asp 565 570 Ala Ser Gly Pro Pro Ser Pro Glu Pro Ser His His Leu Pro His Pro 585 Arg Leu Leu Trp Thr Arg Met Asn Lys Lys Pro Arg Leu Leu Gln Leu 600 Pro Arg Gln Arg Met Pro Arg Ser Arg Ser Leu Arg Pro Arg Arg Ser 615 620 Met Trp Glu Lys Pro Glu Glu Pro Glu Ala Ser Glu Lys Pro Pro Lys 630 635 Ser Val Lys Ser Asp His Lys Lys Glu Thr Glu Glu Glu Pro Glu Asp 645 650 Lys Ala Lys Gly Thr Glu Ala Ile Glu Thr Val Ser Glu Ala Pro Leu 665 Lys Val Glu Lys Ala Gly Ser Lys Ala Ala Val Thr Lys Gly Ser Ser 680 Ser Gly Ala Thr Gln Asp Ser Asp Ser Ser Ala Thr Cys Ser Ala Asp 695 700 Glu Val Asp Glu Pro Glu Gly Gly Asp Lys Gly Arg Leu Leu Ser Pro 710 715 Arg Pro Ser Leu Leu Thr Pro Ala Gly Asp Pro Arg Ala Ser Thr Ser

725 730 Pro Gln Lys Pro Leu Asp Leu Lys Gln Leu Lys Gln Arg Ala Ala Ala 745 740 Ile Pro Pro Ile Val Thr Lys Val His Glu Pro Pro Arg Glu Asp Thr 760 Val Pro Pro Lys Pro Val Pro Pro Val Pro Pro Pro Thr Gln His Leu 775 Gln Pro Glu Gly Asp Val Ser Gln Gln Ser Gly Gly Ser Pro Arg Gly 790 795 Lys Ser Arg Ser Pro Val Pro Pro Ala Glu Lys Glu Ala Glu Lys Pro 805 810 815 Ala Phe Phe Pro Ala Phe Pro Thr Glu Gly Pro Lys Leu Pro Thr Glu 825 830 Pro Pro Arg Trp Ser Ser Gly Leu Pro Phe Pro Ile Pro Pro Arg Glu 840 845 Val Ile Lys Thr Ser Pro His Ala Ala Asp Pro Ser Ala Phe Ser Tyr 855 860 Thr Pro Pro Gly His Pro Leu Pro Leu Gly Leu His Asp Ser Ala Arg 870 875 Pro Val Leu Pro Arg Pro Pro Ile Ser Asn Pro Pro Pro Leu Ile Ser 885 890 Ser Ala Lys His Pro Gly Val Leu Glu Arg Gln Leu Gly Ala Ile Ser 905 910 900 Gln Gln Gly Met Ser Val Gln Leu Arg Val Pro His Ser Glu His Ala 920 Lys Ala Pro Met Gly Pro Leu Thr Met Gly Leu Pro Leu Ala Val Asp 935 Pro Lys Lys Leu Gly Thr Ala Leu Gly Ser Ala Thr Ser Gly Ser Ile 950 955 Thr Lys Gly Leu Pro Ser Thr Arg Ala Ala Asp Gly Pro Ser Tyr Arg 970 Gly Ser Ile Thr His Gly Thr Pro Ala Asp Val Leu Tyr Lys Gly Thr 985 Ile Ser Arg Ile Val Gly Glu Asp Ser Pro Ser Arg Leu Asp Arg Ala 1000 Arg Glu Asp Thr Leu Pro Lys Gly His Val Ile Tyr Glu Gly Lys Lys 1010 1015 1020 Gly His Val Leu Ser Tyr Glu Gly Gly Met Ser Val Ser Gln Cys Ser 1030 1035 Lys Glu Asp Gly Arg Ser Ser Ser Gly Pro Pro His Glu Thr Ala Ala 1045 1050 Pro Lys Arg Thr Tyr Asp Met Met Glu Gly Arg Val Gly Arg Thr Val 1060 1065 Thr Ser Ala Ser Ile Glu Gly Leu Met Gly Arg Ala Ile Pro Glu Gln 1080 1085 His Ser Pro His Leu Lys Glu Gln His His Ile Arg Gly Ser Ile Thr 1090 1095 1100 Gln Gly Ile Pro Arg Ser Tyr Val Glu Ala Gln Glu Asp Tyr Leu Arg 1110 1115 Arg Glu Ala Lys Leu Leu Lys Arg Glu Gly Thr Pro Pro Pro Pro Pro 1125 1130 1135 Pro Pro Arg Asp Leu Thr Glu Thr Tyr Lys Pro Arg Pro Leu Asp Pro 1140 1145 Leu Gly Pro Leu Lys Leu Lys Pro Thr His Glu Gly Val Val Ala Thr 1155 1160 1165 Val Lys Glu Ala Gly Arg Ser Ile His Glu Ile Pro Arg Glu Glu Leu 1170 1175 1180 Arg Arg Thr Pro Glu Leu Pro Leu Ala Pro Arg Pro Leu Lys Glu Gly 1190 1195 Ser Ile Thr Gln Gly Thr Pro Leu Lys Tyr Asp Ser Gly Ala Pro Ser 1205 1210 1

Thr Gly Thr Lys Lys His Asp Val Arg Ser Ile Ile Gly Ser Pro Gly 1220 1225 1230

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Ala Leu Glu Arg Ala Cys Tyr Glu Glu Ser Leu Lys Ser Arg Ser Gly 1250 1255 1260

Thr Ser Ser Gly Ala Gly Gly Ser Ile Thr Arg Gly Ala Pro Val Val 1265 1270 1275 1280 Val Pro Glu Leu Gly Lys Pro Arg Gln Ser Pro Leu Thr Tyr Glu Arg Gly 1295 1290 1295

His Gly Ala Pro Phe Thr Ser His Leu Pro Arg Gly Ser Pro Val Thr \$1300\$ \$1310

Thr Arg Glu Pro Thr Pro Arg Leu Gln Glu Gly Ser Leu Leu Ser Ser

Lys Ala Ser Gln Asp Arg Lys Leu Thr Ser Thr Pro Arg Glu Ile Ala 1330 Lys Ser Pro His Ser Thr Val Pro Glu His His Pro His Pro Ile Ser

1345 1350 1350 1355 1360 Pro Tyr Glu His Leu Leu Arg Gly Val Thr Gly Val Asp Leu Tyr Arg 1365 1370 1375

Gly His Ile Pro Leu Ala Phe Asp Pro Thr Ser Ile Pro Arg Gly Ile 1380 1385 1390

Pro Leu Glu Ala Ala Ala Ala Ala Tyr Tyr Leu Pro Arg His Leu Ala 1395 Pro Ser Pro Thr Tyr Pro His Leu Tyr Pro Pro Tyr Leu Ile Arg Gly

1410 1415

Tyr Pro Asp Thr Ala Ala Leu Glu Asn Arg Gln Thr Ile Ile Asn Asp 1425

1430

1430

1430

Tyr Ile Thr Ser Gln Gln Met His His Asn Ala Ala Ser Ala Met Ala 1445 1450 1455

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Leu Ala Lys Pro Thr Ala Thr Ser Ser Ser Glu Arg Glu Arg Glu Arg
1540 1545 1550

Glu Arg Glu Arg Asp Lys Ser Ile Leu Thr Ser Thr Thr Thr Val Glu 1555 1560 1565

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Thr Ser Met Lys Gly Val Val Thr Ser Val Glu Pro Gly Thr Pro Thr 1620 1625 1630

Val Leu Arg Trp Ala Arg Ser Thr Ser Thr Ser Ser Pro Val Arg Pro 1635 1640 1645

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Glu Gly Val Tyr Pro Thr Leu Met Glu Pro Val Leu Leu Pro Lys Glu 1665 1670 1675 168

Thr Ser Arg Val Ala Arg Pro Glu Arg Ala Arg Val Asp Ala Gly His

1685 1690 169

Ala Phe Leu Thr Lys Pro Pro Gly Arg Glu Pro Ala Ser Ser Pro Ser 1700 1705 1710

Lys Ser Ser Glu Pro Arg Ser Leu Ala Pro Pro Ser Ser Ser His Thr 1715 1720 1725

Ala Ile Ala Arg Thr Pro Ala Lys Asn Leu Ala Pro His His Ala Ser 1730 1730 Pro Asp Pro Pro Ala Pro Thr Ser Ala Ser Asp Leu His Arg Glu Lys

1745 1750 1755 1760 Thr Gln Ser Lys Pro Phe Ser Ile Gln Glu Leu Glu Leu Arg Ser Leu 1765 1770 1775

Gly Tyr His Ser Gly Ala Gly Tyr Ser Pro Asp Gly Val Glu Pro Ile 1780 1780 1785 Ser Pro Val Ser Ser Pro Ser Leu Thr His Asp Lys Gly Leu Ser Lys

1795 1800 1805 Pro Leu Glu Leu Glu Lys Ser His Leu Glu Gly Glu Leu Arg His

1810 1815 1820 Lys Gln Pro Gly Pro Met Lys Leu Ser Ala Glu Ala Ala His Leu Pro

1825 1830 1835 1840 His Leu Arg Pro Leu Pro Glu Ser Gln Pro Ser Ser Pro Leu Leu 1845 1850 1850

Gln Thr Ala Pro Gly Ile Lys Gly His Gln Arg Val Val Thr Leu Ala 1860 1865 1870

Gln His Ile Ser Glu Val Ile Thr Gln Asp Tyr Thr Arg His His Pro 1875 \$1880\$

Gln Leu Ser Gly Pro Leu Pro Ala Pro Leu Tyr Ser Phe Pro Gly 1890 1890 Ala Ser Cys Pro Val Leu Aep Leu Arg Arg Pro Pro Ser Asp Leu Tyr

1905 1910 1915 1920 Leu Pro Pro Pro Asp His Gly Thr Pro Ala Arg Gly Ser Pro His Ser 1925 1930 1930

Glu Gly Gly Lys Arg Ser Pro Glu Pro Ser Lys Thr Ser Val Leu Gly 1945 1940 1945 Ser Ser Glu Asp Ala Ile Glu Pro Val Ser Pro Pro Glu Glv Met Thr

1955 1960 1965 Glu Pro Gly His Ala Arg Ser Thr Ala Tyr Pro Leu Leu Tyr Arg Asp

1970 1975 1980 1975 1980 1976 Gly Glu Gln Gly Glu Pro Arg Met Gly Leu Glu Ser Pro Gly Asn Thr 1985 1990 1995 2000 Ser Gln Pro Pro Thr Phe Phe Ser Lys Leu Thr Glu Ser Asn Ser Ala

2005 2010 2015 Met Val Lys Ser Lys Lys Gln Glu Ile Asn Lys Lys Leu Asn Thr His 2020 2025 2030

Asn Arg Asn Glu Pro Glu Tyr Asn Ile Gly Gln Pro Gly Thr Glu Ile 2035 2040 2045

Phe Asn Met Pro Ala Ile Thr Gly Ala Gly Leu Met Thr Cys Arg Ser 2050 2055 2060

Gln Ala Val Gln Glu His Ala Ser Thr Asn Met Gly Leu Glu Ala Ile 2065 2070 2075 2080

Ile Arg Lys Ala Leu Met Gly Lys Tyr Asp Gln Trp Glu Glu Pro Pro 2095 2090 2095 Pro Leu Gly Ala Asn Ala Phe Asn Pro Leu Asn Ala Ser Ala Ser Leu

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2130 2135 2140

Pro Ser Ser Arg Lys Ala Lys Ser Pro Ala Pro Gly Leu Ala Ser Gly 2145 2150 2155 2156 2156 216 Asp Arg Pro Pro Ser Val Ser Ser Val His Ser Glu Gly Asp Cys Asn

52 2165 2170 Arg Arg Thr Pro Leu Thr Asn Arg Val Trp Glu Asp Arg Pro Ser Ser 2180 2185 Ala Gly Ser Thr Pro Phe Pro Tyr Asn Pro Leu Ile Met Arg Leu Gln 2195 2200 Ala Gly Val Met Ala Ser Pro Pro Pro Pro Gly Leu Ala Ala Gly Ser 2215 2220 Gly Pro Leu Ala Gly Pro His His Ala Trp Asp Glu Glu Pro Lys Pro 2225 2230 2235 Leu Leu Cys Ser Gln Tyr Glu Thr Leu Ser Asp Ser Glu 2245 <210> 10 <211> 7940 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (241) ... (7563) <400> 10 ccaagatggc ggccaaggtg gcgaagcagc agccgcggcg gcggcggcgg ctggagtgag egteegacte geegegeega acgaggteee ggtgtaggge egegegeegt ggeegegtee 120 cactecteag geogggege acgteggete ceaegettag ceageteeeg gtggttteet 180 agaaacatga ttgtttattg gcattgatct cacagtctgg tgaggacttc tttactgata 240 atg tca agt tca ggt tat cct ccc aac caa gga gca ttc agc aca gaa 288 Met Ser Ser Gly Tyr Pro Pro Asn Gln Gly Ala Phe Ser Thr Glu caa agt cgt tat cct cct cac tct gtc cag tat aca ttt ccc aac acc 336 Gln Ser Arg Tyr Pro Pro His Ser Val Gln Tyr Thr Phe Pro Asn Thr 20 cgc cac cag cag gag ttc gca gtc cct gat tat cgt tcc tct cat ctt 384 Arg His Gln Glu Phe Ala Val Pro Asp Tyr Arg Ser Ser His Leu gaa gtg agt cag gca tca cag ctt ttg cag caa cag cag cag caa cag 432 Glu Val Ser Gln Ala Ser Gln Leu Leu Gln Gln Gln Gln Gln Gln Gln Gln ctt cga agg cga cct tcc ttg ctt tca qaa ttt cac cca ggt tct qac 480 Leu Arg Arg Pro Ser Leu Leu Ser Glu Phe His Pro Gly Ser Asp 70 agg cct caa gaa agg aga act agt tat gaa ccq ttt cat cca ggc cca 528 Arg Pro Gln Glu Arg Arg Thr Ser Tyr Glu Pro Phe His Pro Gly Pro tcc cca gtg gat cat gat tca ctg gaa tcg aag cga cca cgt ctg gaa 576 Ser Pro Val Asp His Asp Ser Leu Glu Ser Lys Arg Pro Arg Leu Glu 105 caq qtt tct qat tct cat ttt caq cqt qtc aqt qct qcq qtt ttq cct 624 Gln Val Ser Asp Ser His Phe Gln Arg Val Ser Ala Ala Val Leu Pro 120 tta gtg cac ccg ctg cca gaa ggg ctg agg gct tct gca gat gct aag 672 Leu Val His Pro Leu Pro Glu Gly Leu Arg Ala Ser Ala Asp Ala Lys

nesys: ozton

			ggc Gly					720
			gat Asp					768
			cag Gln					816
			atc Ile					864
			aaa Lys 215					912
			aaa Lys					960
			gca Ala					1008
			ctg Leu					1056
			atc Ile					1104
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			tat Tyr					1200
			aat Asn					1248
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			ttt Phe					1344
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gat ggg ctc tct gag cag gag aat aat gag aaa caa atg cgg cag ctc 1440

NOSSEZEE DEIDO

Asp 385	Gly	Leu	Ser	Glu	Glu	Asn	Asn	Glu	Lys 395	Gln	Met	Arg	Gln	Leu 400	
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				atg Met											1584
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				aag Lys											1680
				gag Glu 485											1728
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				aaa Lys											1872
				aag Lys											1920
				gag Glu 565											1968
				cgt Arg											2016
				agt Ser											2064
				ccg Pro											2112

acc Thr 625	tct Ser	cga Arg	tgg Trp	aca Thr	gaa Glu 630	gaa Glu	gaa Glu	atg Met	gaa Glu	gtt Val 635	gct Ala	aaa Lys	aaa Lys	ggt Gly	cta Leu 640	2160
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														gaa Glu		2400
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														tta Leu		2544
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					att Ile 950											3120
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		Pro			ttg Leu		Thr					Gly				3312
	Thr				acc Thr 1030	Arg					Leu					3360
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			Gln		aca Thr			Pro					Ile			3504
					cag Gln											3552

	Lys					Ser					Asn			cct Pro		3600
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att Ile	tca Ser	gtg Val 1155	Glu	agc Ser	att Ile	cca Pro	tcc Ser 1160	Leu	cgg Arg	ggc Gly	tct Ser	atc Ile 1165	Thr	cag Gln	ggc Gly	3744
		Ala					Gly					Ala		gtg Val		3792
	Ser					Pro					ser			aaa Lys		3840
aga Arg	gag Glu	gaa Glu	gct Ala	gca Ala 1205	Ser	aaa Lys	ggc Gly	cat His	gtt Val 1210	Ile	tat Tyr	gaa Glu	ggc Gly	aaa Lys 121	Ser	3888
gga Gly	cat His	atc Ile	ttg Leu 1220	ser	tat Tyr	gat Asp	aat Asn	att Ile 1225	Lys	aat Asn	gcc Ala	cga Arg	gaa Glu 123	gly aaa	act Thr	3936
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gaa Glu	ggt Gly	gcc Ala	att Ile	acc Thr	aaa Lys	gga Gly	aaa Lys	cca Pro	tat Tyr	gat Asp	ggc Gly	atc Ile	acc Thr	acc Thr	atc Ile	4272

IGENEYES DESIDE

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	att Ile			Ser					Thr					Asp		4416
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gag Glu 1429	aac Asn	ata Ile	aaa Lys	gtg Val	gta Val 1430	Glu	cgg Arg	gga Gly	aaa Lys	tat Tyr 1435	Glu	gat Asp	gtg Val	aaa Lys	gca Ala 1440	4560
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tcc Ser	gtt Val	ctt Leu	agg Arg 1460	Ser	aca Thr	ctg Leu	cat His	gaa Glu 1469	Ala	ccc Pro	aaa Lys	gca Ala	caa Gln 1470	Leu	agc Ser	4656
cct Pro	Gly ggg	att Ile 1479	Tyr	gat Asp	gac Asp	acc Thr	agt Ser 1480	Ala	cgg Arg	agg Arg	acc Thr	cct Pro 1485	Val	agt Ser	tat Tyr	4704
caa Gln	aac Asn 1490	Thr	atg Met	tcc Ser	aga Arg	ggc Gly 149	Ser	ccc Pro	atg Met	atg Met	aac Asn 1500	Arg	act Thr	tct Ser	gat Asp	4752
gtt Val 150	aca Thr	att Ile	cct Pro	cct Pro	aac Asn 151	Lys	tct Ser	acc Thr	aat Asn	cat His 151	Glu	agg Arg	aaa Lys	tcg Ser	aca Thr 1520	4800
ctg Leu	acc Thr	cct Pro	acc Thr	cag Gln 152	Arg	gaa Glu	agt Ser	atc Ile	cca Pro 153	Ala	aag Lys	tct Ser	cca Pro	gtg Val 153	Pro	4848
Gly 999	gtg Val	gac Asp	cct Pro 154	Val	gtg Val	agc Ser	cac His	agt Ser 154	Pro	ttt Phe	gat Asp	ccc Pro	cat His 155	His	aga Arg	4896
ggc Gly	agc Ser	act Thr 155	Ala	ggc Gly	gag Glu	gtt Val	tat Tyr 156	Trp	agc Ser	cac His	ctg Leu	ecc Pro 156	Thr	caa Gln	ttg Leu	4944
gat Asp	cca Pro	gcc Ala	atg Met	cct Pro	ttt Phe	cac His	agg Arg	gct Ala	ttg Leu	gat Asp	cct Pro	gca Ala	gcg Ala	gct Ala	gct Ala	4992

tac ctg ttt cag aga cag ctt tca cca act cca ggt tac cca agt cag

Tyr Leu Phe Gln Arg Gln Leu Ser Pro Thr Pro Gly Tyr Pro Ser Gln

tat cag ctt tac gca atg gag aac aca aga cag aca atc tta aat gat Tyr Gln Leu Tyr Ala Met Glu Asn Thr Arg Gln Thr Ile Leu Asn Asp

						${\tt Gln}$					Asn				gat Asp 1630	Val		5136
					Ser					Pro					tac Tyr			5184
				Gly					Thr					Thr	att Ile			5232
	ma ud		His					Ser					Asp		atc Ile			5280
1000							Ile					Arg			aac Asn		Ala	5328
	y N	tcc Ser	atg Met	tct Ser	cca Pro 170	Gly	cac His	cca Pro	aca Thr	cac His 170	Leu	gca Ala	gct Ala	gct Ala	gca Ala 1710	Ser	gct Ala	5376
1		gag Glu	agg Arg	gaa Glu 171!	Arg	gaa Glu	cgg Arg	gag Glu	cgg Arg 1720	Glu	aag Lys	gag Glu	cgg Arg	gag Glu 172	cgg Arg	gaa Glu	cgg Arg	5424
-				Ala					Leu					Gly	tca Ser			5472
			Gly					His					Ser		tcc Ser			5520
							Thr					Arg			gtt Val		Gln	5568
						Thr					Pro				act Thr 179	Ala		5616
		cta Leu	cga Arg	atc Ile 179	Met	cca Pro	ctg Leu	cct Pro	gct Ala 180	Gly	ggc Gly	cct Pro	tca Ser	ata Ile 180	agc Ser	caa Gln	ggc Gly	5664
		ctg	cca	gcc	tcc										gct			5712

Leu Pro Ala Ser Arg Tyr Asn Thr Ala Ala Asp Ala Leu Ala Ala Leu

	Asp					Āla					gtg Val					5760
					Ala					Glu	aat Asn				Arg	5808
				Ser					Leu		cag Gln			Leu		5856
gtg Val	gag Glu	aag Lys 1875	Arg	tct Ser	gtt Val	cag Gln	tgt Cys 1880	Leu	tac Tyr	act Thr	tct Ser	tca Ser 1885	Ala	ttt Phe	cca Pro	5904
agt Ser	ggc Gly 1890	Lys	ccc Pro	cag Gln	cct Pro	cat His 1895	Ser	tca Ser	gta Val	gtt Val	tat Tyr 1900	Ser	gag Glu	gct Ala	gl ^A aaa	5952
aaa Lys 1909	Asp	aaa Lys	ggg Gly	cct Pro	cct Pro 1910	${\tt Pro}$	aaa Lys	tcc Ser	aga Arg	tat Tyr 1915	gag Glu	gaa Glu	gag Glu	cta Leu	agg Arg 1920	6000
					Thr					Asn	ttc Phe				Ile	6048
				Ile					Asp		agg Arg			Gly		6096
			Asp					Leu			cac His		Tyr			6144
cct Pro	agc Ser 197	Asp	gct Ala	att Ile	gag Glu	gtg Val 197	Ile	agt Ser	cct Pro	gcc Ala	agc Ser 1980	Ser	cct Pro	gcg Ala	cca Pro	6192
ccc Pro 198	Gln	gag Glu	aaa Lys	ctg Leu	cag Gln 199	Thr	tat Tyr	cag Gln	cca Pro	gag Glu 199	gtt Val	gtt Val	aag Lys	gca Ala	aat Asn 2000	6240
caa Gln	gcg Ala	gaa Glu	aat Asn	gat Asp 200	Pro	acc Thr	aga Arg	caa Gln	tat Tyr 201	Glu	gga Gly	cca Pro	tta Leu	cat His 201	His	6288
tat Tyr	cga Arg	cca Pro	cag Gln 202	Gln	gaa Glu	tca Ser	cca Pro	ser 202	Pro	caa Gln	caa Gln	cag Gln	ctg Leu 203	Pro	cct Pro	6336
tct Ser	tca Ser	cag Gln 203	Ala	gag Glu	gga Gly	atg Met	ggg Gly 204	Gln	gtg Val	ccc Pro	agg Arg	acc Thr 204	His	cgg Arg	ctg Leu	6384
											aca Thr					6432

aga aat caa gtt too tog cag act coc cag cag cot cot act tot aca

Arg 2065		Gln	Val	Ser	Ser 2070		Thr	Pro	Gln	Gln 2075	Pro	Pro	Thr	Ser	Thr 2080	
					tct Ser					Thr					Lys	6528
aca Thr	tca Ser	aac Asn	cgt Arg 2100	Tyr	agc Ser	cca Pro	gaa Glu	tcc Ser 2105	Gln	gct Ala	cag Gln	tct Ser	gtc Val 2110	His	cat His	6576
caa Gln	aga Arg	cca Pro 2115	Gly	tca Ser	agg Arg	gtc Val	tct Ser 2120	Pro	gaa Glu	aat Asn	ctt Leu	gtg Val 2125	Asp	aaa Lys	tcc Ser	6624
agg Arg	gga Gly 2130	Ser	agg Arg	cct Pro	gga Gly	aaa Lys 2135	Ser	cca Pro	gag Glu	agg Arg	agt Ser 2140	His	gtc Val	tct Ser	tcc Ser	6672
gag Glu 2145	Pro	tac Tyr	gag Glu	ccc Pro	atc Ile 2150	ser	cca Pro	ccc Pro	cag Gln	gtt Val 2155	Pro	gtt Val	gtg Val	cat His	gag Glu 2160	6720
aaa Lys	cag Gln	gac Asp	agc Ser	ttg Leu 2165	ctg Leu	ctc Leu	ttg Leu	tct Ser	cag Gln 2170	Arg	ggc Gly	gca Ala	gag Glu	cct Pro 217	Ala	6768
gag Glu	cag Gln	agg Arg	aat Asn 2180	Asp	gcc Ala	cgc Arg	tca Ser	cca Pro 218	Gly	agt Ser	ata Ile	agc Ser	tac Tyr 2190	Leu	cct Pro	6816
tca Ser	ttc Phe	ttc Phe 219	Thr	aag Lys	ctt Leu	gaa Glu	aat Asn 220	Thr	tca Ser	ccc Pro	atg Met	gtt Val 2209	Lys	tca Ser	aag Lys	6864
aag Lys	cag Gln 221	Glu	att Ile	ttt Phe	cgt Arg	aag Lys 221!	Leu	aac Asn	tcc Ser	tct Ser	ggt Gly 222	Gly	ggt Gly	gac Asp	tct Ser	6912
gat Asp 222	Met	gca Ala	gct Ala	gct Ala	cag Gln 2230	Pro	gga Gly	act Thr	gag Glu	atc Ile 223	Phe	aat Asn	ctg Leu	cca Pro	gca Ala 2240	6960
gtt Val	act Thr	acg Thr	tca Ser	ggc Gly 224	tca Ser 5	gtt Val	agc Ser	tct Ser	aga Arg 225	Gly	cat His	tct Ser	ttt Phe	gct Ala 225	Asp	7008
cct Pro	gcc Ala	agt Ser	aat Asn 226	Leu	gly ggg	ctg Leu	gaa Glu	gac Asp 226	Ile	atc Ile	agg Arg	aag Lys	gct Ala 227	Leu	atg Met	7056
gga Gly	agc Ser	ttt Phe 227	Asp	gac Asp	aaa Lys	gtt Val	gag Glu 228	Asp	cat His	gga Gly	gtt Val	gtc Val 228	Met	tcc Ser	cag Gln	7104
cct Pro	atg Met	gga Gly	gta Val	gtg Val	cct Pro	ggt Gly	act Thr	gcc Ala	aac Asn	acc Thr	tca Ser	gtt Val	gtg Val	acc Thr	agt Ser	7152

DSSSZYSK CSINO

ggt gag aca cga aga gag gaa ggg gac cca tca cct cat tca gga gga

Gly Glu Thr Arg Arg Glu Glu Gly Asp Pro Ser Pro His Ser Gly Gly

gtt tgc aaa cca aag ctg atc agc aag tca aac agc agg aaa tct aag

Val Cys Lys Pro Lys Leu Ile Ser Lys Ser Asn Ser Arg Lys Ser Lys

tct cct ata cct ggg caa ggc tac tta gga acg gaa cgg ccc tct tca Ser Pro Ile Pro Gly Gln Gly Tyr Leu Gly Thr Glu Arg Pro Ser Ser

				2340)				2345	5				2350)	
	gtc Val	tcc Ser	tct Ser 2355	Val	cat His	tca Ser	gaa Glu	ggg Gly 2360	Asp	tac Tyr	cat His	agg Arg	cag Gln 2365	Thr	cca Pro	Gly 999
	tgg Trp	gcc Ala 2370	Trp	gaa Glu	gac Asp	agg Arg	ccc Pro 237	Ser	tca Ser	aca Thr	ggc Gly	tca Ser 2380	Thr	cag Gln	ttt Phe	cct Pro
i out	tat Tyr 238	Asn	cct Pro	ctg Leu	act Thr	atg Met 239	Arg	atg Met	ctc Leu	agc Ser	agt Ser 2399	Thr	cca Pro	cca Pro	aca Thr	ccg Pro 2400
	att Ile	gca Ala	tgt Cys	gct Ala	CCC Pro 240	Ser	gcg Ala	gtg Val	aac Asn	caa Gln 241	Ala	gct Ala	cct Pro	cac His	caa Gln 241	Gln
in id	aac Asn	agg Arg	atc Ile	tgg Trp 242	gag Glu)	cga Arg	gag Glu	cct Pro	gcc Ala 242	Pro	ctg Leu	ctc Leu	tca Ser	gca Ala 243	Gln	tac Tyr
				Ser	gat Asp				*	act	gcac	aaa 🤉	gtga	gggg:	aa	
	aga ttg aaa	gccc aatg atac agtg	tcc atg agg	tgtg gaag caat agtt	actta ttca tcag	at to tt to tg go	ccct ggag acta ttac	gaga agtc taat atct	tti a aai a ati g tai	ttca tggg agtg agaga	ggag aaaa gagg aaac	agc aaa gtt cat	cagc caaa gaga aatg	cca caa tgt tct	caga aaaa agag ttaa	caaaaa tgatga ctgcct ttttta atcact aaa
	<21 <21	0 > 1 1 > 2 2 > P 3 > H	440 RT	sapi	ens											
	Met 1		Ser		5					10					15	Glu
				20	Pro				25					30		
	Arg	His	Gln	Gln	Glu	Phe	Ala	Val	Pro	Asp	Tyr	Arg	Ser		His	Leu

Glu Val Ser Gln Ala Ser Gln Leu Leu Gln Gln Gln Gln Gln Gln Gln Gln So50 60 Leu Arg Arg Arg Pro Ser Leu Leu Ser Glu Phe His Pro Gly Ser Asp

Arg Pro Gln Glu Arg Arg Thr Ser Tyr Glu Pro Phe His Pro Gly Pro Ser Pro Val Asp His Asp Ser Leu Glu Ser Lys Arg Pro Arg Leu Glu Gln Val Ser Asp Ser His Phe Gln Arg Val Ser Ala Ala Val Leu Pro Leu Val His Pro Leu Pro Glu Gly Leu Arg Ala Ser Ala Asp Ala Lys Lys Asp Pro Ala Phe Gly Gly Lys His Glu Ala Pro Ser Ser Pro Ile Ser Gly Gln Pro Cys Gly Asp Asp Gln Asn Ala Ser Pro Ser Lys Leu Ser Lys Glu Glu Leu Ile Gln Ser Met Asp Arg Val Asp Arg Glu Ile Ala Lys Val Glu Gln Gln Ile Leu Lys Leu Lys Lys Lys Gln Gln Gln Leu Glu Glu Glu Ala Ala Lys Pro Pro Glu Pro Glu Lys Pro Val Ser Pro Pro Pro Val Glu Gln Lys His Arg Ser Ile Val Gln Ile Ile Tyr Asp Glu Asn Arg Lys Lys Ala Glu Glu Ala His Lys Ile Phe Glu Gly 250 255 Leu Gly Pro Lys Val Glu Leu Pro Leu Tyr Asn Gln Pro Ser Asp Thr Lys Val Tyr His Glu Asn Ile Lys Thr Asn Gln Val Met Arg Lys Lys Leu Ile Leu Phe Phe Lys Arg Arg Asn His Ala Arg Lys Gln Arg Glu Gln Lys Ile Cys Gln Arg Tyr Asp Gln Leu Met Glu Ala Trp Glu Lys Lys Val Asp Arg Ile Glu Asn Asn Pro Arg Arg Lys Ala Lys Glu Ser Lys Thr Arg Glu Tyr Tyr Glu Lys Gln Phe Pro Glu Ile Arg Lys Gln Arg Glu Gln Glu Arg Phe Gln Arg Val Gly Gln Arg Gly Ala Gly Leu Ser Ala Thr Ile Ala Arg Ser Glu His Glu Ile Ser Glu Ile Ile Asp Gly Leu Ser Glu Gln Glu Asn Asn Glu Lys Gln Met Arg Gln Leu Ser Val Ile Pro Pro Met Met Phe Asp Ala Glu Gln Arg Arg Val Lys Phe Ile Asn Met Asn Gly Leu Met Glu Asp Pro Met Lys Val Tyr Lys Asp Arg Gln Phe Met Asn Val Trp Thr Asp His Glu Lys Glu Ile Phe Lys Asp Lys Phe Ile Gln His Pro Lys Asn Phe Gly Leu Ile Ala Ser Tyr Leu Glu Arg Lys Ser Val Pro Asp Cys Val Leu Tyr Tyr Tyr Leu Thr Lys Lys Asn Glu Asn Tyr Lys Ala Leu Val Arg Arg Asn Tyr Gly Lys Arg Arg Gly Arg Asn Gln Gln Ile Ala Arg Pro Ser Gln Glu Glu Lys Val Glu Glu Lys Glu Glu Asp Lys Ala Glu Lys Thr Glu Lys Lys Glu Glu Glu Lys Lys Asp Glu Glu Glu Lys Asp Glu Lys Glu Asp Ser Lys Glu Asn Thr Lys Glu Lys Asp Lys Ile Asp Gly Thr Ala Glu Glu

555 545 Thr Glu Glu Arg Glu Gln Ala Thr Pro Arg Gly Arg Lys Thr Ala Asn 570 565 Ser Gln Gly Arg Arg Lys Gly Arg Ile Thr Arg Ser Met Thr Asn Glu 580 585 Ala Ala Ala Ala Ser Ala Ala Ala Ala Ala Thr Glu Glu Pro Pro 600 Pro Pro Leu Pro Pro Pro Pro Glu Pro Ile Ser Thr Glu Pro Val Glu 615 Thr Ser Arg Trp Thr Glu Glu Glu Met Glu Val Ala Lys Lys Gly Leu 630 635 Val Glu His Gly Arg Asn Trp Ala Ala Ile Ala Lys Met Val Gly Thr 650 Lys Ser Glu Ala Gln Cys Lys Asn Phe Tyr Phe Asn Tyr Lys Arg Arg 665 His Asn Leu Asp Asn Leu Leu Gln Gln His Lys Gln Lys Thr Ser Arg 685 680 Lys Pro Arg Glu Glu Arg Asp Val Ser Gln Cys Glu Ser Val Ala Ser 695 700 Thr Val Ser Ala Gln Glu Asp Glu Asp Ile Glu Ala Ser Asn Glu Glu 715 710 Glu Asn Pro Glu Asp Ser Glu Val Glu Ala Val Lys Pro Ser Glu Asp 730 Ser Pro Glu Asn Ala Thr Ser Arg Gly Asn Thr Glu Pro Ala Val Glu 745 750 740 Leu Glu Pro Thr Thr Glu Thr Ala Pro Ser Thr Ser Pro Ser Leu Ala 760 765 755 Val Pro Ser Thr Lys Pro Ala Glu Asp Glu Ser Val Glu Thr Gln Val 775 780 Asn Asp Ser Ile Ser Ala Glu Thr Ala Glu Gln Met Asp Val Asp Gln 790 795 Gln Glu His Ser Ala Glu Glu Gly Ser Val Cys Asp Pro Pro Pro Ala 805 810 Thr Lys Ala Asp Ser Val Asp Val Glu Val Arg Val Pro Glu Asn His 825 Ala Ser Lys Val Glu Gly Asp Asn Thr Lys Glu Arg Asp Leu Asp Arg 840 Ala Ser Glu Lys Val Glu Pro Arg Asp Glu Asp Leu Val Val Ala Gln 855 Gln Ile Asn Ala Gln Arg Pro Glu Pro Gln Ser Asp Asn Asp Ser Ser 870 875 Ala Thr Cys Ser Ala Asp Glu Asp Val Asp Gly Glu Pro Glu Arg Gln 890 885 Arg Met Phe Pro Met Asp Ser Lys Pro Ser Leu Leu Asn Pro Thr Gly 905 900 Ser Ile Leu Val Ser Ser Pro Leu Lys Pro Asn Pro Leu Asp Leu Pro 920 Gln Leu Gln His Arg Ala Ala Val Ile Pro Pro Met Val Ser Cys Thr 940 935 Pro Cys Asn Ile Pro Ile Gly Thr Pro Val Ser Gly Tyr Ala Leu Tyr 955 950 Gln Arg His Ile Lys Ala Met His Glu Ser Ala Leu Leu Glu Glu Gln 970 965 Arg Gln Arg Gln Glu Gln Ile Asp Leu Glu Cys Arg Ser Ser Thr Ser 985 Pro Cys Gly Thr Ser Lys Ser Pro Asn Arg Glu Trp Glu Val Leu Gln 1000 995 Pro Ala Pro His Gln Leu Ile Thr Asn Leu Pro Glu Gly Val Arg Leu 1020 1015 Pro Thr Thr Arg Pro Thr Arg Pro Pro Pro Leu Ile Pro Ser Ser

1030 1035 1025 Lys Thr Thr Val Ala Ser Glu Lys Pro Ser Phe Ile Met Gly Gly Ser 1045 1050 1055 Ile Ser Gln Gly Thr Pro Gly Thr Tyr Leu Thr Ser His Asn Gln Ala 1060 1065 1070 Ser Tyr Thr Gln Glu Thr Pro Lys Pro Ser Val Gly Ser Ile Ser Leu 1075 1080 1085 Gly Leu Pro Arg Gln Gln Glu Ser Ala Lys Ser Ala Thr Leu Pro Tyr 1090 1095 1100 Ile Lys Gln Glu Glu Phe Ser Pro Arg Ser Gln Asn Ser Gln Pro Glu 1105 1110 1115 1120 Gly Leu Leu Val Arg Ala Gln His Glu Gly Val Val Arg Gly Thr Ala 1125 1130 1135 Gly Ala Ile Gln Glu Gly Ser Ile Thr Arg Gly Thr Pro Thr Ser Lys 1140 1145 1150 Ile Ser Val Glu Ser Ile Pro Ser Leu Arg Gly Ser Ile Thr Gln Gly 1155 1160 1165 Thr Pro Ala Leu Pro Gln Thr Gly Ile Pro Thr Glu Ala Leu Val Lys 1170 1175 1180 Gly Ser Ile Ser Arg Met Pro Ile Glu Asp Ser Ser Pro Glu Lys Gly 1185 1190 1195 1200 Arg Glu Glu Ala Ala Ser Lys Gly His Val Ile Tyr Glu Gly Lys Ser 1205 1210 1215 Gly His Ile Leu Ser Tyr Asp Asn Ile Lys Asn Ala Arg Glu Gly Thr 1220 1225 1230 Arg Ser Pro Arg Thr Ala His Glu Ile Ser Leu Lys Arg Ser Tyr Glu 1235 1240 1245 Ser Val Glu Gly Asn Ile Lys Gln Gly Met Ser Met Arg Glu Ser Pro 1250 1255 1260 Val Ser Ala Pro Leu Glu Gly Leu Ile Cys Arg Ala Leu Pro Arg Gly 1265 1270 1275 1280 Ser Pro His Ser Asp Leu Lys Glu Arg Thr Val Leu Ser Gly Ser Ile 1285 1290 1295 Met Gln Gly Thr Pro Arg Ala Thr Thr Glu Ser Phe Glu Asp Gly Leu 1300 1305 1310 Lys Tyr Pro Lys Gln Ile Lys Arg Glu Ser Pro Pro Ile Arg Ala Phe 1315 1320 1325 Glu Gly Ala Ile Thr Lys Gly Lys Pro Tyr Asp Gly Ile Thr Thr Ile 1330 1335 1340 Lys Glu Met Gly Arg Ser Ile His Glu Ile Pro Arg Gln Asp Ile Leu 1345 1350 1355 1360 Thr Gln Glu Ser Arg Lys Thr Pro Glu Val Val Gln Ser Thr Arg Pro 1365 1370 1375 Ile Ile Glu Gly Ser Ile Ser Gln Gly Thr Pro Ile Lys Phe Asp Asn 1380 1385 1390 Asn Ser Gly Gln Ser Ala Ile Lys His Asn Val Lys Ser Leu Ile Thr 1395 1400 1405 Gly Pro Ser Lys Leu Ser Arg Gly Met Pro Pro Leu Glu Ile Val Pro 1415 1420 1410 Glu Asn Ile Lys Val Val Glu Arg Gly Lys Tyr Glu Asp Val Lys Ala 1430 1435 Gly Glu Thr Val Arg Ser Arg His Thr Ser Val Val Ser Ser Gly Pro 1445 1450 Ser Val Leu Arg Ser Thr Leu His Glu Ala Pro Lys Ala Gln Leu Ser 1460 1465 1470 Pro Gly Ile Tyr Asp Asp Thr Ser Ala Arg Arg Thr Pro Val Ser Tyr 1475 1480 1485 Gln Asn Thr Met Ser Arg Gly Ser Pro Met Met Asn Arg Thr Ser Asp 1490 1495 1500

Val Thr Ile Pro Pro Asn Lys Ser Thr Asn His Glu Arg Lys Ser Thr

1505 1510 1515 152 Leu Thr Pro Thr Gln Arg Glu Ser Ile Pro Ala Lys Ser Pro Val Pro 1525 Gly Val Asp Pro Val Val Ser His Ser Pro Phe Asp Pro His His Arg

1540 1545 1550 Gly Ser Thr Ala Gly Glu Val Tyr Trp Ser His Leu Pro Thr Gln Leu

1555 1560 1565 Asp Pro Ala Met Pro Phe His Arg Ala Leu Asp Pro Ala Ala Ala Ala 1570 1580

Tyr Leu Phe Gln Arg Gln Leu Ser Pro Thr Pro Gly Tyr Pro Ser Gln 1595 1595 1600

Tyr Gln Leu Tyr Ala Met Glu Asn Thr Arg Gln Thr Ile Leu Asn Asp 1605 1610 1615

Tyr Ile Thr Ser Gln Gln Met Gln Val Asn Leu Arg Pro Asp Val Ala 1620 1625 1630 Arg Gly Leu Ser Pro Arg Glu Gln Pro Leu Gly Leu Pro Tyr Pro Ala

Arg Gly Leu Ser Pro Arg Glu Gin Pro Leu Gly Leu Pro Tyr Pro Ala 1645 Thr Arg Gly Ile Ile Asp Leu Thr Asn Met Pro Pro Thr Ile Leu Val

1650 1650 1660
Pro His Pro Gly Gly Thr Ser Thr Pro Pro Met Asp Arg Ile Thr Tyr
1665 1670 1675 1680

Ile Pro Gly Thr Gln Ile Thr Phe Pro Pro Arg Pro Tyr Asn Ser Ala 1685 Ser Met Ser Pro Gly His Pro Thr His Leu Ala Ala Ala Ala Ser Ala

ser met ser pro Gly His pro Thr His Leu Ala Ala Ala Ala Ser Ala 1700 1705 1710 Glu Arg Glu Arg Glu Arg Glu Lys Glu Arg Glu Arg Glu Arg

1715 1720 1725

The Ala Ala Ala Ser Ser Asp Leu Tyr Leu Arg Pro Gly Ser Glu Gln
1730 1735 1740

Pro Gly Arg Pro Gly Ser His Gly Tyr Val Arg Ser Pro Ser Pro Ser 1745 Val Arg Thr Gln Glu Thr Met Leu Gln Gln Arg Pro Ser Val Phe Gln

Val Arg Thr Gin Giu Thr Met Leu Gin Arg Flo Ser Val Flee Gin 1765 1770 1775 Gly Thr Asn Gly Thr Ser Val Ile Thr Pro Leu Asp Pro Thr Ala Gln

1780 1785 1790 Leu Arg Ile Met Pro Leu Pro Ala Gly Gly Pro Ser Ile Ser Gln Gly 1795 1800 1805

Leu Pro Ala Ser Arg Tyr Asn Thr Ala Ala Asp Ala Leu Ala Ala Leu 1815 1810 1820 Val Asp Ala Ala Ala Ser Ala Pro Gln Met Asp Val Ser Lys Thr Lys

1825 1830 1835 1840 Glu Ser Lys His Glu Ala Ala Arg Leu Glu Glu An Leu Arg Ser Arg 1845 1850 1855

Ser Ala Ala Val Ser Glu Gln Gln Gln Leu Glu Gln Lys Thr Leu Glu 1860 1865 1870

Val Glu Lys Arg Ser Val Gln Cys Leu Tyr Thr Ser Ser Ala Phe Pro 1875 Ser Gly Lys Pro Gln Pro His Ser Ser Val Val Tyr Ser Glu Ala Gly

Ser Gly Lys Pro Gin Pro His Ser Ser Val Val Tyr Ser Giu Ala Gly 1890 1890 1900 Lys Asp Lys Gly Pro Pro Pro Lys Ser Arg Tyr Glu Glu Glu Leu Arg

1905 1910 1915 1920 Thr Arg Gly Lys Thr Thr Ile Thr Ala Ala Aen Phe Ile Asp Val Ile 1925 1930 1935

Ile Thr Arg Gln Ile Ala Ser Asp Lys Asp Ala Arg Glu Arg Gly Ser 1940 1950 1950 Gln Ser Ser Asp Ser Ser Ser Ser Leu Ser Ser His Arg Tyr Glu Thr

1955 1960 1965 Pro Ser Asp Ala Ile Glu Val Ile Ser Pro Ala Ser Ser Pro Ala Pro

Pro Ser Asp Ala Ile Glu Val Ile Ser Pro Ala Ser Ser Pro Ala Pro 1970 1980 Pro Gln Glu Lys Leu Gln Thr Tyr Gln Pro Glu Val Val Lys Ala Asn

1990 1985 1995 Gln Ala Glu Asn Asp Pro Thr Arg Gln Tyr Glu Gly Pro Leu His His 2005 2010 2015 Tyr Arg Pro Gln Gln Glu Ser Pro Ser Pro Gln Gln Leu Pro Pro 2020 2025 2030 Ser Ser Gln Ala Glu Gly Met Gly Gln Val Pro Arg Thr His Arg Leu 2035 2040 2045 Ile Thr Leu Ala Asp His Ile Cys Gln Ile Ile Thr Gln Asp Phe Ala 2050 2055 2060 Arg Asn Gln Val Ser Ser Gln Thr Pro Gln Gln Pro Pro Thr Ser Thr 2065 2070 2075 2080 Phe Gln Asn Ser Pro Ser Ala Leu Val Ser Thr Pro Val Arg Thr Lvs 2085 2090 Thr Ser Asn Arg Tyr Ser Pro Glu Ser Gln Ala Gln Ser Val His His 2100 2105 2110 Gln Arg Pro Gly Ser Arg Val Ser Pro Glu Asn Leu Val Asp Lys Ser 2115 2120 2125 Arg Gly Ser Arg Pro Gly Lys Ser Pro Glu Arg Ser His Val Ser Ser 2130 2135 2140 Glu Pro Tyr Glu Pro Ile Ser Pro Pro Gln Val Pro Val Val His Glu 2145 2150 2155 Lys Gln Asp Ser Leu Leu Leu Ser Gln Arg Gly Ala Glu Pro Ala 2165 2170 2175 Glu Gln Arg Asn Asp Ala Arg Ser Pro Gly Ser Ile Ser Tyr Leu Pro 2180 2185 2190 Ser Phe Phe Thr Lys Leu Glu Asn Thr Ser Pro Met Val Lys Ser Lys 2195 2200 2205 Lys Gln Glu Ile Phe Arg Lys Leu Asn Ser Ser Gly Gly Asp Ser 2210 2215 2220 Asp Met Ala Ala Ala Gln Pro Gly Thr Glu Ile Phe Asn Leu Pro Ala 2230 2235 Val Thr Thr Ser Gly Ser Val Ser Ser Arg Gly His Ser Phe Ala Asp 2245 2250 2255 Pro Ala Ser Asn Leu Gly Leu Glu Asp Ile Ile Arg Lys Ala Leu Met 2260 2265 2270 Gly Ser Phe Asp Asp Lys Val Glu Asp His Gly Val Val Met Ser Gln 2275 2280 2285 Pro Met Gly Val Val Pro Gly Thr Ala Asn Thr Ser Val Val Thr Ser 2290 2295 2300 Gly Glu Thr Arg Arg Glu Glu Gly Asp Pro Ser Pro His Ser Gly Gly 2310 2315 2320 Val Cys Lys Pro Lys Leu Ile Ser Lys Ser Asn Ser Arg Lys Ser Lys 2325 2330 2335 Ser Pro Ile Pro Gly Gln Gly Tyr Leu Gly Thr Glu Arg Pro Ser Ser 2345 2350 Val Ser Ser Val His Ser Glu Gly Asp Tyr His Arg Gln Thr Pro Gly 2355 2360 2365 Trp Ala Trp Glu Asp Arg Pro Ser Ser Thr Gly Ser Thr Gln Phe Pro 2370 2375 2380 Tyr Asn Pro Leu Thr Met Arg Met Leu Ser Ser Thr Pro Pro Thr Pro 2390 2395 2400 Ile Ala Cys Ala Pro Ser Ala Val Asn Gln Ala Ala Pro His Gln Gln 2405 2410 2415 Asn Arg Ile Trp Glu Arg Glu Pro Ala Pro Leu Leu Ser Ala Gln Tyr 2420 2425

Glu Thr Leu Ser Asp Ser Asp Asp 2435 2440